

Research paper

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RNA interference is ineffective as a routine method for gene silencing in chick embryos as monitored by *fgf8* silencing

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Abstract

The *in vivo* accessibility of the chick embryo makes it a favoured model system for experimental developmental biology. Although the range of available techniques now extends to miss-expression of genes through *in ovo* electroporation, it remains difficult to knock out individual gene expression. Recently, the possibility of silencing gene expression by RNAi in chick embryos has been reported. However, published studies show only discrete quantitative differences in the expression of the endogenous targeted genes and unclear morphological alterations. To elucidate whether the tools currently available are adequate to silence gene expression sufficiently to produce a clear and specific null-like mutant phenotype, we have performed several experiments with different molecules that trigger RNAi: dsRNA, siRNA, and shRNA produced from a plasmid coexpressing green fluorescent protein as an internal marker. Focussing on *fgf8* expression in the developing isthmus, we show that no morphological defects are observed, and that *fgf8* expression is neither silenced in embryos microinjected with dsRNA nor in embryos microinjected and electroporated with a pool of siRNAs. Moreover, *fgf8* expression was not significantly silenced in most isthmus cells transformed with a plasmid producing engineered shRNAs to *fgf8*. We also show that siRNA molecules do not spread significantly from cell to cell as reported for invertebrates, suggesting the existence of molecular differences between different model systems that may explain the different responses to RNAi. Although our results are basically in agreement with previously reported studies, we suggest, in contrast to them, that with currently available tools and techniques the number of cells in which *fgf8* gene expression is decreased, if any, is not sufficient to generate a detectable mutant phenotype, thus making RNAi useless as a routine method for functional gene analysis in chick embryos.

Key words

RNA interference (RNAi), small interfering RNA (siRNA), short hairpin RNA (shRNA), chick embryo, isthmus, *fgf8*

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1. Introduction

The *in ovo* accessibility of the chick embryo makes it a favoured model system for *in vivo* studies of developmental processes. Since the development of *in ovo* gene transfer by electroporation [1], it has been possible to analyse the function of a gene by overexpressing it in specific tissues. However, it is still difficult to knock out the function of a gene of interest because it is not yet possible to make mutant animals by gene targeting. Recently however, the possibility of silencing gene expression by RNA interference (RNAi) has been reported.

RNAi is thought to be an evolutionarily conserved process that responds to the presence of double-stranded RNA (dsRNA) [2, 3, 4, 5]. The mechanism through which RNAi functions is based on the action of a set of enzymes that cleave the dsRNA into small interfering 21-23mer RNAs (siRNA) that act as templates for the specific degradation of the corresponding mRNAs [3, 6, 7, 8]. The key enzyme in this process is Dicer, a Ribonuclease III family molecule containing a PAZ domain and two dsRNA binding domains [9].

RNAi has been successfully used in several invertebrates, e.g. in *C. elegans* and *Drosophila*, both through dsRNA and constructs expressing siRNA [10, 11, 12, 13, 14]. It was initially postulated that this technique could not be used in vertebrates because the intracellular presence of dsRNA may trigger an antiviral response mediated by a dsRNA-dependent protein kinase (PKR) and the interferon system, which inhibits transcription [15]. However, the discovery of siRNAs, which are thought to escape the PKR, raised the possibility of using RNAi in vertebrates, although there are some reports suggesting that in mammalian cells siRNAs may actually upregulate some interferon response genes [16, 17].

Several recent papers have suggested that RNAi is effective in zebrafish, mouse and chick embryos. In zebrafish, Wargelius [18] and Li [19] reported the application of RNAi by injecting dsRNAs at 1-2 cell stage. However, Oates [20], using dsRNAs from several genes, were not able to distinguish the phenotypes produced by them, suggesting a non-specific effect of dsRNA on gene silencing. In mice, the use of RNAi has been reported in cultured cell lines [21, 22, 23], in oocytes [24, 25], and in embryos [23, 26, 27]. In these studies, the expression of the gene to be silenced was only partially reduced in a population of cells, and no morphological alterations were reported, with the exception of a mild mutant phenotype observed in the derivatives of some oocytes when compared with corresponding null mutants [23].

Finally, although the few published works on RNAi in chick embryos are very optimistic, they show discrete quantitative differences in the expression of endogenous target genes and limited, if any, morphological alterations [28, 29, 30, 31]. It is important to note that in all of these studies the transfection efficiency was assessed by using independent reporter plasmids as markers, thereby precluding the identification of the individual cells containing the molecules that trigger RNAi.

We have performed several experiments to elucidate whether the currently available tools are adequate to silence gene expression in chick embryos to a sufficient extent to produce a clear and specific null-like mutant phenotype using internal markers for siRNA and shRNA localisation. These experiments focused on RNAi of *fgf8* in the isthmus. *Fgf8* is a member of the fibroblast growth factor gene family, which codes for ligands that act to promote the growth and differentiation of many mesodermal and ectodermal cells by binding to specific receptors. Its expression is spatially regulated during development, e.g. in the isthmus organizer [32, 33]. The isthmus organizer, located at the mid-hindbrain boundary, exhibits a morphogenetic activity that acts on the midbrain neuroectoderm [34, 35]. Both *fgf8* hypomorph homozygote mutant mice and zebrafish acerebellar mutants, which carry a mutation affecting *fgf8*, lack posterior midbrain and cerebellar tissue [36, 37, 38].

We microinjected different potential triggers for RNAi into the neural tube: *fgf8* dsRNA; a pool of siRNAs to *fgf8* obtained *in vitro* by Dicer-digestion, some of which were labelled with digoxigenin (DIG) to monitor their uptake at a cellular level; and the psiRNA-hH1GFPzeo G2 plasmid engineered to express short hairpin RNAs (shRNAs) to *fgf8*. The shRNA-producing vector also expresses green fluorescent protein (GFP) as an internal transfection marker. Although our results are basically in agreement with previously reported studies on the silencing of endogenous genes in chick embryos, i.e. some cells expressing shRNA to *fgf8* may show a reduction in *fgf8* expression, we suggest that the currently available tools and techniques are inadequate to generate a detectable mutant *fgf8* phenotype at the embryo level, thus making RNAi useless as a routine method to study gene function in chick embryos.

2. Materials and methods

Production of dsRNA and *in vitro* Dicer-digestion

Fgf8 dsRNA was produced from a fragment of cDNA encoding *fgf8b* cloned in pBluescript KS [39]. After linearization with *EcoRI* or *XhoI*, RNA synthesis was carried out *in vitro* from T7 and T3 promoters for antisense and sense RNAs respectively. For each direction, 2 µg DNA was mixed with a final concentration of 4mM rNTPs or DIG-rNTPs (Roche) to synthesise dsRNA or DIG-labelled dsRNA respectively, and with the appropriate concentration of T3 and T7 polymerases (Roche). After 2 h

transcription at 37°C for the T7 promoter and 25°C for T3 promoter (to produce equimolar concentrations), 1 µl of DNase I (Roche) was added and the mixture incubated for 15 min at 37°C. The reaction was stopped with 380 µl of STOP solution (1M NH₄OAc, 10mM EDTA, 0.2% SDS). The RNA was extracted first with phenol:chloroform 1:1 (v/v) and then chloroform. Equal amounts of sense and antisense RNA from the aqueous layer were mixed, heated to 70°C for 10 min and annealed at 37°C for 30 min. After precipitation in ethanol, the dsRNA was dissolved in 10 µl of DMEM (Gibco) supplemented with 0.3 µg/ml Vital Red/DMEM to visualize microinjection, or in 10 µl of H₂O for *in vitro* Dicer-digestion. For quality control, 1µl of each sample was analyzed by gel electrophoresis (0.8% agarose).

To obtain a pool of siRNAs or DIG-labelled siRNAs from *fgf8* dsRNA, dsRNA or DIG-labelled dsRNA dissolved in H₂O were digested and purified using the Dicer Kit (Gene Therapy Systems), according to the manufacturers instructions. For quality controls, samples were analyzed by gel electrophoresis (3% agarose). The efficiency of *in vitro* Dicer-digestion of DIG-dsRNAs was also tested on an agarose gel, showing no differences with respect to control siRNAs (Fig. 1).

Plasmids and shRNA constructs

To monitor siRNA electroporation efficiency we used a GFP expression vector (pEGFP-C2, Clontech; see below). The shRNAs used to interfere with *fgf8* expression were designed according to the general rules of the siRNA Wizard™ program (Invivogen; www.invivogen.com), and the interfering sequence was cloned into the psiRNA-hH1GFPzeo G2 vector (Invivogen). This vector coexpresses the cloned shRNA, under the control of the H1 promoter, with GFP, under the control of the hEF1/HTLV promoter and SV40 enhancer (Fig. 2A).

Two sets of 21-mer oligonucleotides were selected from the *fgf8b* open reading frame (ORF) sequence. The 21-mer sense and antisense RNA sequences were linked to a seven nucleotide spacer (TCAAGAG) as a loop, and nucleotides corresponding to *Hind*III and *Acc*65I restriction sites were added to the 5' and 3' end of the reverse oligonucleotides respectively. The engineered DNAs for the corresponding shRNAs are shown in Fig. 2B. They were annealed and cloned into the psiRNA-hH1GFPzeo G2 vector using the *Hind*III/*Acc*65I restriction sites. Following the instructions of the plasmid supplier, vectors were transformed into the *E.coli* strain GT116, in which *sbcC* and *sbcD* genes are deleted, to increase shRNA stability.

In ovo microinjection and electroporation

Fertilized eggs obtained from a local farm (Granja Gibert) were incubated at 38°C and windowed using Scotch tape. They were staged according to Hamburger and Hamilton [40]. DsRNA, the pool of siRNAs or the shRNA-producing plasmid were microinjected into the neural tube of embryos at HH 9-11 using a Drummond piston microinjector (32 nl at 3-5 µg/µl). Embryos were contrasted with 0.3 mg/ml Fast Green (Sigma) solution, and the pool of siRNAs and the plasmid producing shRNA were mixed with Fast Green at 3 mg/ml (3:1 v/v) prior to microinjection to monitor their localisation in the neural tube. Just after microinjection, the pool of siRNAs and the shRNA-producing plasmid were electroporated using a TSS20 Ovodyne electroporator (Intracel; 25V, 50ms pulse-width, 5 pulses with 30ms intervals between the pulses) and platinum electrodes (Nepagene; 5mm in length, 0.5mm wide) in 0.01x PBS as electroporation medium. After electroporation, the embryo was covered with DMEM (Gibco), the window was sealed with Scotch tape and the egg was incubated at 38°C. The pool of siRNAs was co-electroporated with pEGFP-C2 (U57606) to assess electroporation efficiency and localisation. As described previously [41, 42], transfection occurs on one side of the neural tube, the other side serving as a control.

In situ hybridisation, immunohistochemistry and TUNEL assay

Embryos were fixed overnight (ON) at 4°C in 4% paraformaldehyde in 1xPBS, dehydrated in a methanol series and stored at -20°C prior to use. *In situ* hybridisation was performed as described by Bueno et al. [43] for single probe development, with minor modifications (hybridisation temperature was 55°C, and the incubation time of the anti-DIG antibody was 3 hours at room temperature). The *Fgf8* construct used to produce the antisense riboprobe was from Crossley et al. [39]. The α-peptide construct for antisense riboprobe production was generated by PCR cloning of 237 nt of the α-peptide sequence into pBluescript KS (*Eco*RI/*Xho*I restriction sites). The primers used were the following: forward 5'ccggaattcatggaccctgttgctgctgcaa3' and reverse 5'ccgctcgagcacagtgtcagcctctgggag3'. Antisense riboprobe was transcribed with T7 polymerase after *Eco*RI digestion.

As *in situ* hybridization treatments destroy GFP fluorescence, immunohistochemistry with a polyclonal antibody to GFP (Molecular Probes) was performed to detect its presence after *fgf8 in situ* hybridisation. Briefly, hybridised embryos were post-fixed at 4°C for 2 hours with 4% paraformaldehyde in 1xPBS. For immunohistochemistry on sections, the embryos were dehydrated in an ethanol series, passed through xylene and embedded in paraffin. 10 µm sections were dewaxed in xylene and rehydrated in an ethanol series. For both immunohistochemistry on sections and on whole mount embryos, samples were blocked and incubated with the anti-GFP antibody, at 1/250 or 1/500

respectively, for 48 h at 4°C. After extensive washes, samples were incubated with a FITC-conjugated donkey anti-rabbit polyclonal antibody (Molecular Probes).

To detect the presence of DIG-labelled siRNAs, embryos were processed as for *in situ* hybridisation. After permeabilisation, they were blocked, incubated with an anti-DIG antibody (Roche) and developed with NBT/BCIP.

Apoptotic cells were detected by TUNEL assay on whole mount embryos using standard procedures. Briefly, embryos were fixed overnight at 4°C in 4% paraformaldehyde and permeabilised with PBT (PBS + 0.1% Triton X-100 + 0.1% Sodium Citrate). The 3'-OH ends of DNA were labelled for two hours at 37°C by addition of digoxigenin 11-UTPs using the enzyme TdT. Then, the samples were blocked, the presence of DIG was detected with an anti-DIG antibody coupled to alkaline phosphatase and they were developed with NBT/BCIP.

3. Results

dsRNA microinjection into the neural tube

First of all, to test whether dsRNA triggers specific gene silencing in chick embryos, as reported for invertebrates, *fgf8* dsRNA was microinjected into the neural tube of embryos at HH9-11. After 24 h or 48 h of *in vivo* (*in ovo*) embryonic development, i.e. at HH18-20 or HH23-24 respectively, embryos were killed and examined under the dissecting microscope to search for morphological defects, especially at the midbrain/hindbrain junction. In these experiments no morphological defects were observed, as compared with control embryos (not shown).

To examine whether *fgf8* expression was silenced in these embryos, whole mount *in situ* hybridization with an *fgf8* riboprobe was performed. *Fgf8* expression was detected in all neuroepithelial areas where it has previously been reported to be expressed, i.e. the isthmus and the frontal end of the telencephalon, as well as in the other embryonic sites, e.g. the eyes, branchial arches, the narrow pit and the apical ectodermal ridge of the limbs in HH23-24 embryos, as in control embryos (not shown). Finally, careful microscopic examination of serial sections from the hybridized embryos revealed no subtle morphological defects in the neuroectoderm (not shown).

Electroporation of siRNAs into the isthmus and RNAi spreading

To force the penetration of the putative triggering molecules for *fgf8* gene silencing, a pool of siRNAs to *fgf8* obtained *in vitro* by Dicer-digestion from the corresponding dsRNA was microinjected into the neural tube of embryos at HH9-11 and immediately electroporated into the isthmus cells. In these experiments the siRNAs were co-injected and co-electroporated with the pEGFP-C2 plasmid as an independent marker of electroporation efficiency. After 24 h of *in vivo* development the embryos were killed. Those embryos showing GFP fluorescence on the electroporated side of the isthmus were examined under the dissecting microscope to search for morphological defects. Again, no defects were observed when compared with control embryos (Fig. 3, and data not shown).

To examine whether *fgf8* was silenced on the electroporated side of the isthmus, whole mount *in situ* hybridization with an *fgf8* riboprobe was performed. *Fgf8* expression showed no significant differences between the two sides of the isthmus. Careful microscopic examination of serial sections from the hybridized embryos did not reveal any subtle morphological defect (not shown).

As the pEGFP-C2 plasmid has a different size and overall structure to the siRNAs (circular 4.7kb or linear 21mers respectively) that may affect the electroporation efficiency, we monitored whether the isthmus cells had actually been transfected with the siRNAs by using DIG-labelled siRNAs. The DIG-siRNAs were also co-injected and co-electroporated together with the pEGFP-C2 plasmid. After 24 h of *in vivo* embryonic development embryos showing GFP fluorescence on the electroporated side of the neural tube were immunostained with an antibody to DIG. Most of the GFP positive cells also showed the presence of siRNAs in their cytoplasm (Fig. 3). Interestingly, no significant spreading of the siRNAs from the transfected cells was observed. From these experiments we conclude that the electroporation profile used is appropriate for the transfection of the siRNAs, and thus that electroporation of a pool of siRNAs to *fgf8* is not able to silence *fgf8* expression in the isthmus.

Electroporation of a plasmid producing shRNAs

To test the possibility that the results obtained by electroporation of a pool of siRNAs were due to an insufficient number of siRNA molecules per cell to trigger the degradation of all existing and newly synthesised *fgf8* mRNA molecules, the psiRNA-hH1GFPzeo G2 shRNA-producing plasmid was microinjected into the neural tube and electroporated into the isthmus.

First of all we tested whether the H1 promoter for the shRNA was properly directing the expression of the downstream cloned sequences in chick embryos. Wild type plasmids, which carry the α -peptide sequence of the β -gal gene located under the control of the H1 promoter, were microinjected and electroporated into the isthmus. After 12 h or 24 h of *in vivo* development, embryos showing GFP fluorescence on the electroporated side of the isthmus were hybridized with a riboprobe to the α -peptide. These embryos exhibited a very high level of α -peptide expression (Fig. 4), suggesting that the H1 promoter also directs the expression of the shRNAs in the cells of the chick neural tube. In this way

the number of shRNA molecules present in each particular cell will be much higher than the number of siRNA molecules electroporated after Dicer-digestion of the corresponding dsRNA. Moreover, this experiment also proved that both promoters (i.e. H1 promoter for the shRNA and hEF1/HTLV promoter for GFP) were working simultaneously, with no interferences.

Two different shRNAs to *fgf8*, the best shRNAs to silence *fgf8* expression according to the general rules of the siRNA Wizard™ program (Invivogen), were engineered and independently cloned into the psiRNA-hH1GFPzeo G2 plasmid. These constructs were either independently or simultaneously microinjected and electroporated into the isthmus of chick embryos at HH9-11, in order to maximize the possible gene silencing effect. After 24 h of *in vivo* development, embryos showing GFP fluorescence on the electroporated side of the isthmus were examined under the dissecting microscope to search for morphological defects. Once again, no morphological defects were observed, as compared with control embryos electroporated with the wild type plasmid (Figs. 5 and 6). To examine whether *fgf8* expression was silenced on the electroporated side of the isthmus, whole mount *in situ* hybridization with an *fgf8* riboprobe was performed. *Fgf8* expression examined at whole embryo level showed a slight decrease on the electroporated side of the isthmus as compared with the control side (Figs. 5 and 6), although careful microscopic examination of serial sections from the hybridized embryos did not reveal any subtle morphological defect (Figs. 5 and 6).

To check whether *fgf8* expression was actually silenced in the transformed cells, immunohistochemistry with an antibody to GFP was performed on whole mount embryos and on sections after *fgf8 in situ* hybridization. This experiment allowed us to visualize individual transfected cells and *fgf8* expressing cells simultaneously. Although some GFP positive cells were clearly not expressing *fgf8*, most of the transfected cells did, suggesting that if RNAi occurs in chick embryos, its efficiency is very low (Fig. 7).

Finally, as it has been reported that dsRNA and siRNA molecules may trigger an apoptotic response in vertebrate cells that may mask and/or simulate RNAi effects, we monitored apoptosis to check whether the slight decrease in *fgf8* expression was caused by the death of some of the transfected cells. The pattern of apoptotic cells in adjacent non-transfected structures of the electroporated side as the eye and the otic vesicle were used as controls. Embryos electroporated with the shRNA-producing plasmid showed few apoptotic cells that affect mostly the ectoderm, and no significant differences were observed between these embryos and either control embryos electroporated with the wild type plasmid or non-electroporated embryos (Fig. 8).

4. Discussion

We have analysed the possible use of RNAi as a routine method for specific gene silencing in chick embryos by assessing the capacity of different potential triggers of RNAi (dsRNA, a pool of siRNAs obtained *in vitro* by Dicer-digestion of the corresponding dsRNA, and a plasmid producing shRNA to the gene to be silenced) to silence *fgf8* expression in the isthmus, as well as to provoke morphological malformations in midbrain/hindbrain junction derivatives.

Action of RNAi

All published studies focused on the possible use of RNAi to silence gene expression in chick embryos, in which dsRNA molecules or non-viral vectors producing shRNAs have been electroporated into target cells, have taken advantage of the sealed cavity of the neural tube to introduce the putative trigger molecules prior to electroporation [28, 29, 31]. To facilitate the experimental manipulation of the embryos we also introduced the putative trigger molecules for RNAi into the neural tube. We have focused on *fgf8* expression in the isthmus to monitor gene silencing for several practical reasons. Firstly, the isthmus is easily accessible from the neural cavity, and clearly recognizable under a dissecting microscope to properly localize the molecules that trigger RNAi. Secondly, when the neural tube is completely sealed, at HH9, *fgf8* expression in the isthmus is still faint and is detected in relatively few cells that are restricted to a small area [32, 33], a fact that may facilitate the degradation of all existing mRNA molecules. And thirdly, the phenotype of *fgf8* null-mutant and allelic series of mutations affecting the isthmus has been reported in other vertebrate model systems, and is easily morphologically recognizable as a deletion of the isthmus derivatives [36].

We observed no morphological defects in any of the experiments performed. We have also shown that *fgf8* expression is neither silenced in embryos microinjected with dsRNA nor in embryos microinjected and electroporated with a pool of siRNAs. Moreover, *fgf8* expression is also not significantly silenced in most isthmus cells transformed with the psiRNA-hH1GFPzeo G2 plasmid producing the engineered shRNAs to *fgf8*, and an increase in cell death is not detected either in the transfected neuroectoderm or in adjacent structures serving as controls for the possible toxic effects of the electric field.

Our results are basically in agreement with previously reported studies on RNAi of endogenous genes in chick embryos. Thus, the electroporation of a pSilencer 1.0-U6 vector (Ambion) producing shRNA to *en-1* or *en-2* [29] or to *Islet1* [31] does not produce any recognizable morphological defect and only a discrete reduction of gene expression. In apparent contrast, electroporation of dsRNA to *axonin-1*

and *NrCAM*, which are involved in axonal guidance, produces some morphological defects affecting this process in 20% of the neurons [28], although the possibility that this arises from an increase in cell death due to the presence of dsRNA molecules was not analysed. Nevertheless, the presence of just 20% of affected neurons is unlikely to be sufficient to see any long-term defect, as in many cases embryos are able to balance mild developmental defects.

In a similar way, microinjection of the viral pSuper vector (OligoEngine) engineered to produce shRNA to silence *mkp-3* expression in the limb bud mesenchyme [30] caused an increase in cell death compared with controls microinjected with an RCAS virus expressing GFP, although unfortunately no other shRNAs were used to assess the non-specific effect of these molecules on cell death in the limb mesenchyme.

In these studies the electroporation efficiency and the localisation of the molecules that may trigger RNAi were assessed by co-electroporation of an independent reporter plasmid, thereby precluding analysis of which particular cells were effectively transfected with the RNAi triggering molecules. This is especially important when considering that a maximum of 60% of the cells of the electroporated side are actually transfected with the reporter plasmid [28]. This handicap is avoided with the psiRNA-hH1GFPzeo G2 plasmid used in this study, as it coexpresses GFP as a marker. This allowed us to clearly determine that although some transfected cells do not express *fgf8*, most of them do. This raises the question of whether the reduction of *fgf8* expression in some transfected cells occurs irrespective of the presence of the specific shRNA, or instead whether the presence of shRNA is triggering gene silencing only in certain cells due to an unknown intrinsic difference between them. Nevertheless, the small reduction in gene expression, if any, accounts for the lack of morphological defects in RNAi treated embryos.

A similar situation occurs in mice [21, 22, 23, 24, 25, 27]. Here, with the exception of a mild phenotype reported by Paddison et al. [23] in the derivatives of some oocytes, no phenotypic alterations have been reported.

Better results are obtained when attempting to silence the expression of exogenous genes in chick embryos, although the gene silencing is incomplete, Pekarik [28] reported that co-electroporation of a vector expressing yellow fluorescent protein (YFP) together with dsRNA to YFP in the neural tube causes a reduction of YFP fluorescence in a substantial number of cells. Moreover, Chesnutt and Niswander [31] also reported a significant but incomplete reduction of GFP expression when co-electroporating a vector expressing GFP and a pSilencer 1.0-U6 vector expressing shRNA to this gene.

Taking into account our results and the published works discussed above, we suggest that the currently available tools and techniques for RNAi are not suitable as a routine method for functional gene analysis in chick embryos, as it is in some invertebrates [11, 12, 13, 14]. This suggestion does not preclude a possible use of RNAi to analyze the effect of gene silencing at a molecular level in individual transfected cells, the ones that may show a reduction in gene expression.

Is RNAi an evolutionarily conserved mechanism?

Although it is commonly said that RNAi is an evolutionarily conserved mechanism, reports of molecular differences between several model systems are appearing. Thus, most invertebrates in which RNAi has been successfully used are able to take up dsRNA molecules by themselves, the exception being *Drosophila*. The molecule responsible for dsRNA uptake and siRNA spreading is SID-1, a specific transmembrane protein [44, 45, 46]. *Drosophila* shows RNAi but lacks a *sid-1* ortholog, thus not showing the spreading characteristic [45, 47]. In this way, the amount of dsRNA needed for RNAi in this organism is 10,000 times higher than in controls expressing *sid-1* ectopically.

The presence of *sid* homologs has been reported in mice, although these organisms do not seem to be able to take up dsRNA by themselves, a similar situation occurring in chick embryos. In fact, all reports on the use of dsRNA or siRNA for RNAi in vertebrates have forced these molecules to enter the cells, either by calcium precipitation, electroporation or cytoplasmic microinjection. Moreover, our results have also shown that siRNA molecules do not spread from the transfected cells to the rest of the tissue or the embryo, as has been reported to occur in most invertebrates [2], suggesting the existence of molecular differences between different model systems that may explain the different responses to RNAi. To develop a general procedure for RNAi in chick embryos for routine functional gene analysis, if it is technically and molecularly possible, a better knowledge of the mechanisms leading to gene silencing in vertebrates is needed.

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Conflict of interest

The authors have declared that no conflict of interest exists.

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Figures

Figure 1. siRNA and DIG-siRNA obtained by *in vitro* Dicer-digestion. A 3% agarose gel electrophoresis of siRNA and DIG-labelled siRNA to *fgf8* is shown. Note that Dicer is able to generate a pool of siRNAs from both dsRNA and DIG-labelled dsRNA.

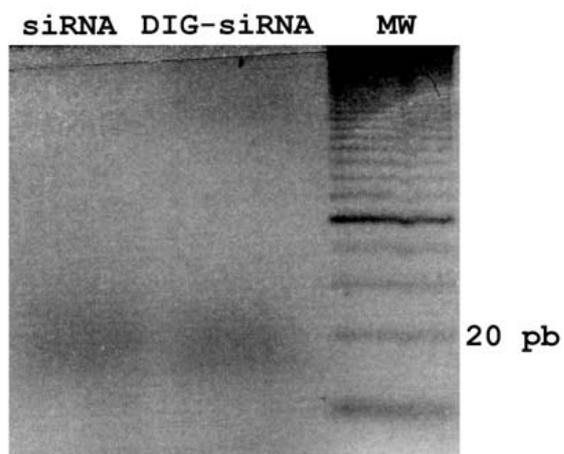
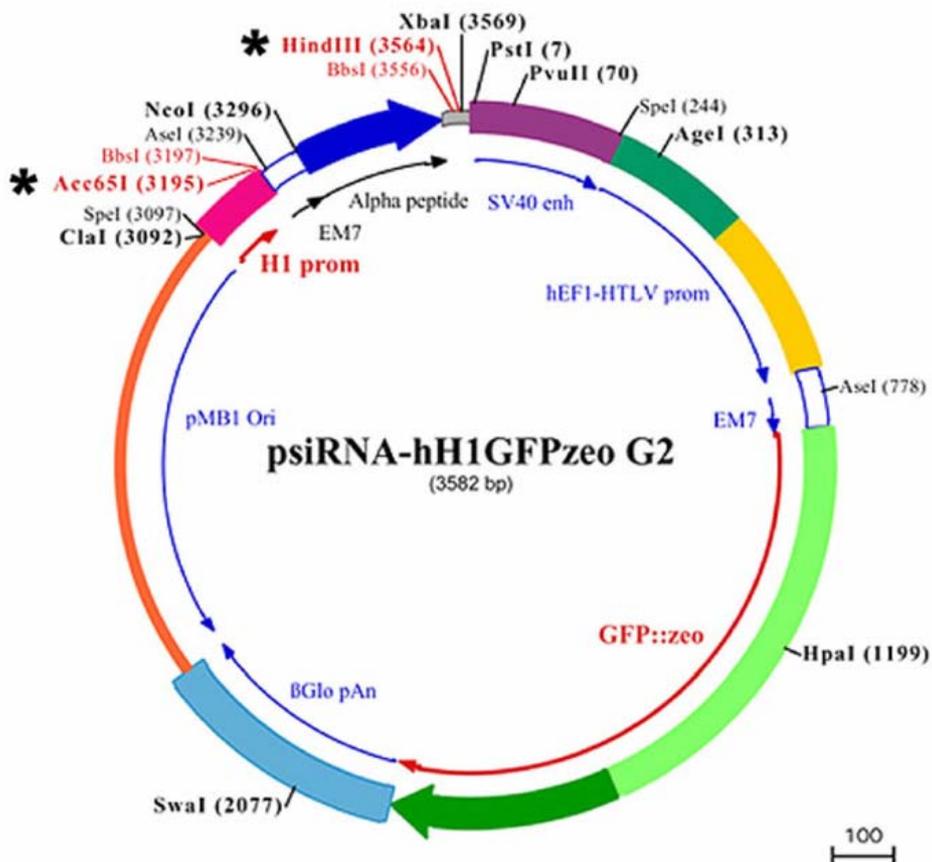


Figure 2. Plasmids and shRNA constructs. A) Wild-type psiRNA-hH1GFPzeo G2 vector map showing *Acc65I/HindIII* restriction sites (*), the α -peptide coding sequence located downstream of the H1 promoter, and the GFP coding sequence located downstream of hEF1-HTLV promoter. B) Engineered DNA oligonucleotides for the shRNAs: 21-mers are shown in green; the spacer is shown in blue; *HindIII* and *Acc65I* restriction sites are shown in orange; and the poly-A sequence is shown in black. The presumptive secondary structure of shfgf8b-1 is also shown.

A



B

shfgf8b-1 5' **GTACCTCACGTGCAGATCTTGGACAACATCAAGAGT**GTGTGTTGTTCCAAGATCTGCACGTTTTTTGGAAA 3'
 3' **GAGTGCACGCTAGAACCTGTTGTAGTTCTCACAACAGGTTCTAGACGTGCAAAAAACCTTTTCGA** 5'

shfgf8b-2 5' **GTACCTCAACGCGAGGTGCACCTTCATGATCAAGAGT**CATGAAGTGCACCTCGCGTTTTTTGGAAA 3'
 3' **GAGTTGCGCTCCACGTGAAGTACTAGTTCTCAGTACTTCACGTGGAGCGCAAAAAACCTTTTCGA** 5'

shfgf8b-1 presumptive secondary structure



Figure 3. Localisation of DIG-siRNA 24 h after electroporation. Embryos co-electroporated with a pool of DIG-labelled siRNAs and the pEGFP-C2 plasmid. A) GFP fluorescence into the electroporated side of the neural tube before DIG immunodetection (arrows). Yellow dashed line is delimiting the GFP expression area. B) GFP fluorescence into the electroporated side of the neural tube of the embryo shown in A) after DIG immunodetection (arrows). Note a decrease in GFP fluorescence due to the presence of the NBT/BCIP precipitate. C) Immunodetection of DIG-labelled siRNA in the neural tube (in blue; arrows) in the same embryo than A) and B). D) Merging of B) and C). Yellow dashed line has been extrapolated from A) and is delimiting the area of GFP expression before NBT/BCIP precipitation. Note that most cells exhibiting GFP fluorescence also show the presence of the DIG-siRNAs, although they do not colocalize in all electroporated cells. Also note the absence of siRNA spreading from the electroporated area. Abbreviations: Ist, isthmus; Oc, optic cup; Tel, telencephalon.

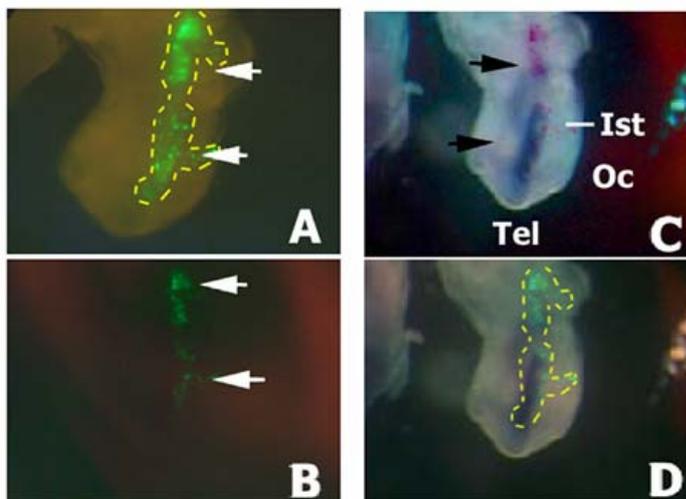


Figure 4. α -peptide expression in embryos transfected with the psiRNA-hH1GFPzeo G2 vector. A-B) Two different embryos 12 h after electroporation hybridised with the α -peptide antisense riboprobe. Note the expression of the α -peptide sequence in the electroporated half-side of the neural tube (arrows). C-D) Two different embryos 24 h after electroporation hybridised with the α -peptide antisense riboprobe. Note that the expression of the α -peptide is properly localized into the isthmus (arrows). E) Facial view of the embryo shown in D. Note that the α -peptide is expressed only in electroporated side (arrowhead).

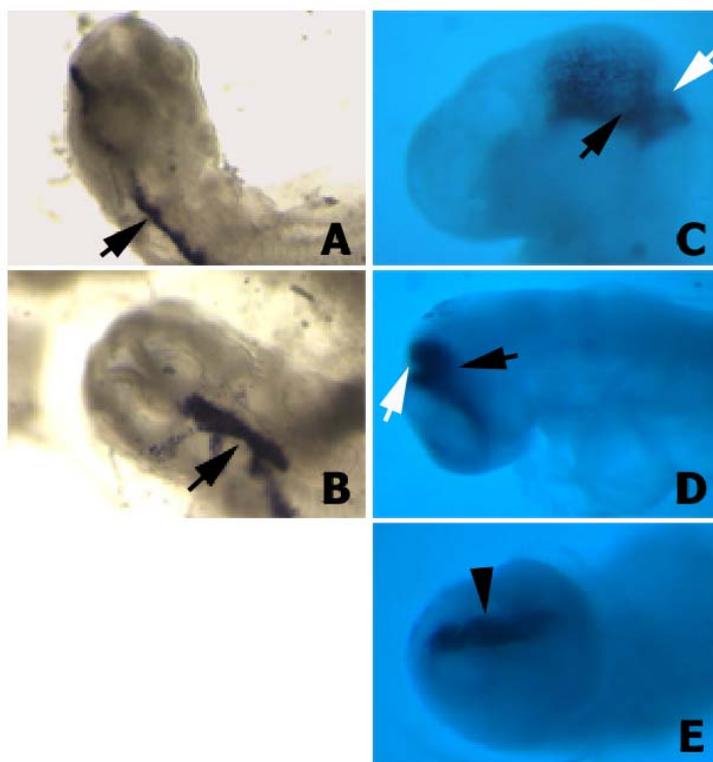


Figure 5. *Fgf8* and isthmus morphology in flattened isthmus of embryos transfected with *shfgf8b-1* and *shfgf8b-2*. Dorsal view of a flattened isthmus hybridised with the *fgf8* riboprobe 24 h after plasmids electroporation. The black line follows the dorsal midline. Note that no significant differences either in *fgf8* expression or neuroectoderm morphology are observed when comparing both sides of the isthmus.

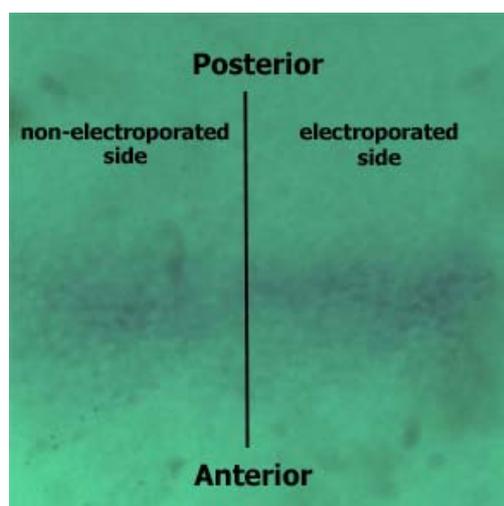


Figure 6. *Fgf8* expression and isthmus morphology in whole mount embryos transfected with *shfgf8b-1* and *shfgf8b-2*. Rostral is to the left. **A)** Embryo 24 h after electroporation, prior to *fgf8* *in situ* hybridisation. Note that the morphology of the isthmus derivatives is completely normal (compare with control embryo shown in **C**). **A')** Magnification of **A**) showing the isthmus area. **B)** GFP fluorescence (arrowheads) in the electroporated side of the same embryo shown in **A**). **B')** Magnification of **B**) showing the area of GFP expression (arrowheads). **C)** *Fgf8* *in situ* hybridisation of a control embryo. **D)** *Fgf8* *in situ* hybridisation in the isthmus of the embryo shown in **A**) (arrow). Note that *fgf8* expression in the isthmus is very similar to that shown in **C**). Abbreviations: AERfl, forelimb apical ectodermal ridge; AERhl, hindlimb apical ectodermal ridge; BA, branchial arches; Ist, isthmus; Met, metencephalon; Mes, mesencephalon; Tel, telencephalon.

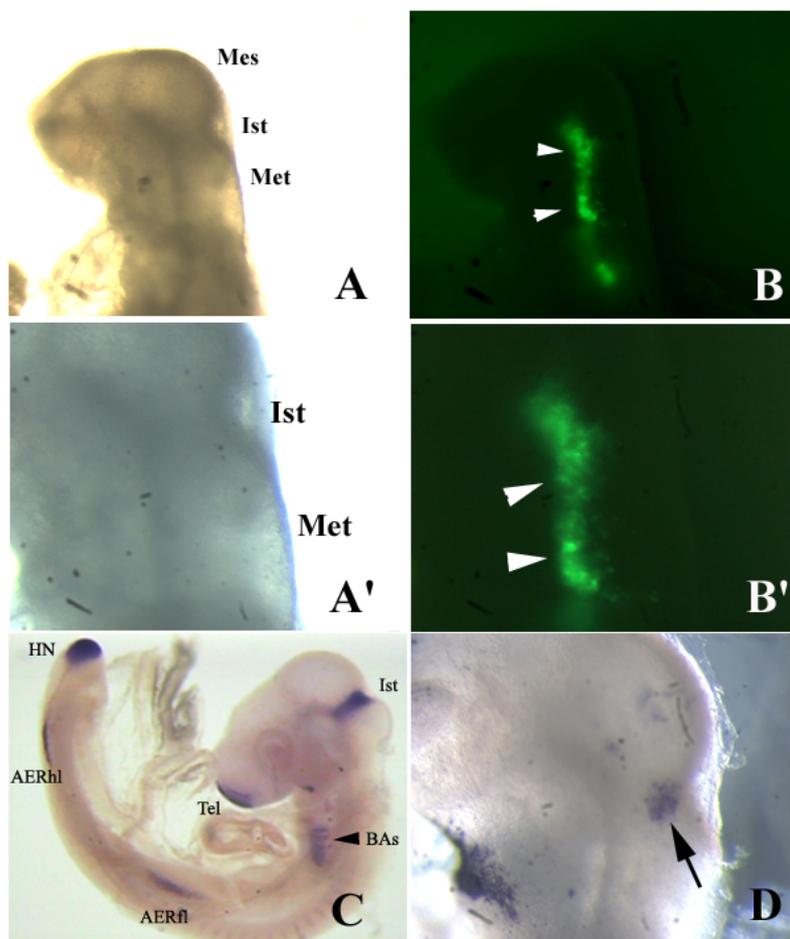


Figure 7. *Fgf8* expression and isthmus morphology in sections of embryos transfected with *shfgf8b-1* and *shfgf8b-2*. Transversal sections of the neural tube at the isthmus level. Both sides of neural tube are shown. The electroporated side is to the left. **A)** *Fgf8* expression detected by *in situ* hybridisation. **B)** GFP expression in the same section shown in **A**) detected by GFP immunostaining. Note that GFP is detected only in the electroporated side (arrowheads). **C)** Merging of **A**) and **B**). The expression of *fgf8* is very similar in electroporated and control neuroepithelium. **D to F)** Magnification of sections shown in **A** to **C** respectively. Note the presence of cells showing *fgf8* and GFP expression simultaneously (arrows and arrowhead). **G to I)** Magnification of sections shown in **A** to **C** respectively in a different focal plane. Note the presence of cells showing GFP expression but not *fgf8* expression (double arrowhead).

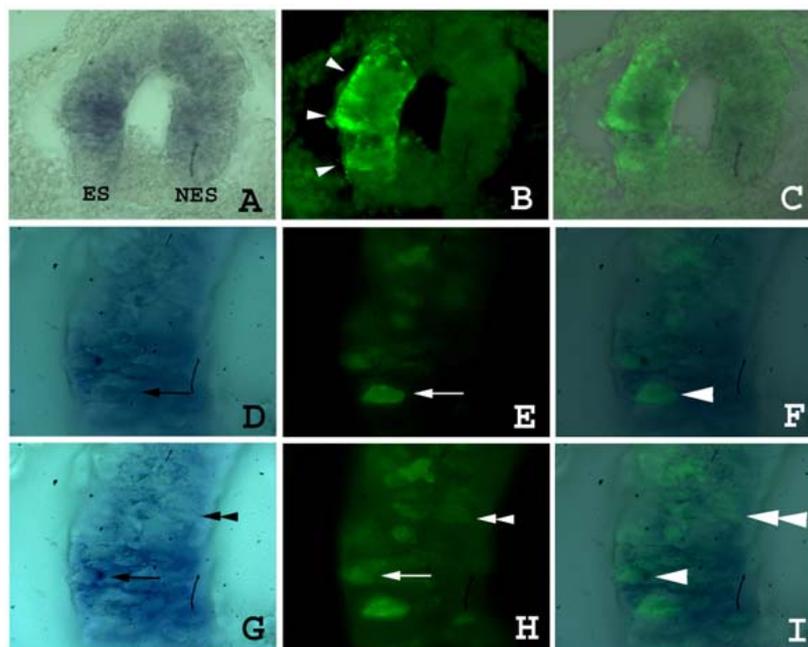


Figure 8. Apoptosis in embryos transfected with shfgf8b-1 and shfgf8b-2. **A)** Dorsal view of a transfected embryo in which apoptotic cells have been detected by TUNEL assay (brown nuclei). Note the absence of morphological defects (the transfected side is to the bottom). **B)** GFP fluorescence in the same embryo. The transfected area includes the isthmus. **C)** Merging of B) and C). **D)** Magnification of the isthmic area of A). Note that both sides of the embryo exhibit a similar amount of apoptotic cells (arrows), which affect mostly the ectoderm. **E)** Magnification of the isthmic area of B). The arrow points an apoptotic cell not expressing GFP. **F)** Magnification of the isthmic area of C). **G)** Dissected cephalic cavity showing the internal side of the mesencephalon, the metencephalon and the isthmus. Apoptotic cells have been detected by TUNEL assay (brown nuclei). The morphology of the transfected side (left hand side) is completely normal, and that apoptotic cells are equally distributed at both sides of the embryo. **H)** GFP fluorescence in the same embryo. The transfected area includes the isthmus. **I)** Merging of G) and H). **J)** Apoptosis control in the eye of the electroporated side. Note that apoptotic cells are located in the dorsal portion of the optic cup as described in the literature for normal embryos (Trousse et al., 2001). **K)** Apoptosis control in the ear of the electroporated side. Note that apoptotic cells are located in the dorsal portion of the otic vesicle as described in the literature for normal embryos (Lang et al., 2000). Abbreviations: Ant, anterior; Mes, mesencephalon; Met, metencephalon; Pos, posterior.

