

Research Paper

Overexpression of Selenoprotein H Reduces Ht22 Neuronal Cell Death after UVB Irradiation by Preventing Superoxide Formation

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Selenoproteins have been shown to exhibit a variety of biological functions, including antioxidant functions, maintaining cellular redox balance, and heavy metal detoxification. UV irradiation-induced damage is partially mediated by increased oxygen radical production. The present study is designed to examine the antioxidative effects of human selenoprotein H (hSelH) after brief period of UVB irradiation on the murine hippocampal neuronal cell line Ht22. Ht22 cells were stably transfected with the hSelH gene or with MSCV empty vector and exposed to UVB irradiation with or without the presence of serum. The results showed that cell viability was significantly higher in hSelH-transfected cells compared to the MSCV vector-transfected cells after 24 h of recovery with or without the presence of serum in the media. Further studies revealed that while the number of superoxide anion (O₂⁻) positive cells was increased following a 7 mJ/cm² of UVB irradiation and 5 h of recovery, overexpression of hSelH significantly reduced superoxide production. These results suggest that hSelH overexpression protects cells from UVB irradiation-induced cell death by reducing the O₂⁻ formation.

Key words: selenoprotein H; ultraviolet B irradiation; reactive oxygen species; superoxide anion; cell viability.

1. Introduction

Ultraviolet (UV) irradiation, especially UVB irradiation (290-320 nm), leads to premature skin aging, suppression of immunity, and skin cancer [1]. In neuronal cells, UV irradiation induces apoptosis; therefore, many research groups have used this model as a tool to study the intracellular mechanisms of apoptosis [2-7]. UV irradiation induces cell death via a variety of cellular mechanisms. These include induction of nuclear DNA damage, activation of cell surface death receptors such as the tumor necrosis factor (TNF) superfamily, and formation of reactive oxygen species (ROS) accompanied by mitochondrial dysfunction and release of pro-apoptotic factors [8, 9]. UV irradiation has been demonstrated to produce reactive oxygen species (ROS) such as superoxide anion (•O₂) radical, hydroxyl radical, hydrogen peroxide and the highly reactive and destructive oxidant, peroxynitrite (ONOO⁻) [8-10].

To date, 25 selenoprotein genes have been identified in human. Known biological roles of selenoproteins include antioxidant enzymes, enzymes involved in thyroid hormone metabolism, and proteins functioning in selenium transport and heavy metal detoxification [11-16]. These include several well-known peroxidases and reductases such as glutathione peroxidases (GPxs), phospholipid hydroperoxide glutathione peroxidases (PHGPxs) and thioredoxin reductases (TRs), as well as other lesser characterized selenoproteins such as selenoprotein H (SelH) [17, 18]. Furthermore, selenoprotein P (SelP)

may also function as a neural survival factor [19, 20]. Emerging evidence suggests that SelP may also possess antioxidant activity [21, 22]. For example, SelP has been shown to function as a protectant against diquat-induced liver necrosis and lipid peroxidation in the rat [23, 24]. SelP is also protective against peroxynitrite-mediated oxidation and nitration [25]. The antioxidative effects of several other selenoproteins such as Gpx, PHGPxs and TRs have been well characterized [26, 27].

The function of SelH is largely unknown. Recently, it was demonstrated that overexpression of *Drosophila melanogaster* SelH (dSelH) significantly increased viability in embryos by decreasing lipid peroxidation [28]. The objectives of this study were to investigate whether human Selenoprotein H (hSelH) protects neuronal cells against UVB irradiation-induced cell death and if so, whether the protective effect may associate with ROS production prior to observable cell death. We first determined cell viability following a 7 mJ/cm² of UVB irradiation injury and 24 h of recovery in the murine hippocampal neuronal cell line Ht22, transfected with the hSelH gene in Murine Stem Cell Virus (MSCV) or the empty retroviral vector. We then measured O₂⁻-production after 5 h of recovery in hSelH or empty vector transfected cells using the oxidized dihydroethidine detection method [29, 30]. Our results are consistent with the concept that the neuroprotective effect of SelH is associated with its ability to reduce ROS production.

2. Materials and Methods

hSelH transfection procedures

The plasmid containing the gene encoding hSelH was purchased from Invitrogen (Carlsbad, CA, USA). The target gene was subcloned into Murine Stem Cell retroviral vector pMSCVpuro, purchased from BD Biosciences (Palo Alto, CA, USA). The packaging cells RetroPack PT67 (BD Biosciences) were plated in a 60 mm plate at 70% optical confluency 12 h before transfection. Cells were then transfected with 2 µg of hSelH-carrying retroviral vector or empty vector using Fugene method (Roche, Indianapolis, IN, USA). Culture medium was aspirated from cultures after 4 h of transfection. PT 67 cells were washed twice with PBS, and grown for 24 h in 3 ml complete medium. The cells were then selected by puromycin. The concentration of puromycin was optimized to kill the untransfected control PT67 cells in 7-10 d. The transfected PT67 cells that survived the puromycin selection stably produced virus. For viral infection of the target cells, Ht22 cells were plated 12 h before infection at ~70% optical confluency. The medium from transfected PT67 cells containing virus were collected, filtered through a 0.45 µm filter, and added to the Ht22 cells in the presence of 4 µg/ml Polybrene. The virus-containing medium was replaced after 24 h of incubation. After infection for 48 h, the target cells were subjected to puromycin selection. The selection lasted for ~10 d until the uninfected control cells were killed. The cells that survived the selection made up the hSelH-transfected Ht22 cell line (hSelH-Ht22) and MSCV-transfectant (MSCV-Ht22) cell lines.

Detection of mRNA overexpression on hSelH by Real time PCR

Total RNA was extracted from hSelH-Ht22 and MSCV-Ht22 cells using RNeasy Midi Kit (Qiagen, Valencia, CA, USA). cDNAs were synthesized from total RNA using the SuperScript III First-Strand Synthesis System (Invitrogen), and PCR amplification of target genes were carried out using the Platinum SYBR Green qPCR SuperMix-UDG kit (Invitrogen). The following specific primers were used to amplify hSelH, mSelH, and mGAPDH (house keeping gene) cDNAs:

hSelH	forward:
(5'-GCTTCCAGTAAAGGTGAACCCG-3')	
hSelH	reverse:
(5'-ACCCAAATCTCCCTACGACAGG-3')	
mSelH	forward:
(5'-GGAAGAAAGCGTAAGGCGGG-3')	
mSelH	reverse:
(5'-GGTTTGGACGGGTTCACTTGC-3')	
mGAPDH	forward:
(5'-TGACATCAAGAAGGTGGTGAAGC-3')	
and mGAPDH	reverse:
(5'-CCCTGTTGCTGTAGCCGTATTC-3')	

All PCR reactions were performed in duplicate 20 µl reaction volumes and detection was carried out with a LightCycler 2 real-time PCR machine (Biorad,

Hercules, CA, USA).

Cell maintenance and treatment

Murine hippocampal Ht22 neuronal cells were infected either with the MSCV empty vector or with hSelH in MSCV expression vector. For propagation, cells were fed with Dulbecco's Modified Eagle Medium (DMEM) with 10% fetal bovine serum (FBS), 2 mM glutamine, and 200 mM streptomycin/penicillin (Invitrogen) and then cultured at 70%-75% relative humidity in 5% CO₂ at 37°C. Media was renewed every 3 d. For cell viability assays, cells were seeded in 48-well cell culture plates (Corning, Aton, MA, USA) 48 h before treatment. The cells were allowed to reach 80 % optical confluency before UVB treatment.

UVB irradiation

Cells were seeded in 96 or 24 wells plate and propagated to reach 80% cell confluency. Prior to UVB irradiation, cells were washed twice with cold PBS to remove residual serum and dead cells. Cells were placed in serum-containing media (10%) media or without serum for 1 h of incubation in normoxic incubator at 37°C. Cells were then, submitted to different doses of UVB irradiation from a 20 watt MEABS-25 bulb (Ultra-Lum, Inc., Claremont, CA, USA), which emits most of its energy (60 %) with the UVB range (290-320 nm) with an emission peak at 310 nm band. The irradiation dosages given were 7 mJ/cm², 11.5 mJ/cm², and 23 mJ/cm². After UVB irradiation, cells were returned to the incubator for 5 h or 24 h of recovery at 37°C.

Measurement of the cytotoxicity

Cell viability was assessed using the Trypan blue exclusion assay as described previously [25]. Briefly, viable and dead cells were discriminated and counted using 0.4% Trypan blue staining with a hemacytometer (Sigma, Saint-Louis, MO, USA). Dead cells were counted as those stained blue by the Trypan blue stain, whereas live and apoptotic cells were those that had excluded the stain.

Detection of superoxide production

MSCV-Ht22 and hSelH-Ht22 cells were exposed to a 7 mJ/cm² of UVB irradiation in the absence of serum. Production of superoxide anion was assessed using dihydroethidine (HET) method [31-33] after 5 h of recovery following 7 mJ/cm² of UVB irradiation, because no frank cell death is seen at this time. HET was purchased from Molecular Probes (Eugene, OR, USA). Stock solution of HET (1 mg/ml) was prepared by dissolving in DMSO and stored, protected from light at -20°C. One µl (1 µg) of HET was added to 500 µl of cultures 15 min before UVB irradiation. HET is a cell-permeable fluorescent probe widely used to detect intracellular superoxide anion. It is proposed that the reaction between superoxide and HET results in the formation of a two-electron oxidized product, which binds to DNA and leads to the enhancement of red fluorescence after being oxidized by superoxide [31, 33, 34]. Twelve microscopic images at the magnification of

20X were captured and the number of oxidized HET cells, which represent superoxide production, were counted and presented as percentage of total counted cells. Because HET gives a light background staining, only cells stained with strong bright red color were counted as oxidized HET-positive cells.

Statistical analysis

All experiments were performed at least in triplicate, repeated two to three times and presented as mean \pm SD. ANOVA followed by Scheffe's test was employed for statistical analysis (GraphPad Prism software, SAS, San Diego, CA, USA). A *p*-value of < 0.05 was considered significant.

3. Results

hSelH infection

hSelH was stably infected into Ht22 cells using MSCV vector. The success of this infection was determined by detection of hSelH mRNA levels using quantitative real-time PCR. The mSelH was a low copy number gene in Ht22 cells. The hSelH mRNA level was increased by about 30 folds compared to endogenous mSelH mRNA level in hSelH-Ht22 cells (Table 1). In control MSCV-Ht22 cells, the murine SelH (mSelH) mRNA level, is comparable to that in SelH-Ht22 but the hSelH mRNA is undetectable.

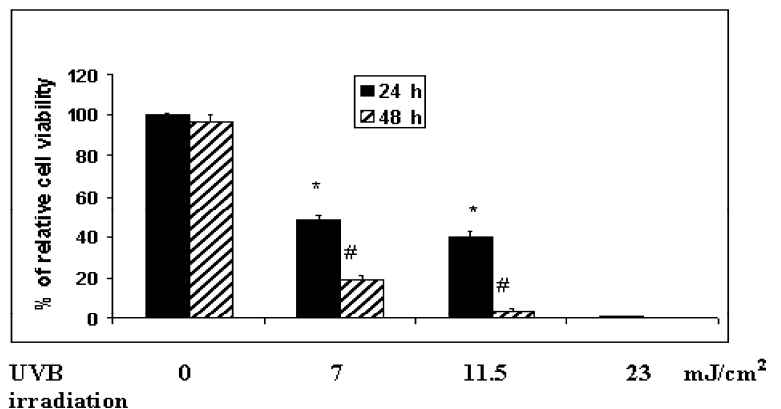
Table 1: Detection of mRNA overexpression on hSelH by Real time PCR. Both human SelH (hSelH) and mouse SelH (mSelH) genes were measured in hSelH-Ht22 and MSCV-Ht22 cells. Each target gene was amplified in duplicates. The table shows a significant drastic increased level of hSelH in Ht22-hSelH cells compared to controls (means \pm SD, *P* < 0.0001).

Target genes	MSCV-Ht22	SelH-Ht22
hSelH	Non-Detectable	0.557 \pm 0.017*
mSelH	0.0149 \pm 0.0019	0.0163 \pm 0.0005

Effects of variable UVB irradiation duration on cell viability

In order to identify an optimal time of UVB exposure, we exposed MSCV-Ht22-transfected and hSelH-Ht22-transfected cells to 0 mJ/cm², 7 mJ/cm², 11.5 mJ/cm², and 23 mJ/cm² of UVB irradiation. Cell viabilities were measured after 24 and 48 h of recovery. Cell viability after 24 h of recovery, was reduced from 100% in non-irradiated control to an average of 50%, 40% and 6% in 7 mJ/cm², 11.5 mJ/cm², and 23 mJ/cm² of UVB irradiation, respectively (Figure 1). Cell viabilities were further decreased after 48 h of recovery. Thus, the average survival rate dropped to 19%, 4% and 0% following 7 mJ/cm², 11.5 mJ/cm², and 23 mJ/cm² of UVB irradiation, respectively (Figure 1). This experiment showed that 7 mJ/cm² of UVB irradiation resulted in approximately 50% cell death at 24 h of recovery, which was determined to be the optimal length of injury. Therefore, a 7 mJ/cm² of UVB irradiation was used for the remaining studies.

Fig. 1. Variable UVB irradiation exposure parameters on cell viability in MSCV (empty vector) transfected Ht22 cells. The irradiation doses given to the cultures were 0 mJ/cm², 7 mJ/cm², 11.5 mJ/cm², and 23 mJ/cm², in DMEM medium with serum, respectively. The percentage of cell viability was determined after 24 and 48 h of recovery using the Trypan blue exclusion assay. About 100%, 50%, 40% and 6% of cell viability were observed at 24 h of recovery 0 mJ/cm², 7 mJ/cm², 11.5 mJ/cm², and 23 mJ/cm² of UVB irradiation, respectively. Cell viabilities were further significantly decreased to 21 %, 4% and 0% at 48 h of recovery at the indicated UVB doses in the Figure. Experiments were performed in triplicate and presented as means \pm SD. *Significantly different from non-irradiated control at 24 h (*P* < 0.05) at the indicated UVB dose. #Significantly different from 24 h of recovery at 48 h of recovery (*P* < 0.05).



Effects of hSelH overexpression on cell viability after UVB irradiation

Both MSCV-Ht22 and hSelH-Ht22 cells were exposed to a 7 mJ/cm² of UVB irradiation in the presence or absence of serum. The cell viabilities were examined at 24 h of recovery and the results were compared to MSCV and hSelH transfectants without UVB irradiation. As shown in Figures 2 and 3, MSCV-Ht22 cells exposed to 7 mJ/cm² of UVB irradiation in the presence of serum exhibited 48% cell viability compared to controls, as expected. However, the cell viability was significantly increased in cells overexpressing hSelH. That is, hSelH overexpression resulted in appreciatively 80% cell survival compared to controls and this was found to be significantly different than that observed with vector control (*P* < 0.0001). In the absence of serum, cell survival rate decreased to about 70% even without UVB irradiation, with no major difference between the MSCV-Ht22 and hSelH-Ht22 cells. Cell survival rate was further decreased to 23% in MSCV-Ht22 cells after a 7 mJ/cm² of UVB irradiation. However, with the overexpression of hSelH, cell survival only decreased to 52% (*P* = 0.0004) when cells were exposed to 7 mJ/cm² of UVB irradiation without supplementation of serum. Figure 2 shows representative photomicrographs taken from MSCV-Ht22 and hSelH-Ht22 cells treated with or without UVB irradiation in the presence or absence of serum. Figure 3 summarizes the cell viability data

collected from three independent experiments.

Fig. 2. Photomicrographs showing effects of hSelH overexpression cell viability in Ht22 cells treated with 7 mJ/cm² of UVB irradiation followed by 24 h of recovery. Increased cell debris was present in UVB radiated MSCV-Ht22 cell compared to non-treatment control cells. More viable cells were observed when hSelH transfected Ht22 cells were exposed to UVB irradiation. Images were taken at 20X magnification with a Zeiss LSM5 laser-scanning confocal microscope. Bar = 10 μm.

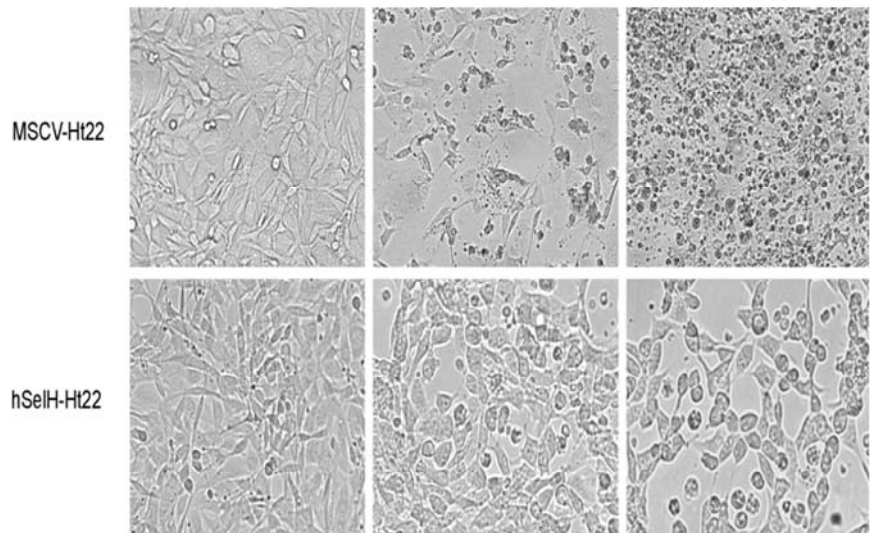
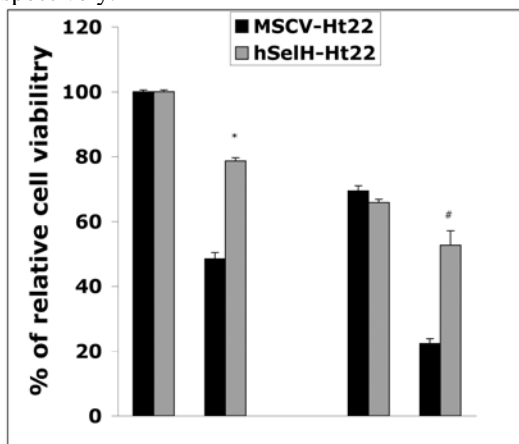


Fig. 3. Effects of hSelH overexpression on Ht22 cells exposed to 7 mJ/cm² of UVB irradiation with or without serum in media. Cell viability was examined after 24 h of recovery and presented as percentage (mean ± SD). Overexpression of hSelH protected cells from UVB-induced damage. *Significantly different from MSCV control, P < 0.0001 and #Significantly different from MSCV transfectant, P = 0.0004, in the presence or in the absence of serum, respectively.



Serum	+	+	-	-
UV	-	+	-	+

Overexpression of hSelH reduces superoxide formation

In order to determine the effects of UVB irradiation and overexpression of hSelH on free radical production, we measured superoxide production using the oxidized HET method. Since a majority of MSCV-transfected cells die after 24 h of recovery, we selected an early recovery time point (5 h) for detection of superoxide formation. A 7 mJ/cm² of UVB irradiation resulted in 48% of oxidized HET positive stained cells in MSCV-Ht22 after 5 h of recovery. In contrast, the percentage was decreased to 16% (P =

0.0017) in hSelH-Ht22 cells (Figure 4). A set of fluorescent photomicrographs from MSCV-Ht22 and hSelH-Ht22 cells that were exposed to 7 mJ/cm² of UVB irradiation is shown in Figure 5.

4. Discussion

Previous published studies on neuronal cells have shown that UV irradiation induced apoptosis is associated with activations of caspase, calpain, JNK, and the mitochondrial apoptosis pathway [4-7]. UV irradiation provokes oxidative stress through the generation of ROS and to induce apoptosis by activation of both receptor-mediated and mitochondrion-initiated cell death pathways [8, 9, 35, 38]. In this study, O₂⁻ production increased significantly after 5 h of recovery following a 7 mJ/cm² of UVB irradiation. This is consistent with the results published by Maglio and colleagues who observed a 5-fold increase in superoxide production in the skin after UV irradiation [39]. Under physiological conditions, superoxide dismutase (SOD) reduces O₂⁻ to H₂O₂ that is further reduced to H₂O by catalase or glutathione peroxidase. However, excess production of O₂⁻ leads to formation of the highly toxic hydroxyl radical, •OH, in the presence of metal or peroxynitrite (ONOO⁻) in the presence of nitric oxide (NO). Hydroxyl radicals and ONOO⁻ are potent oxidants of proteins and DNA, and alter the function of key enzymes through nitration and nitrosylation. Increased ROS in the mitochondrion induces formation of a mitochondrial permeability transition (MPT) pore. The MPT allows release of proapoptotic proteins that subsequently activate a cascade of proteases, ultimately leading to cell death [40, 41].

The selenoprotein family contains 25 members in the human proteome [13]. A majority of published studies have been focused on the expression and functions of GPxs, TRs, iodothyronine deiodinases, and SelP. Studies have shown that these enzymes

possess antioxidative effects, in addition to their roles in male fertility, thyroid hormone metabolism, heavy metal detoxification and neurological function and development [20, 26, 42]. For example, glutathione peroxidase acts as peroxynitrite reductase, preventing oxidation and nitration reactions resulting from peroxynitrate [17, 18]. Similarly, SelP protects low-density lipoproteins (LDL) against oxidation (traps ONOO⁻ and protect astrocytes from various injuries including irradiation and *tert*-butyl hydroperoxide-induced oxidative stress [10, 21, 22, 27]. The functions of most of the recently identified selenoproteins, including SelH, have not been elucidated.

To study whether overexpression of hSelH is capable of protecting cells against UVB-induced oxidative stress, we first transfected the murine hippocampal Ht22 neuron cell line with the hSelH or the empty vector MSCV. As shown in Table 1, the hSelH mRNA level was 30 fold higher than the endogenous mSelH mRNA level in hSelH-Ht22, suggesting a high transcriptional rate of the transfected hSelH gene. We then exposed the cells to a 7 mJ/cm² of UVB irradiation. Overexpression of hSelH reduced cell death caused by UVB irradiation. This result is consistent with a previous publication showing that overexpression of a selenoprotein homologous to glutathione peroxidase blocks UV-induced cell death [43]. We finally measured O₂⁻ production in cultured cells at 5 h after a 7 mJ/cm² of UVB irradiation. The results showed that overexpression of hSelH inhibited O₂⁻ formation in neurons in the early recovery phase after irradiation. Concurrently, the percent of oxidized HET-positive cells detected at 5 h of recovery was correlated with the percent of cell death observed at 24 h of recovery, suggesting increased superoxide production might be the underlying cause for the observed cell death. Our findings are in accordance with a previous study reported by Morozova and colleagues [22]. In that study, it was demonstrated that silencing the expression of *Drosophila melanogaster* SelH (dSelH) gene significantly reduced the total antioxidant capacity and embryo viability. In contrast, overexpression of dSelH aided cell survival by reducing lipid peroxidation *in vivo* [28].

It is not clear through what mechanism(s) hSelH reduces ROS production. A parallel study carried by Panee and colleagues has shown that overexpression of SelH sustains the total glutathione (GSH) level in Ht22 cells during oxidative stress (Panee et al., manuscript in preparation). GSH plays multiple roles in the protection of cells from ROS, electrophiles and xenobiotics [44, 45]. More interestingly, SelH overexpression upregulated the expression of genes involved in antioxidant defense (Panee et al., manuscript in preparation) and phase II detoxification [44]. Therefore, it is likely that SelH overexpression decreases the ROS level and sustains cell viability in Ht22 cells after UVB irradiation via these pathways.

Taken together, our study shows that

overexpression of hSelH protects cells against UVB irradiation-induced cell death by ameliorating ROS formation in the early recovery period. The results are in line with previous publications showing that selenoproteins possess antioxidative capacity.

Fig. 4. Percentage of oxidized HET positive cells in MSCV-Ht22 and hSelH-Ht22 cells after 5 h of recovery follow a 7 mJ/cm² of UVB irradiation. hSelH overexpression significantly reduced oxidized HET positive cells compared with MSCV-Ht22 cells. *Significantly different from MSCV-Ht22, P = 0.0017.

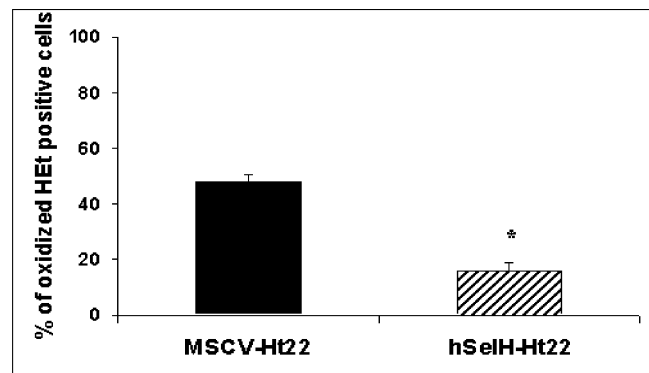
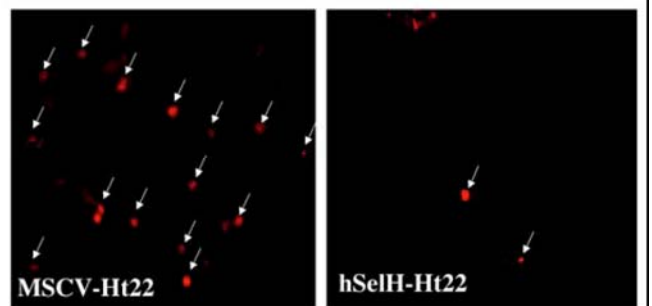


Fig. 5. Representative photomicrographs showing oxidized HET positive staining in MSCV-Ht22 (A) and hSelH-Ht22 cells (B) after 5 h of recovery follow 7 mJ/cm² of UVB irradiation. Arrows denote oxidized HET cells which indicating the formation of superoxide. Images were captured at 20X magnification with a Zeiss LSM5 laser-scanning confocal microscope. Bar = 10 μm.



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Conflict of interests

The authors have declared that no conflict of interest exists.

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