

Research Paper

Temporal Expression of Mutant LRRK2 in Adult Rats Impairs Dopamine Reuptake

Hongxia Zhou^{1,*}, Cao Huang^{1,*}, Jianbin Tong¹, Weimin C Hong², Yong-Jian Liu^{2,}, Xu-Gang Xia^{1,}

1. Department of Pathology, Anatomy & Cell Biology, Thomas Jefferson University, 1020 Locust Street, Philadelphia, PA 19107, USA
2. Department of Neurobiology, University of Pittsburgh School of Medicine, Pittsburgh, PA 15213, USA

* These authors contributed equally to this work.

✉ Corresponding author: H.Z. (Hongxia.zhou@jefferson.edu), Y.J.L. (yjliu@pitt.edu), or X.G.X. (xugang.xia@jefferson.edu).

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Abstract

Parkinson's disease (PD) results from progressive degeneration of dopaminergic neurons. Most PD cases are sporadic, but some have pathogenic mutation in the individual genes. Mutation of the *leucine-rich repeat kinase-2 (LRRK2)* gene is associated with familial and sporadic PD, as exemplified by G2019S substitution. While constitutive expression of mutant *LRRK2* in transgenic mice fails to induce neuron death, transient expression of the disease gene by viral delivery causes a substantial loss of dopaminergic neurons in mice. To further assess *LRRK2* pathogenesis, we created inducible transgenic rats expressing human *LRRK2* with G2019S substitution. Temporal overexpression of *LRRK2*^{G2019S} in adult rats impaired dopamine reuptake by dopamine transporter (DAT) and thus enhanced locomotor activity, the phenotypes that were not observed in transgenic rats constitutively expressing the gene throughout life time. Reduced DAT binding activity is an early sign of dopaminergic dysfunction in asymptomatic subjects carrying pathogenic mutation in *LRRK2*. Our transgenic rats recapitulated the initiation process of dopaminergic dysfunction caused by pathogenic mutation in *LRRK2*. Inducible transgenic approach uncovered phenotypes that may be obscured by developmental compensation in constitutive transgenic rats. Finding in inducible *LRRK2* transgenic rats would guide developing effective strategy in transgenic studies: Inducible expression of transgene may induce greater phenotypes than constitutive gene expression, particularly in rodents with short life time.

Key words: LRRK2; Parkinson's disease; rats; genetic model; dopamine transporter; dopaminergic neurons

Introduction

Parkinson's disease (PD) is a common neurodegenerative disease caused by progressive degeneration of dopaminergic neurons in the substantia nigra pars compacta (SNpc). How neurons degenerate in PD is largely unknown. While most PD cases are sporadic, approximately 10% of the cases have genetic mutation in the individual genes, including

a-synuclein, *parkin*, *DJ-1*, *pink1*, and *leucine-rich repeat kinase 2 (LRRK2)* (1-6). Unlike mutation in the other PD genes, mutation of *LRRK2* causes late-onset Parkinsonism indistinguishable from idiopathic PD (7-10), suggesting a role for *LRRK2* in sporadic PD. Indeed, a common mutation in *LRRK2* (G2019S substitution) occurs in sporadic PD (10), underscoring the

importance of LRRK2 for understanding PD pathogenesis.

LRRK2 is a member of the newly defined ROCO protein family, which belongs to the Ras-GTPase superfamily (5, 6). Limited information from orthologous proteins in *Dictyostelium Discoideum* suggests a role for ROCO proteins in regulating cytoskeletal structures (11). Like the other ROCO proteins, LRRK2 contains a conserved GTPase-like domain (Roc) and a mixed lineage kinase-like kinase domain (5, 12). G2019S mutation located within the kinase domain of LRRK2 augments kinase activity implicated in LRRK2 neurotoxicity (12-15). Mutation of LRRK2 within its Roc domain (R1440C substitution) also enhances its kinase activity (16), suggesting an intrinsic role of kinase activity in the neurotoxicity. To understand LRRK2 pathogenesis, a critical step is establishing animal models reproducing PD phenotypes observed in patients. Similar to the other genetic mouse models of PD (17-21), mutant mice with endogenous *LRRK2* deleted or with mutant LRRK2 introduced fail to develop dopaminergic neuron death (22-29), although some strains display dystrophy of dopaminergic neurites and disturbance to dopaminergic neurotransmission (28, 29). Like transgenic mice, LRRK2 transgenic *Drosophila* of different strains also develops diverse phenotypes (30, 31). Phenotypic expression in LRRK2 transgenic animals largely depends on transgenic strains (22-31). Even though chromosomal positional effect on transgene expression is ameliorated by using a bulk transgene, such as bacterial artificial chromosome (BAC), BAC transgenic mice expressing mutant LRRK2 develop varying phenotypes without neuronal loss (28, 32). Such findings in transgenic animals underscore the importance of diverse models for dissecting LRRK2 pathogenesis.

To further assess LRRK2 mutation, we developed transgenic rats expressing mutant human LRRK2 in a controlled manner. Temporal, but not constitutive, expression of human LRRK2 with G2019S substitution enhanced locomotor activity in aged rats. Microdialysis analysis of living rats attributed the enhanced locomotor activity to impaired dopamine reuptake by dopamine transporter (DAT). As a result of compromised DAT activity, amphetamine-evoked dopamine release and amphetamine-elicited locomotor activity were reduced in mutant LRRK2 transgenic rats. Intriguingly, reduced DAT binding is the earliest indication of dopaminergic dysfunction in asymptomatic subjects carrying a pathogenic mutation in *LRRK2* (33). Thus, our transgenic rats recapitulated the initiation process of dopaminergic dysfunction caused by mutation in *LRRK2*.

Materials and Methods

Transgenic rat production and behavioral analyses

LRRK2 and tTA transgenic rats were produced by pronuclear injection as described previously (34). TRE-LRRK2^{G2019S} transgenic rats were crossed with CAG-tTA transgenic rats to produce the double transgenic offspring that expressed mutant human LRRK2 in the absence of Doxycycline (Dox). To mimic constitutive gene expression, breeding rats and their offspring were not given Dox through their lifetime. To achieve temporal expression of LRRK2 transgene in adulthood, breeding rats and their offspring were constantly given Dox in drinking water (50µg/ml) until the double transgenic offspring reached 5 months of age. LRRK2 transgenic rats and the control rats were subject to open field activity assay (Med Associates), which measured the total distance and the total stereotypic move the rat made in 20 minutes. Elicited activities were measured at 5 minutes after administration of amphetamine (1mg/kg, I.p.) or 20 minutes after administration of nomifensine (1mg/kg, I.p.). Animal use followed NIH guidelines and the animal use protocol was approved by the Institutional Animal Care and Use Committees at Thomas Jefferson University.

Microdialysis and HPLC analysis

As described previously (35), the extracellular level of striatal dopamine was measured by intra-brain microdialysis combined with HPLC in free-moving rats. Anesthetized rats were implanted with a guide cannula the day before dialysis. On the following day, dialysis probe (BASi) was inserted through the guide cannula into the left striatum (coordinates: 1 mm anterior to bregma; 2.6 mm left from sagittal line; 4 mm ventral to the skull). Continuous perfusion was maintained with modified Ringer's solution (147 mM Na⁺, 4 mM K⁺, 1.3 mM Ca²⁺, 1 mM Mg²⁺, and 155.6 mM Cl⁻). Basal dopamine release was established within two hours when comparable levels of released dopamine were obtained in three consecutive samplings (20 minutes each). Amphetamine (1.3 mM) and nomifensine (200 µM) were applied to dialysis solution for 40 minutes and evoked dopamine release was measured subsequently. Dopamine concentration in dialysis solution was instantly measured by HPLC. By the end of dialysis, the right striatum of dialyzed rats was dissected and homogenized in 0.1 M perchloric acid and cleared tissue lysates were measured by HPLC for the concentration of dopamine and dopamine metabolites. Tissue pellets were dissolved in 1M NaOH and protein concentration was

determined. Tissue contents of striatal dopamine were adjusted with protein concentration. HPLC analysis was done as described previously (36).

Histology and stereological cell counting

Immunostaining and stereological cell counting were done as described previously (36, 37). Rat's brain was cut into serial sections of 30 μm on a Cryostat. Every fourth section through the SNpc and every fourth section through the locus coeruleus (LC) were immunostained for TH (Pel-Freez; 1: 1000). TH positive neurons in the SNpc and in the LC were estimated by stereological cell counting. Coronal sections of rat's brain through SNpc and striatum were also immunostained with mouse monoclonal antibody to α -synuclein (1: 500; Chemicon), chicken antibody to ubiquitin (1: 1000; Sigma), or rabbit polyclonal antibody to phospho-tau (1: 500; Sigma).

[³H]-dopamine uptake assay

The assay was performed as described previously (38). The human neuroblastoma SH-SY5Y cells that stably expressed human DAT-EGFP-C1 were seeded into 96-well plates and were then transfected with LRRK2 constructs using Lipofectamine 2000 (Invitrogen, CA). Two days following transfection, cells were washed with PBSCM (PBS with 0.1mM CaCl₂, 1 mM MgCl₂), and were incubated with 50 nM [³H]-dopamine plus varying concentrations of unlabeled dopamine for 5 min at room temperature. Retained radioactivity in cells was determined by liquid scintillation spectrometry. Data from triplicate samples were analyzed according to the Michaelis-Menten kinetic equation using GraphPad Prism 4.0.

Results

Enhanced locomotor activity in aged rats expressing mutant LRRK2 in adulthood

To further examine the pathogenesis of mutation in *LRRK2*, we created transgenic rats overexpressing a mutant LRRK2 with G2019S substitution (34). Transgenic studies of amyotrophic lateral sclerosis suggest that mice and rats develop diverse phenotypes, even if the same disease gene is overexpressed (37, 39). Rats confer advantages over mice in behavior tests that are crucial to the modeling of neurological diseases such as PD (40, 41). To increase the diversity of animal models for PD, we chose rats to model the phenotypes of pathogenic mutation in *LRRK2*. Since developmental compensation for transgene expression may compromise phenotypic expression in transgenic animals, we used a Tet-inducible system to express

mutant LRRK2 transgene temporally in adult rats (34). As described previously (34), one transgenic line carrying two copies of LRRK2 transgene was established and this line expressed mutant human LRRK2 robustly in the midbrain when crossed onto CAG-tTA transgenic background.

The mutant LRRK2 transgene was driven by the TRE promoter and thus its expression depends on tTA activation and is subject to Dox regulation (34). In the absence of Dox, mutant LRRK2 transgene was constitutively and fully expressed (34), exhibiting a constitutive expression pattern (Fig. 1A). To obtain temporal overexpression of mutant LRRK2 in adult rats, we supplied transgenic rats with Dox in drinking water during embryonic and postnatal development such that the mutant LRRK2 transgene was constantly suppressed from expression in transgenic rats (34). Dox-treated rats were deprived of Dox at the age of 5 months, and they soon began to express mutant LRRK2 after Dox withdrawal (Fig. 1A). Constitutive overexpression of mutant LRRK2 failed to induce behavioral phenotypes in transgenic rats (Fig. 1B). In contrast to constitutive overexpression, temporal overexpression of mutant LRRK2 caused abnormal locomotor activity in aged, but not in young, transgenic rats (Fig. 1B). The results suggest that temporal gene expression uncovered the phenotypes obscured by developmental compensation.

Unaltered number of dopaminergic neurons and unaltered content of striatal dopamine in LRRK2 transgenic rats

PD is characterized by progressive loss of dopaminergic neurons in the SNpc (42). We then examined dopaminergic neurons by stereological cell counting. Neither constitutive nor temporal overexpression of mutant LRRK2 altered the number of dopaminergic neurons in transgenic rats at advanced ages (Fig. 2A-1G). Noradrenergic neurons in the locus coeruleus (LC) are often lost at the early stages of PD (43). We also assessed the TH-positive neurons in the LC and found no loss of the neurons in aged rats constitutively or temporally expressing mutant LRRK2 (Fig. 2H). The axonal terminals of dopaminergic neurons are the primary targets of degeneration in PD (44). We measured the contents of the neurotransmitter dopamine in the striatum, but did not detect any change to the total striatal dopamine in mutant LRRK2 transgenic rats, although the transgenic rats expressing mutant LRRK2 temporally in adulthood developed abnormal locomotor activity in advanced ages (Figs. 1B and 2I-2K). Proteinopathy is considered a characteristic of PD (42, 45). In our mutant LRRK2 transgenic rats, we did not detect any

inclusion positive for α -synuclein, ubiquitin, or phosphorylated Tau.

Impaired reuptake of dopamine in aged rats overexpressing mutant LRRK2 in adulthood

Temporal overexpression of mutant LRRK2 enhanced locomotor activity in aged rats (Figs. 1 and 4), indicating that dopaminergic neurotransmission was altered. In the aged rats, however, the content of striatal dopamine was unaltered (Fig. 2I), suggesting that dopamine synthesis and storage may not be interrupted by temporal overexpression of mutant LRRK2. The extracellular level of dopamine released from the neuronal terminals is virtually related to dopaminergic function. Using intra-brain microdialysis, we assessed dopamine release in the striatum of free-moving transgenic rats and detected a significant increase in dopamine release in temporally, but not in constitutively, mutant LRRK2-expressing rats (Fig. 3). Enhanced locomotor activity in mutant LRRK2 rats is attributable to elevated levels of dopamine in the synaptic cleft. Elevation of extracellular dopamine levels may result from increased release or reduced reuptake of dopamine. Nomifensine, a selective DAT

inhibitor, increased dopamine release to comparable levels between the mutant LRRK2 transgenic rats and the control rats (Fig. 3A). This increased release of dopamine upon nomifensine stimulation was significantly reduced by temporal, but not by constitutive, expression of mutant LRRK2 (Fig. 3B). The results suggest that dopamine release was unaltered but that dopamine reuptake was impaired in the mutant LRRK2 rats. Amphetamine is structurally similar to dopamine and has a higher affinity for DAT and vesicular monoamine transporter type 2 (VMAT2) than dopamine. The affinity of amphetamine is higher for DAT than for VMAT2. Amphetamine-evoked release of dopamine was compromised in transgenic rats expressing mutant LRRK2 in adulthood (Fig. 3), confirming that DAT activity was impaired in the mutant rats at advanced ages. Accordingly, amphetamine- and nomifensine-elicited locomotor activity was reduced in aged rats that temporally, but not constitutively, overexpressed mutant LRRK2 (Fig. 4A-4D). Collectively, the results suggest that DAT activity was impaired in aged rats expressing mutant LRRK2 in adulthood.

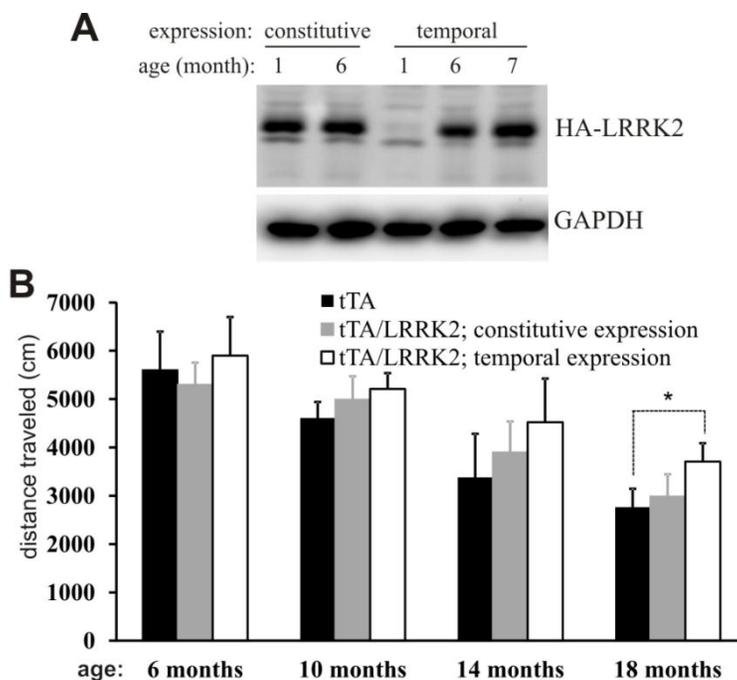


Figure 1. Temporal overexpression of mutant LRRK2 alters locomotor activity in aged rats. **A**, Immunoblotting revealed a full recovery of LRRK2 transgene expression in the brainstem after Doxycycline (Dox) withdrawal. Brain tissues were dissected from transgenic rats carrying CAG-tTA and TRE-hLRRK2^{G2019S}-HA transgenes. The double-transgenic rats were untreated with Dox throughout their lives (constitutive expression) or treated with Dox (50 μ g/ml in drinking water) from the embryonic stage to 5 months of age (temporal expression). Human LRRK2 was detected with an antibody to HA tag fused to the C-terminal of the mutant human LRRK2. GAPDH immunoreactivity was detected as a control for equal loading. **B**, Open field activity assay revealed an increase in locomotor activity in aged rats temporally, but not constitutively, overexpressing the mutant LRRK2. Data are means + SEM (n = 10). * $p < 0.05$.

Figure 2. Overexpression of mutant LRRK2 causes no loss of nigral dopaminergic neurons and striatal dopamine contents in aged rats. A-F, Representative photos of low (A-C) or high (D-F) magnification show the SNpc of transgenic rats at 18 months of age. Transgenic rats carried a CAG-tTA single transgene (1: tTA), or CAG-tTA and TRE-LRRK2^{G2019S} double transgenes (2 and 3: LRRK2). Constitutive expression of mutant LRRK2 (2: constitutive LRRK2) was achieved by withholding Dox throughout rat's lives, and temporal expression of the transgene (3: temporal LRRK2) was achieved by withdrawing Dox from rats at the age of 5 months. **G, H**, Stereological cell counting revealed no loss of TH-positive neurons in the SNpc and locus coeruleus (LC). Data are means \pm SEM ($n = 7$). **I-K**, HPLC revealed the contents of striatal dopamine (DA) and DA metabolites (DOPAC and HVA) in transgenic rats. Data are means \pm SEM ($n = 10$). Experimental group number in panels G-K corresponds to that in panels A-F.

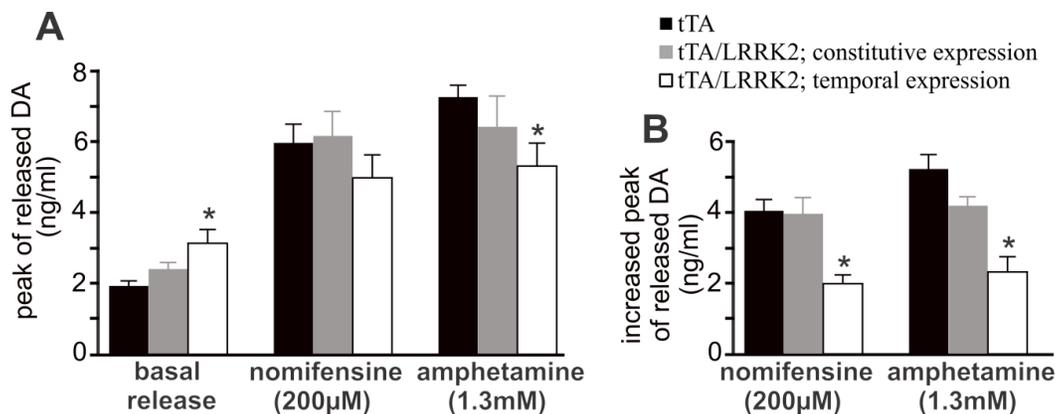
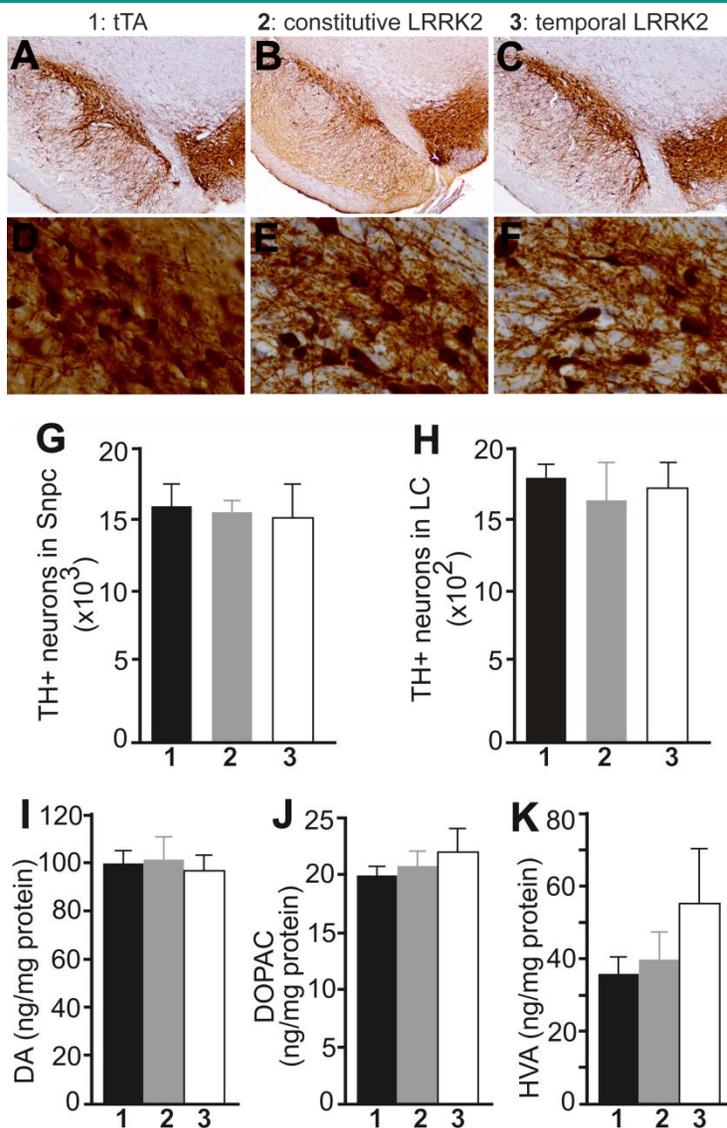


Figure 3. Overexpression of mutant LRRK2 impairs DA reuptake in aged rats. **A**, Intra-brain microdialysis revealed the extracellular levels of DA in the striatum of free-moving rats at 18 months of age. Basal DA release was established in living rats when three consecutive samplings resulted in a sustainable level of DA in dialysates in the absence of stimulation. The peaks of stimulated DA release were detected after nomifensine (200 μ M) or amphetamine (1.3 mM) was applied to dialysate. Data are means \pm SEM ($n = 8$). * $p < 0.05$ compared to CAG-tTA single transgenic rats (tTA). **B**, Increased peak of stimulated DA release was calculated for each animal by subtracting basal release from stimulated release. * $p < 0.05$ compared to either tTA single transgenic rats or constitutive LRRK2-expressing rats.

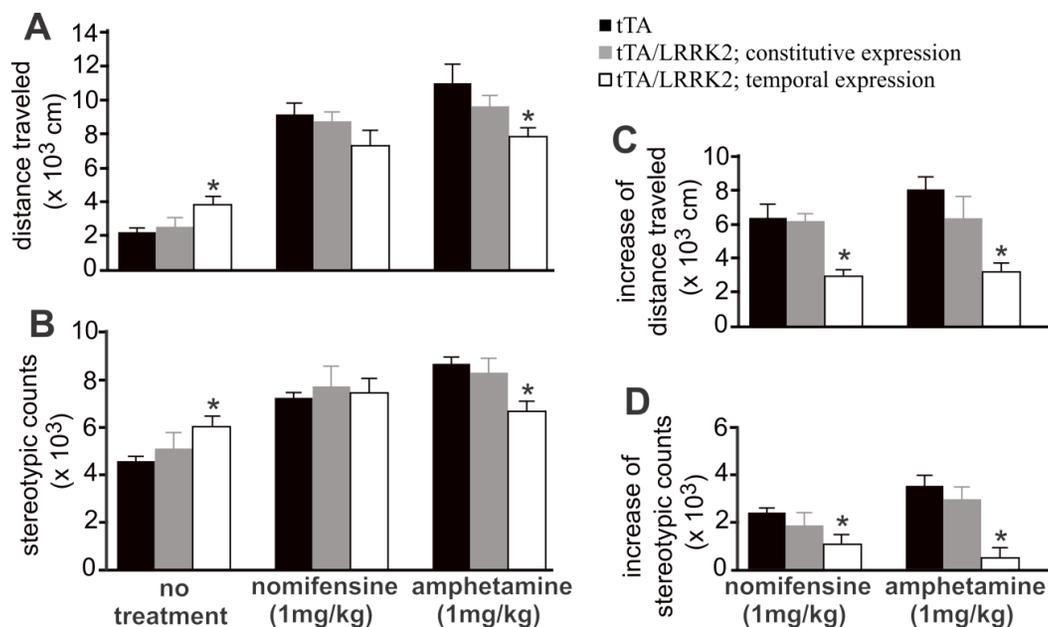


Figure 4. Temporal overexpression of mutant LRRK2 increases spontaneous, but reduces evoked, locomotor activity in aged rats. A, B, Open field activity assay measured locomotor activity in transgenic rats treated or untreated with nomifensine or amphetamine. Transgenic rats (18 months old) were subject to behavior test. C, D, Increased locomotor activity in response to drug stimulation was calculated by subtracting spontaneous activity counts from evoked activity counts. Data are means \pm SEM ($n = 10$). * $p < 0.05$ compared to CAG-tTA single transgenic rats (tTA).

No direct interaction of mutant LRRK2 with dopamine transporter

Microdialysis revealed that dopamine reuptake was impaired in aged rats expressing mutant LRRK2 temporally in adulthood (Fig. 3). To further assess the potential interaction of mutant LRRK2 with DAT, we used the neuroblastoma cells SH-SY5Y as an *in vitro* model of dopaminergic neurons. The SH-SY5Y cells that were stably transfected with DAT-expressing plasmid were transiently transfected with normal or mutant LRRK2-expressing constructs. In the presence of forcedly overexpressed LRRK2 variants, dopamine uptakes were determined and no significant difference between LRRK2 variants tested was observed (Fig. 5). The results suggest that normal and mutant LRRK2 do not interact with DAT directly.

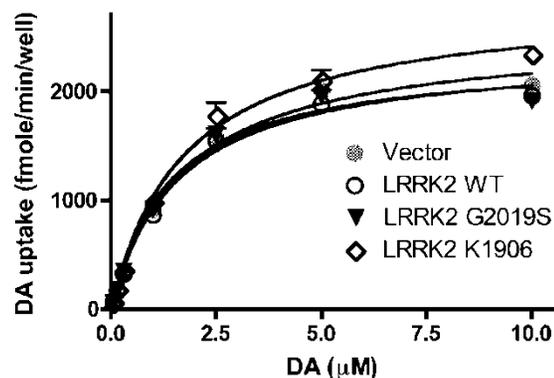


Figure 5. Overexpression of LRRK2 variants has no direct effect on the transport activity of DAT heterogeneously expressed in dopaminergic cells. SH-SY5Y cells stably expressing GFP-DAT were seeded in 96-well plates and transiently transfected with indicated constructs. [3 H]-DA uptake was determined 2 days after transfection. Data were analyzed according to the Michaelis-Menten kinetics using GraphPad Prism 4.0 software. The V_{max} and K_m values derived from the regression curves were not significantly different.

Discussion

Because G2019S mutation in *LRRK2* is observed in both the familial and sporadic PD (10), we chose *LRRK2*^{G2019S} as an example of *LRRK2* mutations for transgenic study. Temporal, but not constitutive, overexpression of the mutant *LRRK2* induced unexpected phenotypes, including enhanced locomotor activity and impaired dopamine reuptake. Altered locomotor activity in transgenic rats was attributed to elevated levels of extracellular dopamine revealed by intra-brain microdialysis. The extracellular level of dopamine is an outcome of the balance between dopamine release and reuptake. Blocking dopamine reuptake with the DAT-specific inhibitor nomifensine had a reduced effect on extracellular dopamine. DAT-mediated reuptake of dopamine can be reverted by amphetamine. In *LRRK2* transgenic rats, amphetamine had a reduced effect on the extracellular level of released dopamine. Pharmacologic analyses suggest that DAT-mediated dopamine reuptake was impaired in mutant *LRRK2* transgenic rats. The behavioral change was observed in aged, but not in young, transgenic rats, implying that DAT activity was impaired gradually as mutant *LRRK2* was overexpressed. The phenotypes were induced by temporal, but not by constitutive, overexpression of pathogenically mutated *LRRK2*, although the expression levels of mutant *LRRK2* were comparable between temporal and constitutive expression patterns. The findings suggest that damage to DAT activity was an indirect effect of mutant *LRRK2*. This notion is further supported by *in vitro* analysis, which did not detect any direct interaction of mutant or normal *LRRK2* with DAT. Temporal expression of mutant *LRRK2* in adult rats caused gradual damage to DAT-mediated dopamine reuptake.

Neuronal death in PD is considered a dying-back process and the neurites of dopaminergic cells are thought to be the primary targets for degeneration (28, 44). Most studies of genetic PD models reveal no dopaminergic neuron death, but do reveal damage to dopamine neurotransmission (18, 23, 29, 32, 46). As impaired D2 autoreceptor function is revealed in DJ-1 knockout and in *LRRK2* knockin mice (18, 29), DAT-mediated dopamine reuptake was, for the first time, revealed impaired in a transgenic model for PD. Our results, though unexpected, are consistent with clinical findings in mutant *LRRK2* carriers who display reduced DAT binding at asymptomatic stages (33). Elevation of extracellular dopamine may cause oxidative damage to dopaminergic neurites and is possibly an early event in PD pathogenesis. Compared to constitutive gene expression, temporal

overexpression of mutant *LRRK2* in adult rats induced a greater phenotype, possibly because it avoided developmental compensation for transgene expression. Our finding may help interpret why transient expression of PD genes by viral delivery often produces great disease phenotypes that could not be reproduced by constitutive expression of the disease genes in transgenic animals (26, 28, 29, 32, 47, 48).

Inducible *LRRK2* transgenic rats recapitulated the early pathology observed in asymptomatic subjects who carry pathogenic mutation in *LRRK2*. However, our transgenic rats did not develop dopaminergic neurodegeneration in their lifetime. The limited copy of mutant *LRRK2* transgene (only 2 copies) may yield insufficient disease protein, which was below the threshold to induce dopaminergic neuron death in a rat's lifetime. Phenotypic expression in rats may be improved by increasing expression of the *LRRK2* transgene. As PD is considered a multifactorial disease, multiple genetic and environmental factors may interact to induce dopaminergic neuron death (37). Indeed, varied disease penetrance and varied onset time are observed in the subjects carrying pathogenic mutation in *LRRK2* (9), favoring multifactorial pathogenesis in PD. Mutation of a single PD gene, particularly with low yield, may not be sufficient to induce neuron death within the short lifespan of rodents. Based on the multifactorial pathogenesis of PD, a combination of multiple disease factors (including environmental and genetic factors) will improve phenotypic expression in animal models.

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Conflict of Interests

The authors have declared that no conflict of interest exists.

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