

Research Paper

P-selectin-mediated LOX expression promotes insulinoma growth in Rip1-Tag2 mice by increasing tissue stiffness

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Abstract

P-selectin, a cell adhesion molecule, is an important member of the selectin family. Recent studies have shown that P-selectin deletion inhibits tumor growth in Rip1-Tag2 mice by suppressing platelet accumulation in tumor tissues. This study aimed to evaluate whether and how P-selectin affects tumor stiffness in Rip1-Tag2 mice. To explore the role of P-selectin in tissue stiffness, we demonstrated that tumor progression in Rip1-Tag2 mice was correlated with tissue stiffness using immunofluorescence and histological staining. Furthermore, we showed that P-selectin deficiency significantly decreased tissue stiffness by inhibiting lysyl oxidase (LOX) expression. Our experiments involving Rip1-Tag2 mice treated with the LOX inhibitor BAPN showed that BAPN significantly abolished collagen deposition to decrease tumor stiffness and thus inhibit tumor growth. These results indicate that P-selectin deletion significantly decreases tumor stiffness in Rip1-Tag2 mice by inhibiting LOX expression. Further study demonstrated that P-selectin-mediated platelet accumulation increases tissue stiffness mainly by increasing LOX expression and thus promotes tumor growth. Therefore, P-selectin may be an effective therapeutic targeting for treating human insulinomas.

Key words: P-selectin, insulinoma, tissue stiffness, LOX, Rip1-Tag2 mice

Introduction

P-selectin (CD62P), a cell adhesion molecule, is an important member of the selectin (CD62) family. Many studies have reported that P-selectin has significant effects on the growth and metastasis of many cancers including melanoma, lung cancer, and colorectal carcinoma. Furthermore, our previous study demonstrated that P-selectin deletion inhibited melanoma metastasis by suppressing platelet adhesion and that P-selectin-mediated platelet adhesion promoted intestinal tumor growth in *Apc^{Min/+}* mice [1,2]. *Apc^{Min/+}* mice are a spontaneous intestinal adenomatous polyps model, as they contain an *Apc* gene mutation that causes multiple intestinal

neoplasia (*Min*). Importantly, we found that P-selectin deficiency inhibited insulinoma progression using Rip1-Tag2 transgenic mice and Rip1-Tag2;*P-sel*^{-/-} mice [3]. We also found that large numbers of platelets accumulated in Rip1-Tag2 mouse insulinomas via a P-selectin-dependent mechanism [3]. Rip1-Tag2 mice are a spontaneous insulinoma model resulting from the expression of the SV40 T antigen, which functions as an oncogene, in β -cells [4]. Tumor development in Rip1-Tag2 mice involves a multistep process including a hyperplastic islet stage, an angiogenic islet stage, a tumor stage and an invasive cancer stage. Angiogenic islets appear in

six-week-old Rip1-Tag2 mice, and some of these angiogenic islets develop into tumors in nine-week-old Rip1-Tag2 mice [4]. However, the relationships between P-selectin and P-selectin-mediated platelet aggregation and insulinoma ECM stiffness remain unclear.

Some studies have shown that many types of tumors are stiffer than adjacent tissues and that tumor ECM stiffness is strongly correlated with solid tumor progression and aggression [5,6]. Collagen is the most abundant component of tumor extracellular matrix (ECM) [7]. Evidence from various studies regarding human cancers and experimental tumor models indicates that cross-linked collagen accumulation can enhance tumor ECM stiffness. Acerbi *et al* reported that human breast and epithelial cancer progression is facilitated by collagen deposition and remodeling, which are closely associated with tumor stiffness [8-10]. However, the relationship between tumor stiffness and insulinoma development and progression remains unknown.

Lysyl oxidase (LOX), a secreted copper-dependent amine oxidase, is a collagen cross-linker [11]. Studies have demonstrated that LOX is highly expressed in tumor cells and that LOX inhibitors can inhibit tumor progression by decreasing the amount of collagen cross-links in tumors [12]. Breast cancer patients exhibiting high levels of LOX expression have a greater chance of developing metastasis and thus experience shorter survival times than patients exhibiting low levels of LOX expression [13-17]. LOX-mediated cross-linking and increases in collagen concentrations can heighten collagen stiffness, indicating that ECM stiffness promotes breast cancer growth and metastasis [18] and that the extent of collagen deposition and the amount of collagen cross-links in tumors control tumor progression [19].

In this study, we sought to investigate the relationship between P-selectin expression and insulinoma ECM stiffness and the role of P-selectin-mediated ECM stiffness in insulinoma growth in Rip1-Tag2 mice. We found that collagen deposition increased with tumor progression in Rip1-Tag2 mice and Rip1-Tag2;P-sel^{-/-} mice. Our previous study also demonstrated that P-selectin deletion inhibit insulinoma growth in Rip1-Tag2 mice [9]. Furthermore, we investigated the relationship between P-selectin expression and insulinoma ECM stiffness using Rip1-Tag2 mice. Our findings indicated that P-selectin deletion decrease insulinoma ECM stiffness during every stage of insulinoma progression in Rip1-Tag2 mice, thereby inhibiting insulinoma growth in these mice. More importantly, we demonstrated that the LOX inhibitor BAPN

suppressed insulinoma growth in Rip1-Tag2 mice by reversing P-selectin-mediated increases in insulinoma ECM stiffness. We previously demonstrated that P-selectin promotes platelet accumulation in insulinomas in Rip1-Tag2 mice, thereby promoting insulinoma growth [3]. We also found that P-selectin-mediated platelet accumulation promotes increases in tissue stiffness mainly by increasing LOX expression.

Materials and Methods

Mice

P-selectin knockout (P-sel^{-/-}) mice and Rip1-Tag2 transgenic mice were purchased from the Jackson Laboratory (the Jackson Laboratory, Bar Harbor, ME, USA) and the National Cancer Institute (NCI, Washington, DC, USA). Male Rip1-Tag2 mice were crossed with female P-sel^{-/-} mice to establish Rip1-Tag2;P-sel^{-/-} mice. The genotypes of the Rip1-Tag2 mice and P-sel^{-/-} mice were identified as previously described [9]. The animals were housed under specific pathogen-free conditions, and all experiments were performed in accordance with institutional guidelines. The animal protocol was approved by the Medical Research Animal Ethics Committee of Guangdong Pharmaceutical University.

Immunofluorescence staining

Pancreases from Rip1-Tag2 mice and Rip1-Tag2;P-sel^{-/-} mice were fixed in 4% paraformaldehyde overnight and then embedded in optimum cutting temperature compound (OCT) and sectioned. For immunofluorescence staining, the sections were rehydrated in distilled water, blocked with 10% bovine serum albumin (BSA) and incubated with a primary antibody against collagen type I (Abcam, Cambridge, CB, UK) overnight at 4 °C. The next day, DyLight 488 or 555-conjugated anti-GPIIb/3 antibodies (Invitrogen, Carlsbad, CA, USA) were added to the sections, which were counterstained with 4'-6-diamidino-2-phenylindole (DAPI). For immunofluorescence staining quantitation, we randomly chose fields from at least five tumors per mouse (typically 10-15) in five to eight mice per group using a 40 × objective lens. The slides were collected, and the images were quantified using Image-Pro Plus software (IPP, version 6.0, Media Cybernetics).

Histopathological staining

Pancreases from Rip1-Tag2 mice and Rip1-Tag2;P-sel^{-/-} mice were fixed in 10% formalin and then embedded with paraffin and sectioned. The tissue sections (3 μm) were stained with H&E, Masson's trichrome and reticulin using reagents and kits from Maxim-Bio (Maxim-Bio, Fuzhou, Fujian,

China), as previously described [20]. For histopathological staining quantitation, we randomly chose fields from at least five tumors per mouse (typically 10-15) in five to eight mice per group using a 40× objective lens. The IPP software was used to quantify the images based on the histochemical staining results.

Immunohistological staining

After being deparaffinized and rehydrated, the tissue sections were incubated with anti-LOX (1:100 dilution, Abcam, Cambridge, CB, UK) or HIF- α antibodies overnight at 4 °C. Primary antibody binding to the tissues was detected using an HRP-conjugated secondary antibody. All the sections were stained with DAB and counterstained with hematoxylin. Images were taken in a 400 × field, quantified using IPP 6.0 software and assessed by two experimenters.

Hydroxyproline analysis

Hydroxyproline, a biochemical index of collagen deposition, was quantified as previously described [21]. Angiogenic islets and insulinomas from three mice were hydrolyzed in 6 N HCL overnight at 100 °C. After the tissues were neutralized and centrifuged, their corresponding cell supernatants were collected. To oxidize hydroxyproline, the supernatant was incubated with chloramine-T buffer at room temperature for 20 minutes. The p-dimethylaminobenzaldehyde was added to the samples, which were incubated for 20 minutes at 60 °C, and was then detected by a microplate reader at an absorbance of 575 nm. The results are expressed as

$\mu\text{g/g}$ wet tissue.

LOX inhibition *in vivo* by BAPN

Eight-week-old Rip1-Tag2 mice were intraperitoneally injected with the LOX inhibitor β -aminopropionitrile (BAPN, 100 mg/kg in 200 μL of PBS; Sigma-Aldrich, St Louis, MO, USA) or sterile PBS daily. Four weeks later, tumor numbers and volumes were measured as previously described [9]. Survival times were observed in another group of Rip1-Tag2 mice treated with BAPN or PBS.

Statistical analysis

For statistical analysis, a two-tailed Student's *t*-test was used to determine statistical significance. Data were analyzed using the GraphPad Prism 5 software package (GraphPad Software, CA), and charts were constructed using SigmaPlot software. *P* values < 0.05 and < 0.01 were considered significant and very significant, respectively.

Results

Collagen fiber deposition increases in conjunction with tumor progression in Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mice

After the pancreases of Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mice were isolated and sectioned, the sections were subjected to immunofluorescence staining for collagen type I. There were very few collagen fibers in the angiogenic islets of Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mice (Figure 1). However, fiber deposition was significantly increased in the tumors of both mice, especially in invasive carcinomas (Figure 1). These results show that fiber deposition in insulinomas can play an important role in tumor progression.

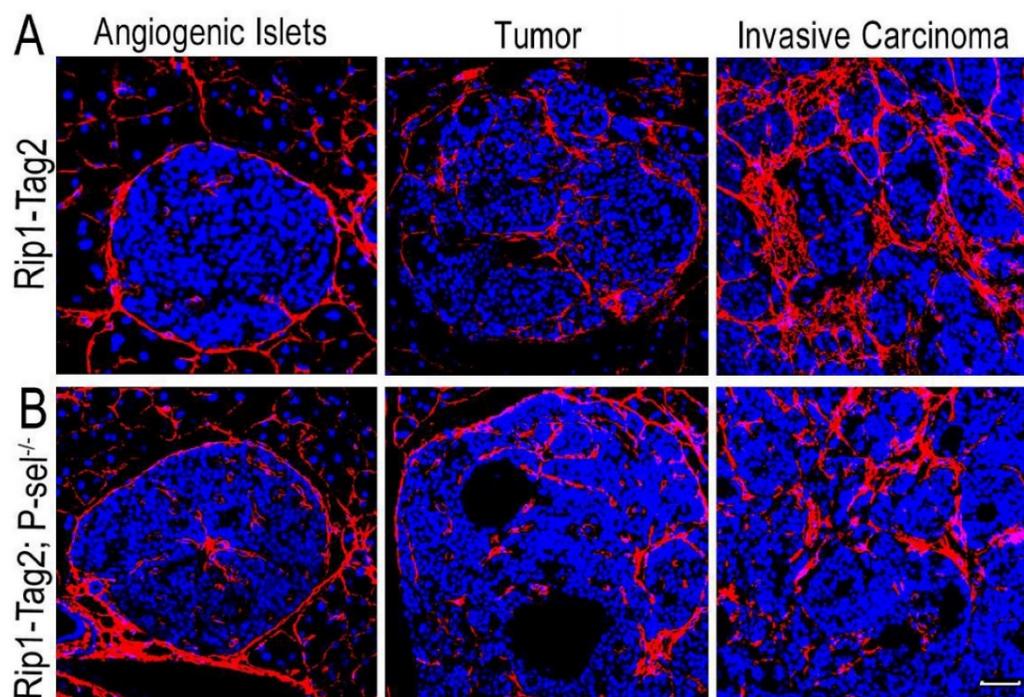


Figure 1. Collagen fiber deposition in Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mouse insulinomas. (A and B) Representative images of immunofluorescence staining of insulinoma sections from Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mice. The levels of collagen fiber deposition significantly increased with tumor progression in Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mice. Bar = 50 μm .

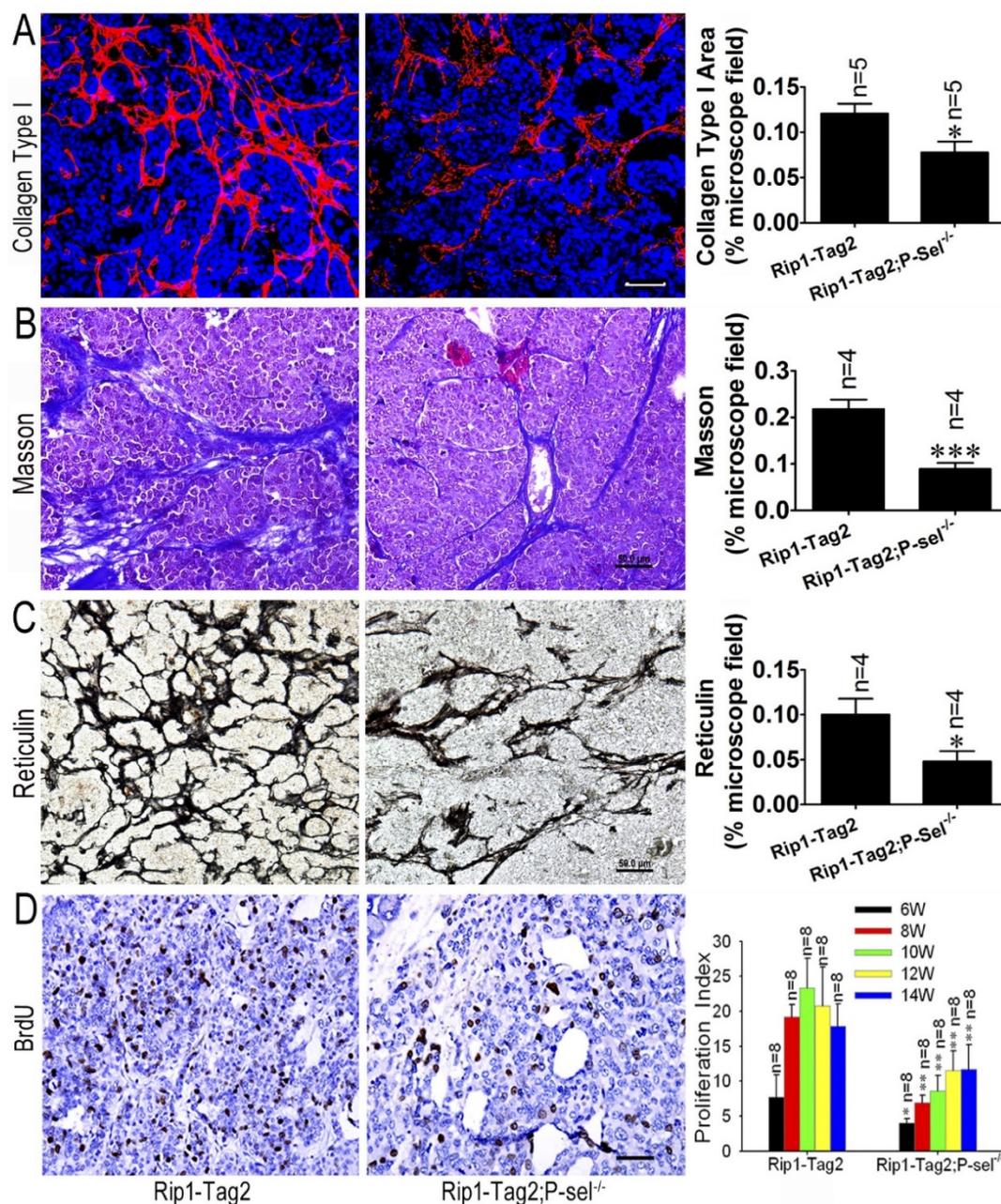


Figure 2. P-selectin deletion decreases tissue stiffness in Rip1-Tag2 mouse insulinomas. (A) Immunofluorescence staining showing collagen type I deposition in the insulinomas of 12-week-old Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mice. Rip1-Tag2;P-sel^{-/-} mice exhibited less collagen type I deposition than Rip1-Tag2 mice. (B) Representative images and quantitation of Masson's staining of insulinoma sections from 12-week-old Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mice. Collagen fiber deposition levels were determined by evaluating the blue areas as percentages of the total microscope area. (C) Representative images and quantitation of reticulin staining of insulinoma sections from 12-week-old Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mice. Rip1-Tag2;P-sel^{-/-} mice exhibited fewer reticular fibers than Rip1-Tag2 mice. (D) Insulinoma proliferative index. Proliferating cells were identified via immunohistochemical staining against BrdU antibodies. The numbers of positive cells per unit area were counted. At least ten sections from five mice were examined for statistical analysis. *, $P < 0.05$; **, $P < 0.01$. Bar = 50 μm .

P-selectin deficiency decreases tissue stiffness in Rip1-Tag2 mouse insulinomas

We previously reported that P-selectin deletion significantly inhibits insulinoma growth in Rip1-Tag2 mice [9] and that tumor tissue stiffness increases in conjunction with insulinoma progression (Figure 1). To investigate the effects of P-selectin on insulinoma tissue stiffness, we subjected Rip1-Tag2 and

Rip1-Tag2;P-sel^{-/-} mouse insulinomas to immunofluorescence, Masson's trichrome and reticulin staining for histological analysis. Immunofluorescence and Masson's trichrome staining showed that Rip1-Tag2;P-sel^{-/-} mouse insulinomas exhibited a significantly decrease in collagen density (Figure 2A and 2B). Accordingly, reticulin staining showed that reticular fiber density was significantly decreased in Rip1-Tag2;P-sel^{-/-}

mouse insulinomas compared to Rip1-Tag2 mouse insulinomas (Figure 2C). These results demonstrate that P-selectin deficiency decreases fiber deposition in Rip1-Tag2 mouse insulinomas and that fiber deposition is closely associated with tissue stiffness. Consistent with these findings, P-selectin deficiency suppressed tumor cell proliferation, as demonstrated via BrdU staining for proliferating cells (Figure 2D).

P-selectin deficiency decreases hydroxyproline levels in Rip1-Tag2 mouse insulinomas

Hydroxyproline, a component amino acid of collagen, is a parallel biochemical marker of fibrosis [22]. To investigate the role of P-selectin in insulinoma tissue stiffness further, we measured hydroxyproline levels in Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mouse insulinomas. Rip1-Tag2;P-sel^{-/-} mice exhibited significantly decreased-insulinoma hydroxyproline concentrations compared to control mice (Figure 3).

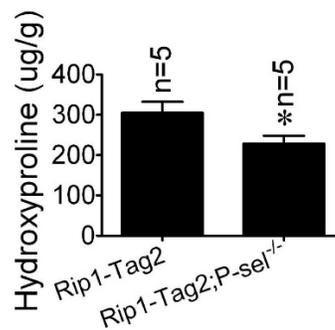


Figure 3. Hydroxyproline concentrations in the insulinomas of 12-week-old Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mice. Hydroxyproline levels were higher in the insulinomas of Rip1-Tag2 mice than in the insulinomas of Rip1-Tag2;P-sel^{-/-} mice. *, P < 0.05.

The LOX inhibitor BAPN inhibits tumor growth in Rip1-Tag2 mice

To determine how P-selectin affects insulinoma tissue stiffness in Rip1-Tag2 mice, we performed immunohistochemical staining to investigate LOX protein expression in the insulinomas of 12-week-old Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mice. LOX expression in Rip1-Tag2 mouse insulinomas was strong, while LOX expression in Rip1-Tag2;P-sel^{-/-} mouse insulinomas was very weak (Figure 4A). Our results also demonstrated that LOX expression levels were significantly lower in Rip1-Tag2;P-sel^{-/-} mouse insulinomas than in the Rip1-Tag2 mouse insulinomas (Figure 4B).

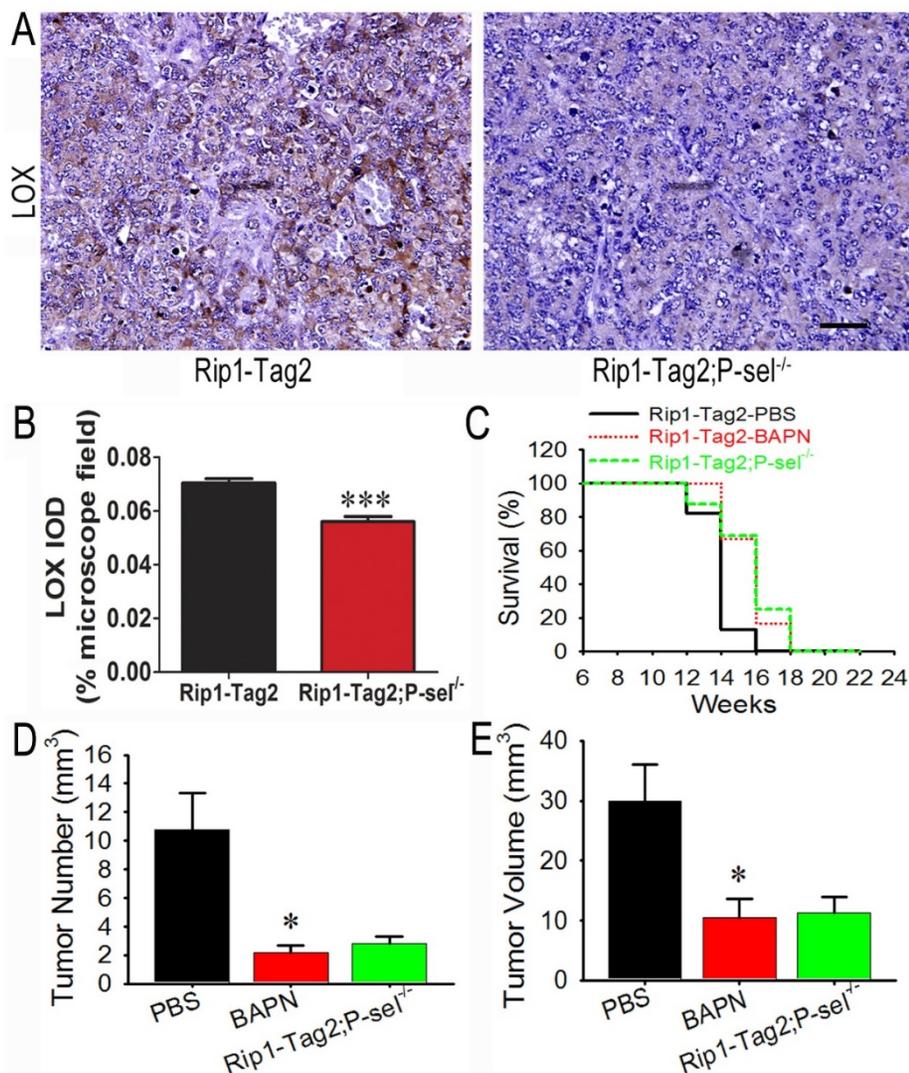


Figure 4. The effects of the LOX inhibitor BAPN on tumor growth in Rip1-Tag2 mice. (A and B) Immunohistochemical staining was performed to detect LOX expression in Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mouse insulinomas. Representative images and semi-quantitation of LOX expression in the insulinomas of 12-week-old Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mice. (C) The survival times of BAPN-treated Rip1-Tag2;P-sel^{-/-} mice and Rip1-Tag2 mice were significantly prolonged compared to those of PBS-treated Rip1-Tag2 mice. (D and E) Tumor volumes (D) and numbers (E) were significantly decreased after BAPN treatment and P-selectin deletion. *, P < 0.05; ***, P < 0.001. Bar = 50 μm.

Because LOX expression was down-regulated in Rip1-Tag2;P-sel^{-/-} mouse insulinomas, the biological effects of LOX were investigated in the setting of tumor progression in Rip1-Tag2 mice. BAPN, a well-recognized inhibitor of LOX activity [23], was administered to Rip1-Tag2 mice. The survival rates of BAPN-treated Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mice were significantly higher than those of normal water-treated Rip1-Tag2 mice. Furthermore, there was no significant difference in survival time between BAPN-treated Rip1-Tag2 mice and Rip1-Tag2;P-sel^{-/-} mice (Figure 4C). Rip1-Tag2;P-sel^{-/-} mice and BAPN-treated Rip1-Tag2 mice also exhibited fewer and smaller tumors than normal water-treated Rip1-Tag2 mice (Figure 4D and 4E). There were no differences in tumor numbers or volumes between BAPN-treated Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mice (Figure 4D and 4E). These results show that P-selectin deletion inhibits tumor progression in Rip1-Tag2 mice

mainly by activating LOX.

The LOX inhibitor BAPN decreases tissue stiffness in Rip1-Tag2 mouse insulinomas

To investigate the effects of the LOX inhibitor BAPN on insulinoma tissue stiffness, Rip1-Tag2 mouse insulinomas treated with BAPN and PBS were subjected to immunofluorescence, Masson's trichrome and reticulin staining for histological analysis. The immunofluorescence and Masson's trichrome staining results showed that collagen density was significantly decreased in Rip1-Tag2 mouse insulinomas treated with BAPN (Figure 5A and 5B). Furthermore, reticulin staining demonstrated that reticular fiber density was significantly decreased in Rip1-Tag2 mouse insulinomas treated with BAPN compared to control mice (Figure 5C). To determine the role of P-selectin in insulinoma tissue stiffness, we measured hydroxyproline levels in Rip1-Tag2 mouse insulinomas treated with BAPN

and PBS. Rip1-Tag2 mice treated with BAPN exhibited significantly decreased insulinoma hydroxyproline levels compared to control mice (Figure 5D). These results show that BAPN inhibits fiber deposition in Rip1-Tag2 mouse insulinomas and that fiber deposition is closely associated with tissue stiffness.

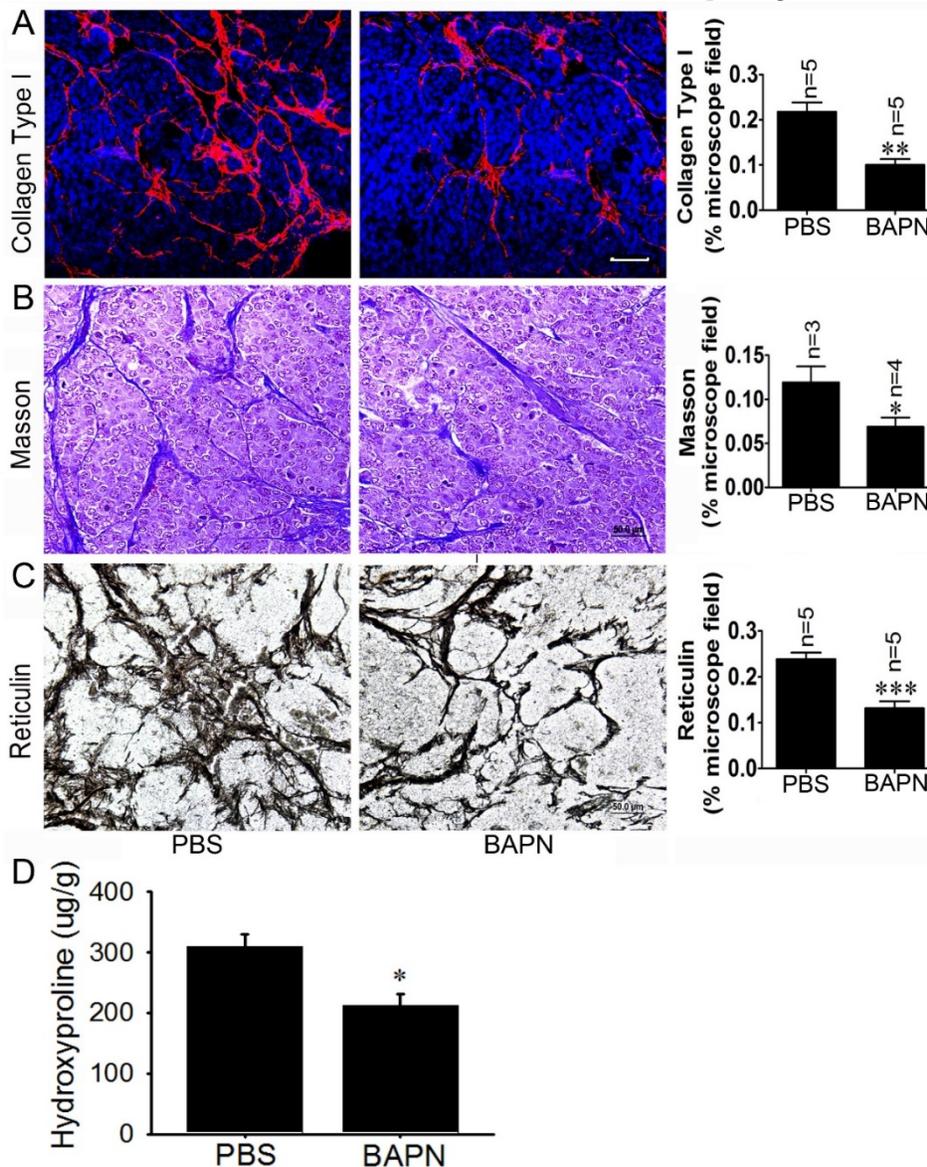


Figure 5. BAPN decreases tissue stiffness in Rip1-Tag2 mouse insulinomas. (A) The insulinoma sections of Rip1-Tag2 mice treated with normal water or BAPN water were subjected to immunofluorescence staining for collagen type I. Representative images and quantitation of immunofluorescence staining in the insulinoma sections of Rip1-Tag2 mice treated with BAPN or PBS. Rip1-Tag2 mice treated with BAPN exhibited much less type I collagen deposition than control mice. (B) Representative images and quantitation of Masson's staining of insulinoma sections from Rip1-Tag2 mice treated with BAPN or PBS. Collagen fiber deposition levels were determined by evaluating the blue areas as percentages of the total microscope area. (C) Representative images and quantitation of reticulin staining of insulinoma sections from Rip1-Tag2 mice treated with BAPN or PBS. Rip1-Tag2 mice treated with BAPN exhibited less reticular fiber deposition than control mice. (D) Hydroxyproline concentrations were detected in the insulinomas of 12-week-old Rip1-Tag2 mice treated with BAPN and PBS. Hydroxyproline levels were lower in the insulinomas of Rip1-Tag2 mice treated with BAPN than in the insulinomas of control mice. * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$. Bar = 50 μ m.

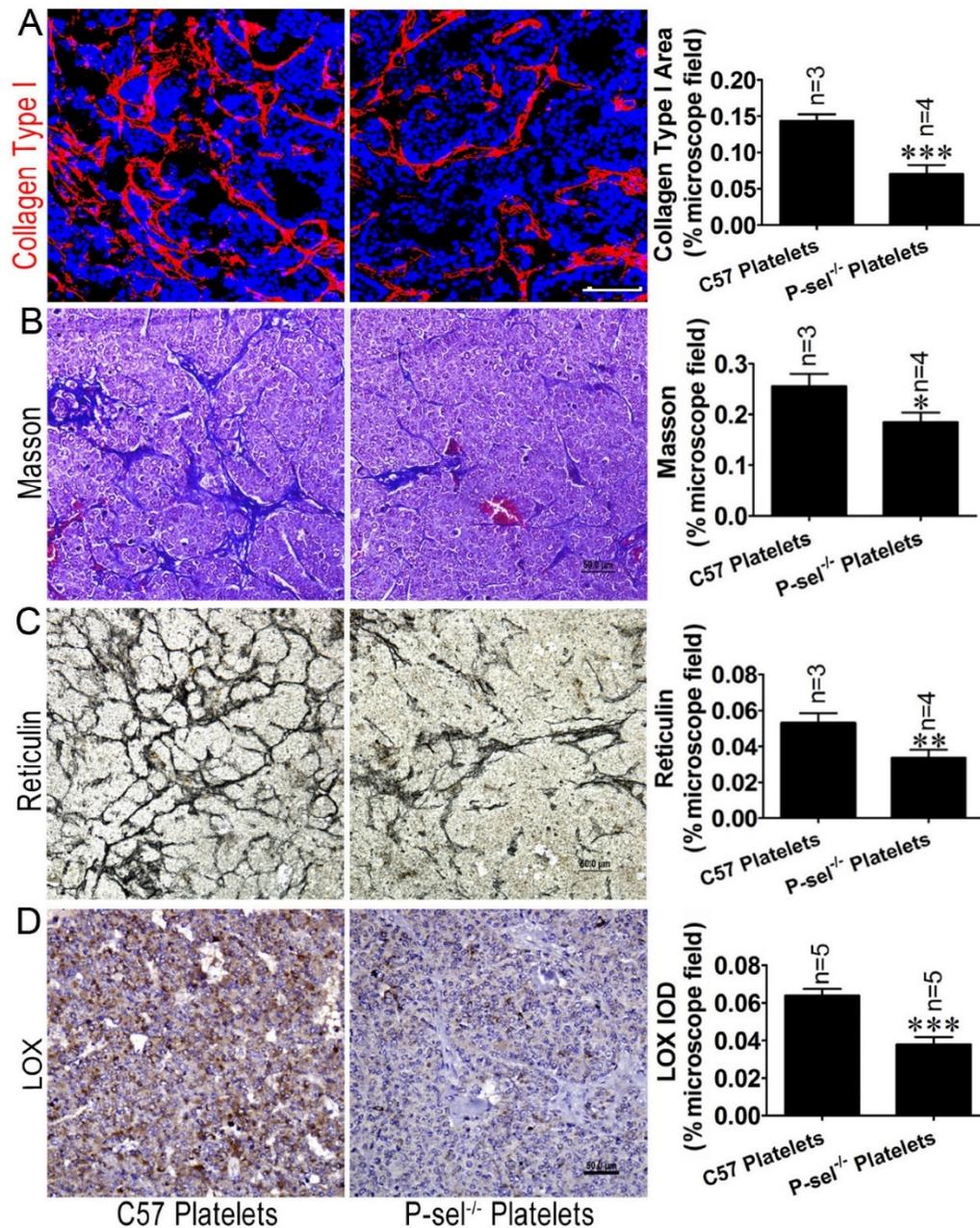


Figure 6. P-selectin-mediated platelet adhesion increases tissue stiffness by increasing LOX expression. (A) The insulinoma sections of Rip1-Tag2;P-sel^{-/-} mice treated with C57 platelets or P-sel^{-/-} platelets were subjected to immunofluorescence staining for collagen type I. Representative images and quantitation of immunofluorescent staining of insulinoma sections from Rip1-Tag2;P-sel^{-/-} mice treated with C57 platelets or P-sel^{-/-} platelets. Rip1-Tag2;P-sel^{-/-} mice treated with P-sel^{-/-} platelets exhibited lower levels of collagen type I deposition than control mice. (B) Representative images and quantitation of Masson's staining of insulinoma sections from Rip1-Tag2;P-sel^{-/-} mice treated with C57 platelets or P-sel^{-/-} platelets. Collagen fiber deposition levels were determined by evaluating the blue areas as percentages of the total microscope area. (C) Representative images and quantitation of reticulin staining of insulinoma sections from Rip1-Tag2;P-sel^{-/-} mice treated with C57 platelets or P-sel^{-/-} platelets. Rip1-Tag2;P-sel^{-/-} mice treated with P-sel^{-/-} platelets exhibited less reticular fiber deposition than control mice. (D) Immunohistochemical staining was performed to detect LOX expression in the insulinomas of Rip1-Tag2;P-sel^{-/-} mice treated with C57 platelets or P-sel^{-/-} platelets. Representative image and semi-quantitation of LOX expression in the insulinomas of 12-week-old Rip1-Tag2;P-sel^{-/-} mice treated with C57 platelets or P-sel^{-/-} platelets. *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$. Bar = 50 μ m.

P-selectin-mediated platelet adhesion increases tissue stiffness by increasing LOX expression

To investigate the effects of P-selectin-mediated platelet adhesion on insulinoma tissue stiffness, Rip1-Tag2;P-sel^{-/-} mice were treated with C57 and P-sel^{-/-} platelets. Their insulinomas were subjected to immunofluorescence, Masson's trichrome and

reticulin staining for histological analysis. The immunofluorescence and Masson's trichrome staining results demonstrated that collagen density was significantly decreased in the insulinomas of Rip1-Tag2 mice treated with P-sel^{-/-} platelets (Figure 6A and 6B). Reticulin staining showed that reticular fiber density was significantly decreased in the insulinomas of Rip1-Tag2 mice treated with P-sel^{-/-} platelets compared to control mice (Figure 6C).

Given that P-selectin-mediated platelet adhesion increases tissue stiffness, we elected to evaluate how P-selectin-mediated platelet adhesion affects tumor stiffness. We detected LOX protein expression in the insulinomas of Rip1-Tag2;P-sel^{-/-} mice treated with C57 platelets and P-sel^{-/-} platelets via immunohistochemical staining. LOX expression was increased in the insulinomas of Rip1-Tag2;P-sel^{-/-} mice treated with C57 platelets, while LOX expression was weak in the insulinomas of Rip1-Tag2;P-sel^{-/-} mice treated with P-sel^{-/-} platelets (Figure 6D). These results show that P-selectin-mediated platelet adhesion significantly affects fiber deposition, which is closely associated with tissue stiffness.

Discussion

It was previously reported by us that P-selectin deficiency can inhibit insulinoma growth in Rip1-Tag2 mice by suppressing platelet accumulation in tumor tissues [9]. Here, we showed that a correlation exists among ECM mechanical properties, P-selectin and insulinoma growth in Rip1-Tag2 mice. Our findings demonstrate that tumor progression in Rip1-Tag2 mice corresponds with significant increases in collagen deposition and linearization, which increase ECM stiffness. Importantly, P-selectin deletion significantly inhibits collagen deposition and linearization, thereby suppressing insulinoma growth in Rip1-Tag2 mice. The LOX inhibitor BAPN significantly suppresses insulinoma growth in Rip1-Tag2 mice, but not Rip1-Tag2;P-sel^{-/-} mice, in conjunction with changes in ECM remodeling and stiffening. These results demonstrate that a correlation exists between tumor aggression and P-selectin-mediated increases in tumor stiffness in Rip1-Tag2 mice and that tumor stiffness is determined by collagen cross-links, especially the total amount of collagen cross-links.

Previous investigations have demonstrated that tissue stiffness contributes to tumor progression. P-selectin-mediated collagen deposition significantly increases ECM stiffness, which thereafter affects β -cell proliferation in Rip1-Tag2 mice (Figure 2), thereby promoting insulinoma growth in these mice. Our previous results indicated that P-selectin (an important member of the selectin family) deficiency suppresses melanoma metastasis and insulinoma growth by preventing platelet adhesion [7,9]. Furthermore, increases in tumor stiffness can exert diverse effects on tumor cell proliferation, differentiation, and migration and hence significantly modify tumor progression [24-27].

Increases in insulinoma ECM stiffness play an important role in insulinoma growth in Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mice. Increases in ECM

stiffness are, at least in part, caused by increases in the total amount of collagen crosslinks [28]. Collagen crosslinking is induced predominantly by LOX and LOX-like (LOXLs) enzymes, which are synthesized by stromal cells during the early stages of cancer and synthesized by both stromal cells and cancer cells in response to hypoxia [29, 30]. Furthermore, LOX is frequently highly expressed in many cancers [28], and we found that LOX expression levels were high in Rip1-Tag2 mouse insulinomas. P-selectin deletion significantly decreased HIF-1 α and LOX expression in Rip1-Tag2 mouse insulinomas. In addition, we treated Rip1-Tag2 and Rip1-Tag2;P-sel^{-/-} mice with BAPN, a LOX inhibitor, to determine the effects of LOX on insulinomas. BAPN treatment prevented collagen crosslinking and suppressed ECM stiffness in Rip1-Tag2 mice, thereby reducing tumor volumes and prolonging survival times in Rip1-Tag2 mice. However, in Rip1-Tag2;P-sel^{-/-} mice, BAPN treatment had no effect on tumor growth. Using a breast cancer mouse model, Levental KR *et al* also demonstrated that LOX inhibitor prevents tissue stiffening, thereby preventing breast cancer progression [28]. Our results indicate that P-selectin mediated-platelet adhesion induces LOX synthesis, which leads to collagen crosslinking.

In this study, we found that P-selectin mediated-platelet adhesion increases collagen deposition in Rip1-Tag2 mouse insulinomas by promoting LOX expression and thus increasing ECM stiffness. Increases in ECM stiffness promote insulinoma growth in Rip1-Tag2 mice. These findings indicate that ECM stiffness is associated with increased-insulinoma aggressiveness, which is mediated by P-selectin. Therefore, P-selectin may be an effective therapeutic targeting for treating human insulinoma.

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Competing Interests

The authors have declared that no competing

interest exists.

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