

Research Paper

Prediction of CircRNA-Disease Associations Using KATZ Model Based on Heterogeneous Networks

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Abstract

Circular RNAs (circRNAs) are a large group of endogenous non-coding RNAs which are key members of gene regulatory processes. Those circRNAs in human play significant roles in health and diseases. Owing to the characteristics of their universality, specificity and stability, circRNAs are becoming an ideal class of biomarkers for disease diagnosis, treatment and prognosis. Identification of the relationships between circRNAs and diseases can help understand the complex disease mechanism. However, traditional experiments are costly and time-consuming, and little computational models have been developed to predict novel circRNA-disease associations. In this study, a heterogeneous network was constructed by employing the circRNA expression profiles, disease phenotype similarity and Gaussian interaction profile kernel similarity. Then, we developed a computational model of KATZ measures for human circRNA-disease association prediction (KATZHCDA). The leave-one-out cross validation (LOOCV) and 5-fold cross validation were implemented to investigate the effects of these four types of similarity measures. As a result, KATZHCDA model yields the AUCs of 0.8469 and 0.7936 \pm 0.0065 in LOOCV and 5-fold cross validation, respectively. Furthermore, we analyze the candidate association between hsa_circ_0006054 and colorectal cancer, and results showed that hsa_circ_0006054 may function as miRNA sponge in the carcinogenesis of colorectal cancer. Overall, it is anticipated that our proposed model could become an effective resource for clinical experimental guidance.

Key words: CircRNA-disease association, similarity measure, KATZ model

Introduction

Circular RNAs (circRNAs) are a class of endogenous non-coding RNAs with a covalently closed continuous loop that lacks 5'-3' polarity structure. The first circRNA was found in 1976 in an electron microscopy-based study of RNA viruses [1]. Nonetheless, due to the structural specificity, unknown function and low abundance of circRNAs, they were initially assumed to be artefacts or mis-splicing products and did not attract much attention [2]. In recent years, thanks to the development of high-throughput sequencing technology and other techniques [3], an increasing number of circRNAs have been identified in thousands of living organisms including archaea, plants and animals [4-7]. The biogenesis mechanisms

of circRNAs mainly include intron pairing-driven [8-10], lariat-driven [11], and RNA-binding protein-driven circularization [12]. In addition, circRNAs can regulate gene expression at transcriptional or post-transcriptional levels by titrating microRNAs (miRNAs), regulating transcription and interfering with splicing [13, 14]. CircRNAs play a critical role in biological processes including transcription, mRNA splicing, RNA decay and translation [15]. Therefore, the misregulation of circRNAs may cause abnormal cellular functions and growth defects and are involved in diseases.

Several circRNAs have been reported to be associated with human diseases, such as glioma tumorigenesis, osteosarcoma, colorectal cancer and so

on. Circ-FBXW7 is reduced in glioblastoma clinical samples compared with their paired tumor-adjacent tissues, and circ-FBXW7 expression is also positively associated with glioblastoma patients overall survival [16]. circPVT1 is significantly up-regulated in the osteosarcoma tissues, and circPVT1 may be a biomarker for the diagnosis of osteosarcoma [17]. hsa_circ_0081001 is reported correlated with poor prognosis, and its expression level may dynamically monitor the condition changes of osteosarcoma [18]. Has_circ_001569 is highly expressed in cell proliferation and invasion of colorectal cancer, compared with non-cancerous samples [19]. Has_circ_0054633 is found to be upregulated in the peripheral blood of patients with type 2 diabetes, and it could be used as a candidate biomarker for the diagnosis for pre-diabetes and type 2 diabetes [20]. Upregulation or downregulation of circRNAs in specific disease tissues compared with normal samples could indicate their diagnostic potential to be new biomarkers. However, disease-related circRNAs are generally detected by analyzing RNA-seq or microarray data, then low throughput biological experiments are used to validate the abnormal expression such as RT-PCR, northern blot and so on. Furthermore, these experiments are expensive and time-consuming.

The CircR2Disease database is constructed, which collects experimentally validated circRNA-disease associations from published literatures [21]. Detecting the potential circRNA-disease associations is significant to understand the complex disease mechanism. To date, little efforts have made to develop the computational methods for circRNA-disease association prediction. Network-based models and machine learning methods have been widely applied to the prediction of bipartite biological associations. For example, KATZ measure is a network-based method to calculate the similarity between nodes in a heterogeneous network [22]. The KATZ measure has been successfully applied to social networks [23], and predict gene-disease associations [24], miRNA-disease associations [25, 26], lncRNA-disease associations [27] and microbe-disease associations [22]. Here, we propose a KATZ-based method for human circRNA-disease association prediction (KATZHCD). In this study, the heterogeneous network is constructed by integrating known circRNA-disease associations, circRNA similarity network and disease similarity networks, where circRNA similarity is calculated from the circRNA expression and Gaussian interaction profiles (GIP) kernel while disease similarity is calculated from the disease phenotype and GIP kernel. By integrating the

heterogeneous network and KATZ measure, the KATZHCD model is developed to predict novel circRNA-disease associations.

To evaluate the performance of KATZHCD model, the leave-one-out cross validation (LOOCV) and 5-fold cross validation (5-fold CV) are implemented. In addition, the prediction performance of different kinds of similarity measures are investigated. As a result, the KATZHCD model achieved the best performance with AUCs of 0.8469 and 0.7936 \pm 0.0065 based on LOOCV and 5-fold cross validation, respectively. Finally, we analyze the candidate association between hsa_circ_0006054 and colorectal cancer, and results shown that hsa_circ_0006054 may be targeted by six miRNAs including hsa-miR-153, hsa-miR-194, hsa-miR-217, hsa-miR-431, hsa-miR-503, hsa-miR-646, and function as hsa-miR-217 sponge in the carcinogenesis of colorectal cancer. Overall, our framework is an effective resource for clinical experimental guidance.

Materials and Methods

Human circRNA-disease associations

The known circRNA-disease associations were derived from the CircR2Disease, which is a manually curated database for experimentally validated circRNA-disease associations. Here, we extract the circRNA-disease associations of human, in which the circRNAs own common circRNA ID with circBase and the diseases listed in the Online Mendelian Inheritance in Man (OMIM) [28, 29]. The gold standard dataset contains 312 distinct circRNA-disease associations involving 275 circRNAs and 36 diseases (Figure 1). The degree distributions of circRNAs and diseases in the bipartite of gold standard circRNA-disease association network are illustrated in Figure 2. The adjacency matrix A of gold standard circRNA-disease associations is constructed, and $A(i,j)$ is equal to 1 when there is a known association between circRNA $c(i)$ and disease $d(j)$, otherwise 0.

CircRNA expression profile similarity

To calculate the similarity among circRNAs, we downloaded the expression profiles of circRNAs from exoRBase database (<http://www.exorbase.org>) [30]. After converting the circRNA ID of CircR2Disease into exoRBase, we obtained expression profile data for 149 circRNAs. Each record of circRNA expression profiles has 90 dimensions representing the expression levels based on RNA-seq data spanning normal individuals and patients with various diseases. A quick overview of the expression levels of 149 circRNAs in different kinds of samples is also shown (Figure 3). Based on the circRNA expression

profiles, the Pearson correlation coefficient was used for similarity measurement, and a pair of circRNAs with a higher correlation score is considered to be more similar. Given expression profiles of two circRNAs denoted by $X=(x_1, \dots, x_n)$ and $Y=(y_1, \dots, y_n)$, then the Pearson correlation coefficient can be calculated as follows:

$$SC_{EP}(X, Y) = \frac{\sum_{i=1}^N (x_i - \bar{x})(y_i - \bar{y})}{\sqrt{\sum_{i=1}^N (x_i - \bar{x})^2 \sum_{i=1}^N (y_i - \bar{y})^2}} \quad (1)$$

where x_i and y_i represent the expression values of two circRNAs in different human tissues or cell lines respectively, N denotes the number of components in the circRNA expression profiles.

Disease phenotype similarity

The disease phenotype similarity was downloaded from MimMiner [31], which measure disease similarity by computing similarity between Mesh terms appearing in the phenotype descriptions of diseases from OMIM database [32]. Here, let SD_{DP} represent the disease phenotype similarity matrix in gold standard dataset, in which each entry is the phenotype similarity score between diseases $d(i)$ and $d(j)$.

GIP kernel similarity

Based on assumption that similar circRNAs exhibit a similar interaction or non-interaction with the diseases and vice versa, the GIP kernel is used to measure the topology similarity for circRNAs and diseases as follows [33]:

$$SC_{cGIP}(c(i), c(j)) = \exp(-\gamma_c \|c(i) - c(j)\|^2)$$

$$\gamma_c = 1 / \left(\frac{1}{n_c} \sum_{i=1}^{n_c} \|c(i)\|^2 \right) \quad (2)$$

$$SD_{dGIP}(d(i), d(j)) = \exp(-\gamma_d \|d(i) - d(j)\|^2)$$

$$\gamma_d = 1 / \left(\frac{1}{n_d} \sum_{i=1}^{n_d} \|d(i)\|^2 \right) \quad (3)$$

where $c(i)$ denotes the binary vector encoding the presence or absence with each disease, i.e., the i -th row of the adjacency matrix A . n_c denotes the number of circRNAs. Similarly, $d(i)$ denotes the binary vector encoding the presence or absence with each circRNAs according to the i -th column of the adjacency matrix A . n_d denotes the number of diseases.

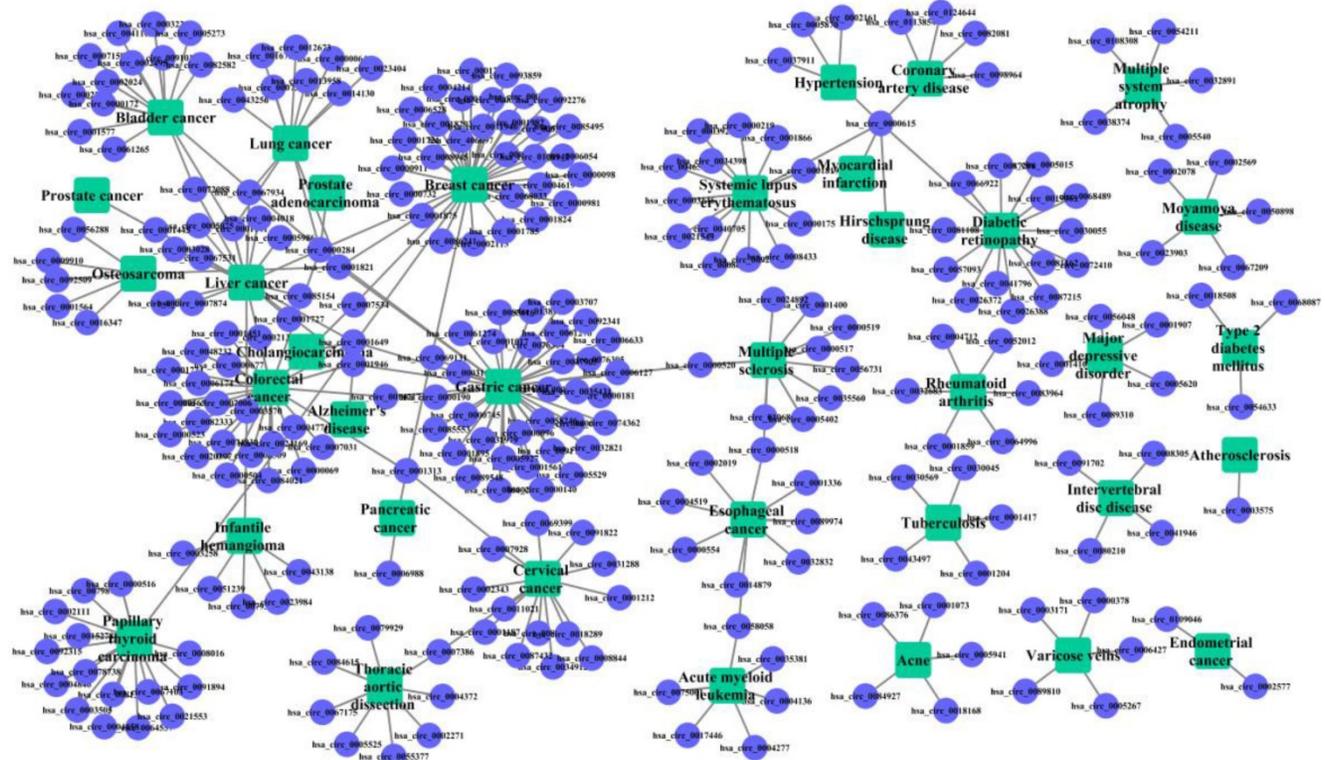


Figure 1. Bipartite graph of the circRNA-disease associations. The rectangles represent the diseases, and the circles represent the circRNAs. An edge corresponds to the gold standard circRNA-disease associations.

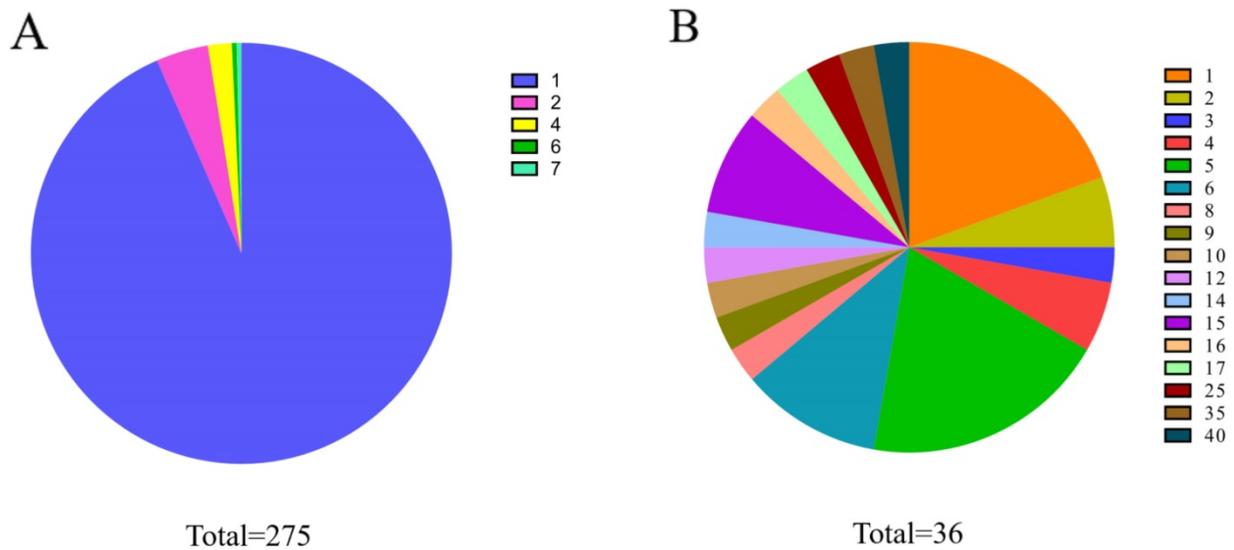


Figure 2. Pie graph of degree distribution for circRNAs or diseases. (A) Degree proportion of circRNAs. (B) Degree proportion of diseases.

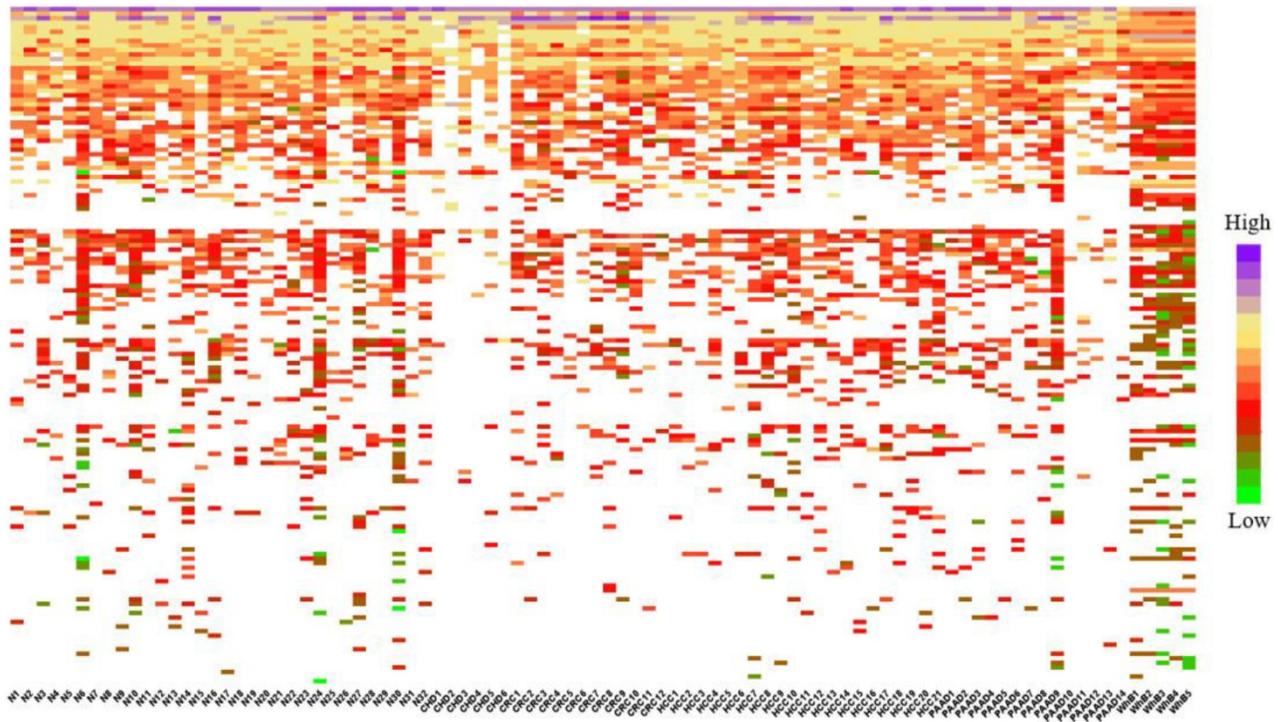


Figure 3. Heatmap of circRNAs in different samples.

Integrated similarity for circRNAs and diseases

The circRNA expression profile similarity and disease phenotype similarity cannot cover all the circRNAs and diseases in the gold standard dataset. Therefore, the GIP kernel similarity for circRNAs and diseases were integrated based on known circRNA-disease associations. Finally, the circRNA similarity matrix was calculated by integrating circRNA expression profile similarity (SC_EP) and

GIP kernel similarity (SC_cGIP) for circRNAs as in Equation (4), and the disease similarity matrix was computed by integrating the disease phenotype similarity (SD_DP) and GIP kernel similarity (SD_dGIP) for diseases as in Equation (5).

$$SC = \alpha SC_EP + (1 - \alpha) SC_cGIP \quad \alpha \in [0, 1] \quad (4)$$

$$SD = \beta SD_DP + (1 - \beta) SD_dGIP \quad \beta \in [0, 1] \quad (5)$$

KATZHCD

KATZ measure is a graph-based method to calculate the similarity between nodes in a heterogeneous network [23]. The number of walks between nodes and walk length are considered to be two effective similarity metrics in the network. So, counting the number of walks of different lengths between circRNA node and disease node can be used to measure the circRNA-disease associations. Here, heterogeneous network is constructed by integrating the circRNA similarity network, disease similarity network and the known circRNA-disease associations (Figure 4). However, with the variation of parameters α and β , the heterogeneous network is also changed. Anyway, the adjacency matrix of the heterogeneous network could be denoted as follows:

$$A^* = \begin{bmatrix} SC & A \\ A^T & SD \end{bmatrix} \quad (6)$$

where SC represents the adjacency matrix of circRNA similarity network, SD represents the disease similarity network, A represents the adjacency matrix of known circRNA-disease associations.

Based on the known circRNA-disease association adjacency matrix, the number of walks of length l between circRNA c_i and disease d_j can be

calculated by computing $A^l(i,j)$. Furthermore, heterogeneous network A^* can be used for the prediction of new circRNA-disease associations. To get a single measurement of each circRNA-disease pair, all walks of different lengths are integrated. Because different lengths of walks have different contribution, a nonnegative parameter γ is introduced to dampen the contribution of longer walks. Therefore, the potential associations between circRNA c_i and disease d_j could be calculated by:

$$S(c(i),d(j)) = \sum_{l=1}^k \gamma^l A^{*l}(i,j) \quad (7)$$

All walks of network can be shown as:

$$S = \sum_{l \geq 1} \gamma^l A^{*l} = (I - \gamma A^*)^{-1} - I \quad (8)$$

where S represents the similarities of all the circRNA-disease pairs. However, it is not necessary to consider all the path lengths. Path of longer lengths in sparse circRNA-disease network may be insignificant or meaningless. Therefore, we set k to be 2, 3 and 4, and the final prediction scores for potential associations are calculated.

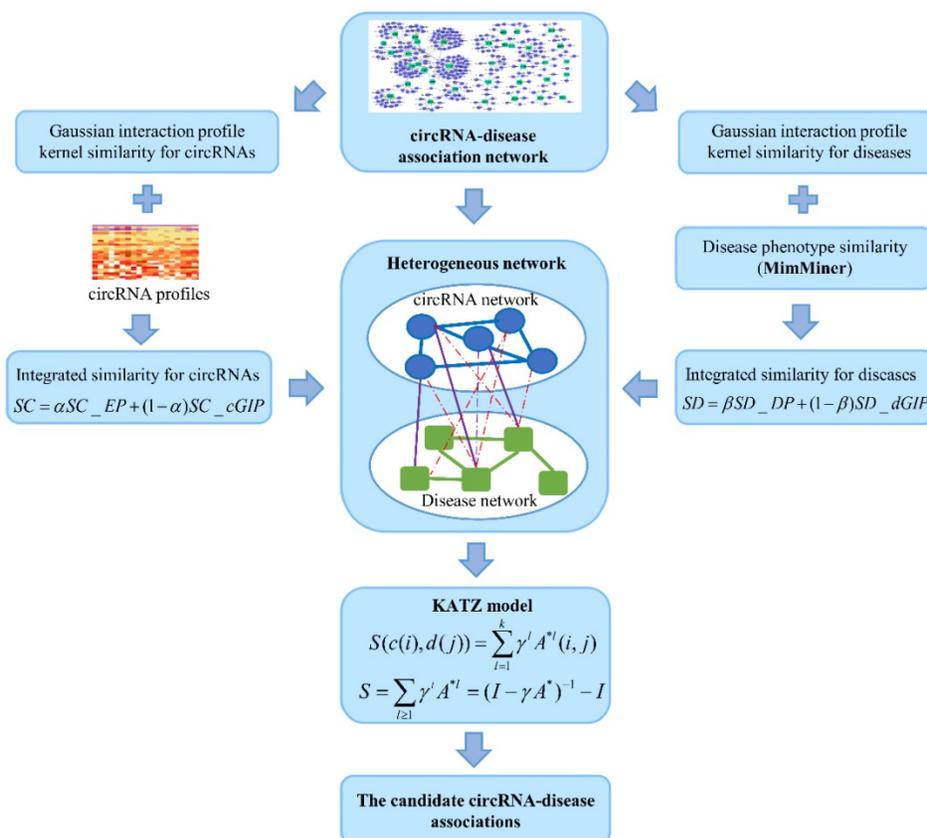


Figure 4. The flowchart of the KATZHCD model.

Experiments and Results

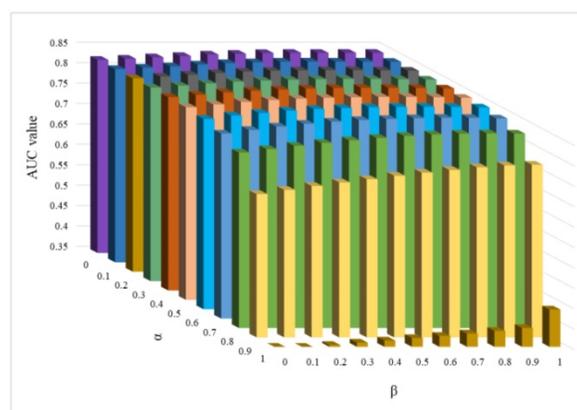
Performance evaluation

The LOOCV and 5-fold CV are implemented on the gold standard human circRNA-disease associations in CircR2Disease database to evaluate the performance of KATZHCD model. In each round of LOOCV, a known circRNA-disease pair is removed in turn as test sample and the other known circRNA-disease pairs are used as training samples. Then, the receiver operating characteristic (ROC) curve is used to evaluate the prediction performance of KATZHCD model, which plots the true positive rate (TPR, sensitivity) against false positive rate (FPR, 1-specificity) over various score thresholds. Here, sensitivity (SEN) refers to the percentage of positive cases that are correctly identified, and specificity (SPE) means the percentage of negative cases that are correctly predicted. In this way, the area under the curve (AUC) is calculated from the corresponding area under ROC curve. In addition, in the framework of 5-fold CV, the gold standard human circRNA-disease associations are randomly divided into five equal size subjects. One of them is retained as testing samples while the remaining datasets are regarded as training samples in turns. This process randomly repeated 100 times, and the average AUC values and standard deviations were obtained.

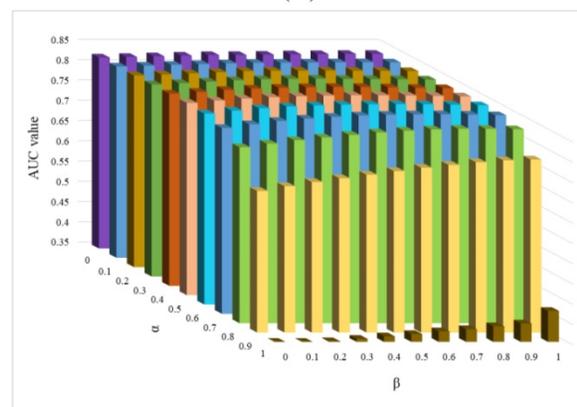
Effect of parameters

There are four parameters in the KATZHCD model, including the a and β , the number of walks k and nonnegative parameter γ . When $a(\beta)=1$, only circRNA expression profile similarity matrix (disease phenotype similarity) was considered. While when $a(\beta)=0$, only topology similarity of known circRNA-disease network was used. To evaluate the performances with different types of similarity matrices, various values were set for a and β . Here, when the values of a and β take 0, 0.1, ..., 0.9, 1, the values of LOOCV AUC were calculated. In addition, based on the suggestion by Zou et al., γ is selected on the basis of $\gamma < 1/||A||^2$ [26]. Therefore, γ is set as 0.01 by following precious studies [22]. Furthermore, k is another parameter that have a very large influence on the prediction results for circRNA-disease associations. Because of the circRNA-disease network is very sparse, so too long walks may be meaningless. Here, we implement a series of comparison experiments to evaluate the performance of KATZHCD by adjusting the parameter k . LOOCV was implemented when k was set as 2, 3, 4 (Figure 5). The highest LOOCV AUC values were 0.8469 ($a=0.5$, $\beta=0.8$), 0.8443 ($a=0.6$, $\beta=0.7$), and 0.8438 ($a=0.6$, $\beta=0.7$) when k was set from 2 to 4, respectively. As a result,

we found the LOOCV AUC value of this model are highest when a , β and k was set as 0.5, 0.8 and 2, respectively. Because little gap between these highest LOOCV AUC values, 5-fold CV was also implemented (Table 1). Results showed that the average AUC values keep an increasing trend when k increased from 2 to 4. Based on the opposite trend of LOOCV and 5-fold, we infer it may be caused by the 5-fold CV each round was removed too many effective edges of the heterogeneous network and the longer walks maybe make up this shortcoming. Therefore, we set a as 0.5 and β as 0.8 in the following analysis.



(A)



(B)

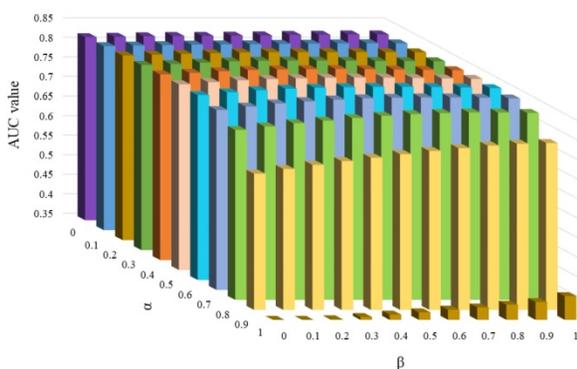


Figure 5. Effect of parameter a and β when k was set as 2, 3 and 4.

Table 1. 5-fold CV experimental results of setting parameter *k*.

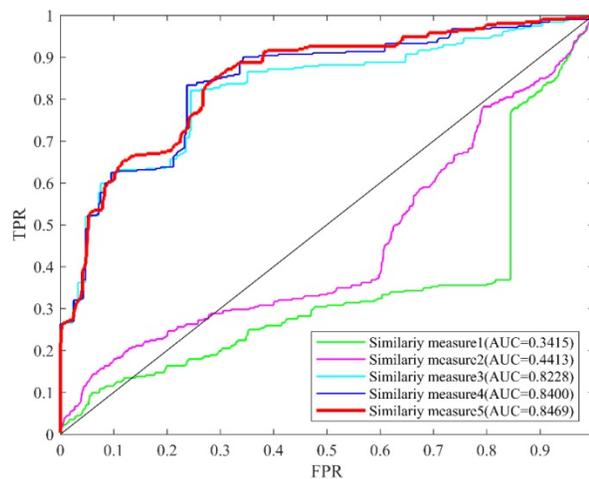
5-fold CV (Average AUC)	<i>k</i> = 2	<i>k</i> = 3	<i>k</i> = 4
$\alpha=0.5, \beta=0.8$	0.7616+/-0.0062	0.7764+/-0.0066	0.7833+/-0.0075
$\alpha=0.6, \beta=0.7$	0.7561+/-0.0067	0.7710+/-0.0070	0.7763+/-0.0065

Comparison with different similarity measures

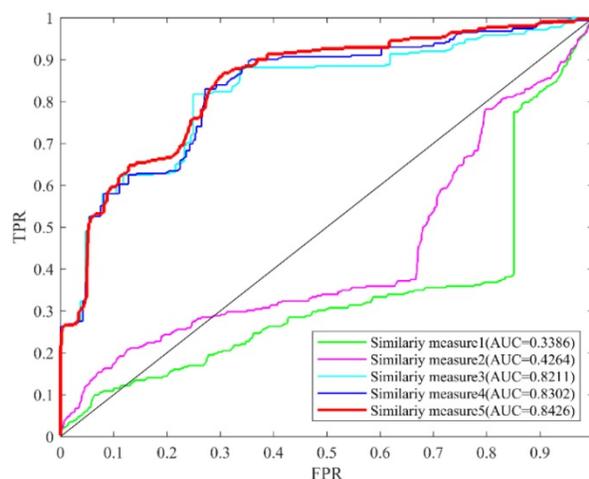
In this study, different kinds of information were used to measure the similarity of circRNAs and diseases. These types of similarity measures conclude circRNA expression profile similarity, disease phenotype similarity, GIP kernel similarity. In order to evaluate the usefulness of these similarity measures, LOOCV and 5-fold were implemented to compare the experimental results (Table 2, Figure 6). From the LOOCV results, we obtained the highest AUC of 0.8469 by integrating the circRNA expression profile similarity, disease phenotype similarity, GIP kernel similarity for circRNAs and diseases. However, the highest average AUC of 0.7936 by integrating disease phenotype similarity and GIP kernel similarity for circRNAs in 5-fold CV. In both LOOCV and 5-fold CV, the experiment results showed that GIP kernel similarity for circRNAs and diseases are very effective for the prediction of circRNA-disease associations.

Candidate circRNA for the specific disease

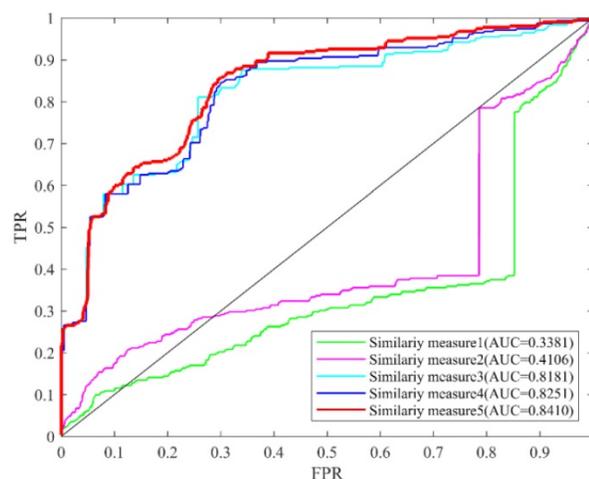
To illustrate the application of KATZHCD model for the disease-associated circRNA prediction, the miRNA-circRNA and miRNA-disease associations are introduced. It has reported that circRNAs can act as competing endogenous RNAs (ceRNAs) to sequester miRNAs and prevent their interactions with target mRNAs [13]. Here, we analyze hsa_circ_0006054 as a putative candidate for playing a role in colorectal cancer as shown in Figure 7. Firstly, the potential miRNA targets on hsa_circ_0006054 are predicted with the TargetScan perl script, and strong experimentally validated miRNA targets are obtained from miRTarBase [34]. Then, the hsa_circ_0006054-miRNA-mRNA regulatory network is constructed. In addition, miRNA-related dysfunctions are widely associated with various diseases. Therefore, we download the experimentally verified miRNA-disease associations from HMDD [35], and mark the colorectal cancer related miRNAs in the hsa_circ_0006054-miRNA-mRNA network. The results showed that hsa_circ_0006054 may be targeted by six miRNAs including hsa-miR-153, hsa-miR-194, hsa-miR-217, hsa-miR-431, hsa-miR-503, hsa-miR-646, and function as hsa-miR-217 sponge in the carcinogenesis of colorectal cancer.



(A)



(B)



(C)

Figure 6. Prediction performance of KATZHCD model with different similarity measures in the framework of LOOCV. (A) Performance comparison among five similarity measures when *k* was set as 2. (B) Performance comparison among five similarity measures when *k* was set as 3. (C) Performance comparison among five similarity measures when *k* was set as 4.

effectiveness of our proposed method. The KATZ model employs the simple measure on the heterogeneous network, and succeed in predicting disease-related biological molecules. Secondly, reliable biological datasets are used to establish the circRNA similarity network and disease similarity network. Furthermore, KATZHCDCA can predict the scores between circRNA and diseases simultaneously for all diseases. Hence, KATZHCDCA is a useful biomedical resource for circRNA-disease association identification.

Despite the effectiveness of KATZHCDCA model, it should be noted that KATZHCDCA still has some limitations. First, the model mainly depends on the known circRNA-disease associations, which may cause possible bias by imbalanced training samples. The parameters were also influenced by the sparsity of the associations. With the addition of more links between circRNAs and diseases, the model may obtain higher prediction scores. Moreover, other kinds of information of circRNAs or diseases could be integrated to improve the predictive performance, such as miRNA-based circRNA similarity, circRNA-protein similarity, disease semantic similarity and so on. In addition, KATZHCDCA is not applicable to predict novel circRNA-disease associations that without any known associations. Therefore, integrating more data sources and appropriate algorithms would improve the predictive performance.

Abbreviations

circRNAs: circular RNAs; miRNAs: microRNAs; GIP: Gaussian interaction profiles; LOOCV: leave-one-out cross validation; 5-fold CV: 5-fold cross validation; OMIM: Online Mendelian Inheritance in Man; ROC: receiver operating characteristic; TPR: true positive rate; FPR: false positive rate; SEN: sensitivity; SPE: specificity; AUC: area under the curve; ceRNAs: competing endogenous RNAs.

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Competing Interests

The authors have declared that no competing interest exists.

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