

Supplemental materials

Quantitative proteomics of HFD-induced fatty liver uncovers novel transcription factors of lipid metabolism

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Supplemental methods

Glucose tolerance tests and insulin tolerance tests

Glucose tolerance was monitored by glucose tolerance tests (GTT) 10 days before sacrificing the mice. One week later, the same mice were used for insulin tolerance tests (ITT). For GTT, mice were fasted for 12 h. After measuring the baseline blood glucose level via a tail nick using a glucometer, 1.5 g/kg glucose was administered via intraperitoneal injection, and glucose levels were measured at 0, 30, 60, and 120 min after the glucose injection. For ITT, mice fasted for 6 h were injected intraperitoneally with recombinant human insulin at a dose of 1 U/kg, and their blood glucose concentrations were determined at 0, 15, 30, 60, and 120 min after the insulin injection.

Primary hepatocytes isolation, cell culture, and Oil red O staining

Primary hepatocytes were isolated from mice fed with normal chow diet. Briefly, after anesthesia, mice were perfused with a buffer solution lacking Ca^{2+} and Mg^{2+} through the portal vein, and then perfused with 0.05% type IV collagenase. After digestion, the liver was excised, minced, and filtered through a 70 μm filter. Then, hepatocytes were separated by centrifugation at $50 \times g$ for 2 min twice. The obtained hepatocytes were resuspended in DMEM supplemented with 10% FBS and 1% penicillin streptomycin, and seeded in six-well plates. 0.5mM of palmitate acid (PA) and 1mM oil acid (OA) was added to the medium for 24 h to establish the in vitro model of lipid accumulation in hepatocytes.

AML12 and HepG2 cells were obtained from the Type Culture Collection of the Chinese Academy of Science and were cultured in DMEM/F12 supplemented with 10% FBS and 1% penicillin streptomycin. Cell cryopreservation using serum-free cell freezing medium according to manufacturer's instructions (UUBio, Suzhou, China). The cells were seeded in 12-well plates at 1×10^5 cells/well. 0.5mM palmitic acid (PA) and 1mM oil acid (OA) were added to the medium for 24 h to establish the in vitro model of lipid accumulation in AML12 cells. The cells were fixed with 4% paraformaldehyde

for 20 min, and then stained with 60% Oil Red O working solution for 1 min to examine intracellular lipid accumulation.

RNA extraction and RT-qPCR

Total RNA was extracted from mouse livers and hepatocytes using Total RNA extraction kit according to manufacturer's instructions (UUBio, Suzhou, China). 1 µg of total RNA was reverse-transcribed to cDNA using the HiScript II 1st Strand Cdna Synthesis Kit (Vazyme, Nanjing, China). RT-qPCR was performed using the SYBR Green PCR kit (Vazyme, Nanjing, China) on the Light Cycler 480 realtime PCR system (Roche, Switzerland). Gene expression was normalized to the expression of housekeeping gene Gapdh. The primers are listed in Table S1. The relative expression of mRNA was calculated using the $2^{-\Delta\Delta C_t}$ method.

Table S1. siRNA and primer sequences used are listed below.

Gene name	siRNA sequences (5'-3')	
Tcea1-1	CGAGAUACAUAUGUUUCAU	
Tcea1-2	GUACGAAGUAGGAUAUCAA	
Tcea1-3	GUAGUGCUGAUGAACCAAU	
Ilf2-1	GCAGGUAGGAUCAUAUAAA	
Ilf2-2	CCCUAGAACAACAGGAUUAU	
Ilf2-3	GCCTTGCTGAAGAGGAATCA	
Rbbp4-1	CACCAGAAUUGUUGUUUAU	
Rbbp4-2	GCUGAAGUGAAUUGCUUAU	
Rbbp4-3	CCUGCAUCAUUGCAACAAA	
	Forword primers (5'-3')	Reverse primers (5'-3')
Tcea1	GAGAAGAAAGTAGTTCAGCAGC	GCCCGAGGAAAAGATGAAACAT
ILF2	CCTGGGGAACAAAGTCGTGG	TGAGAATTTTCACCGTAGCATCA
Rbbp4	GACGACGCAGTGGAAGAAC	CTGGGCAGTTAAGCTGGGC
Acc1	ATGGGCGGAATGGTCTCTTTC	TGGGGACCTTGTCTTCATCAT
Fasn	GGAGGTGGTGATAGCCGGTAT	TGGGTAATCCATAGAGCCCAG
Acox1	TAACCTCCTCACTCGAAGCCA	AGTTCCATGACCCATCTCTGTC
Cpt1 α	CTCCGCCTGAGCCATGAAG	CACCAGTGATGATGCCATTCT
Srebp1	GCAGCCACCATCTAGCCTG	CAGCAGTGAGTCTGCCTTGAT
Gapdh	AGGTCGGTGTGAACGGATTTG	TGTAGACCATGTAGTTGAGGTCA

Figure legends

Figure S1. HFD mice displayed enhanced glucose intolerance and insulin resistance compared to NCD mice. (A) glucose tolerance test (GTT) on HFD and NCD-fed mice. (B) insulin tolerance test (ITT) on HFD and NCD-fed mice. N=7 biological replicates. All data represent the mean \pm SEM. Student's t test, *** $p < 0.001$.

Figure S2. (A) Biological processes enrichment of DEPs. (B) Weight gene analysis of DEPs.

Figure S3. (A-C) RT-qPCR showing knockdown efficiency of Tcea1, Rbbp4, and ILF2 in AML12 cells transfected with corresponding siRNA for 48 h. (D-F) RT-qPCR showing knockdown efficiency of Tcea1, Rbbp4, and ILF2 in HepG2 cells transfected with corresponding siRNA for 48 h.

Figure S4. Tcea1, Rbbp4, and ILF2 deficiency promoted lipid accumulation of HepG2 cells, respectively. (A) Representative images of Oil red O (ORO) staining of HepG2 cells after Tcea1, Rbbp4, and ILF2 knockdown, respectively. Scale bar: 100 μ m. (C) Quantification of relative ORO positive area. All data represent the mean \pm SEM. Student's t test, ** $P < 0.01$, *** $P < 0.001$

Figure S1

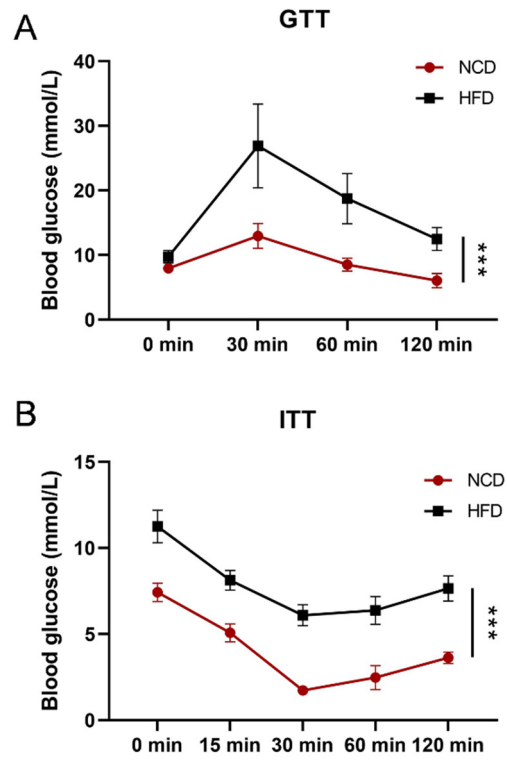


Figure S2

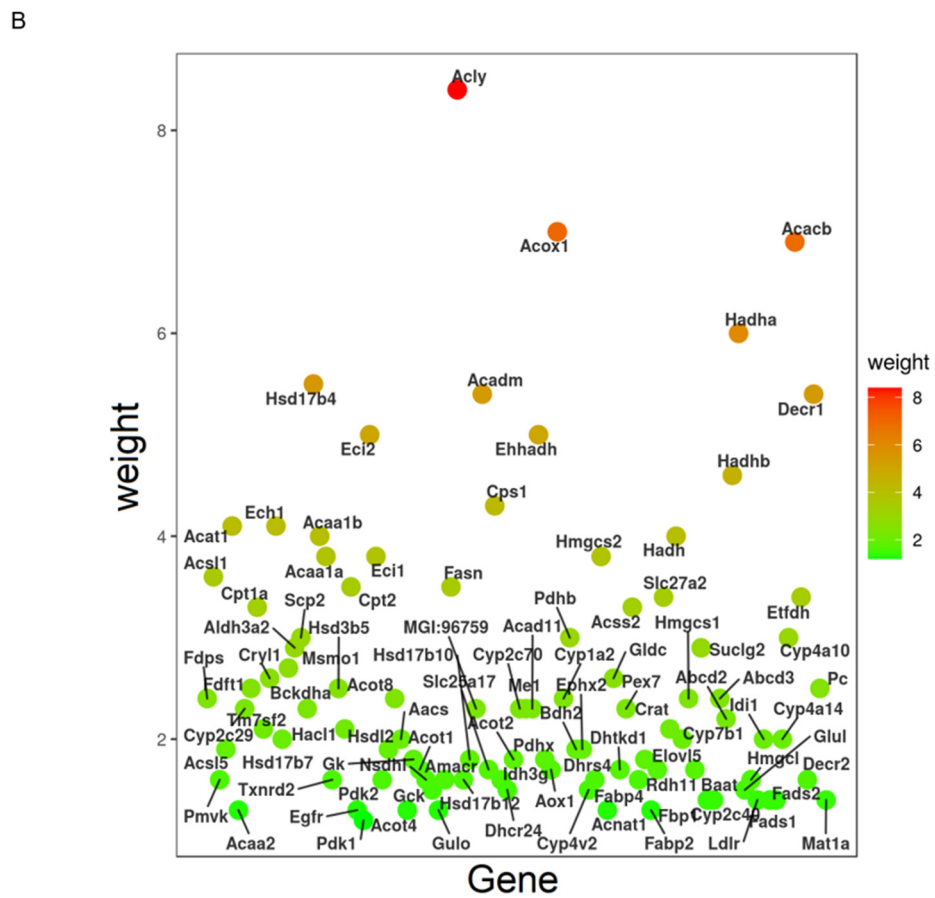
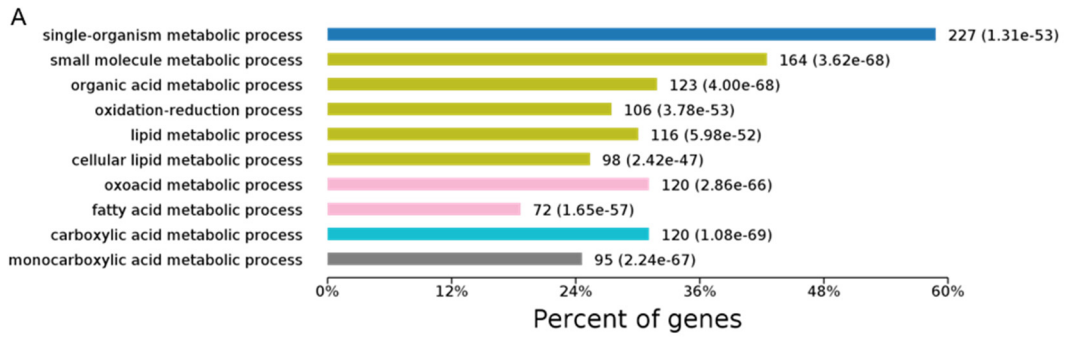


Figure S3

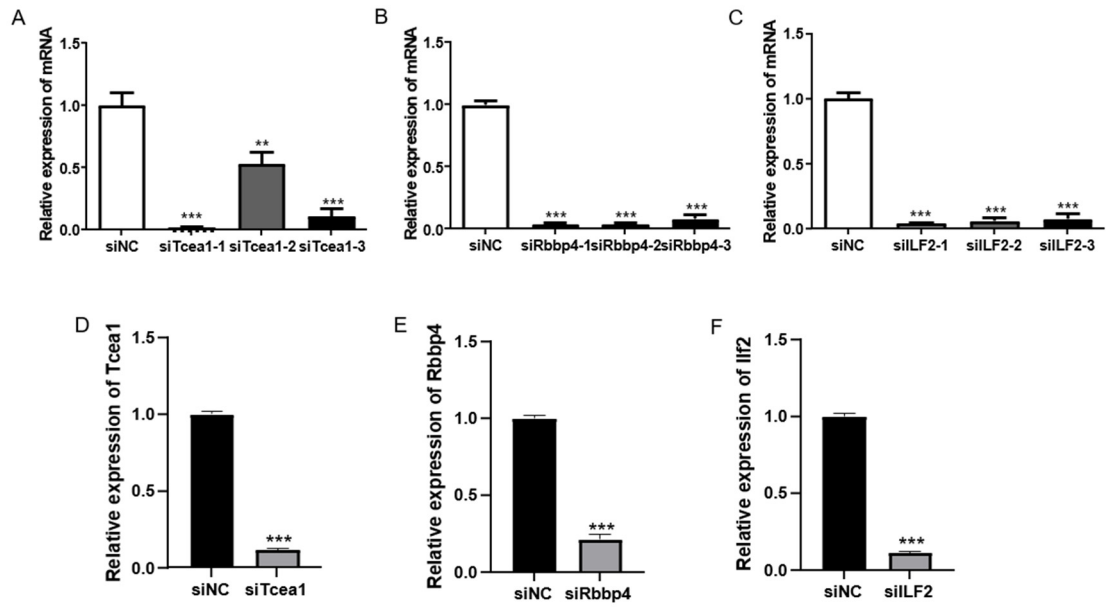


Figure S4

