

**Supplementary material for**  
**Ubiquitin like protein FAT10 repressed cardiac fibrosis after myocardial ischemic via mediating degradation of Smad3 dependent on FAT10-proteasome system**

**Running title: FAT10 and cardiac fibrosis**

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**1. Supplemental Methods**

### **1.1 Masson, Sirius Red, Immunochemistry and Immunofluorescence staining**

Heart sections were prepared at 4- $\mu\text{m}$  thickness through a routine procedure. The sections were subsequently stained with Masson trichrome and Sirius Red staining to determine the extent of fibrosis<sup>1</sup>. For immunochemistry and immunofluorescence staining (IHC), the sections were incubated with primary antibodies which listed in **Supplementary Table 1**.

### **1.2 Ischemic animal model and surgical procedure**

Myocardial infarction (MI) model was created in *Fat10*<sup>-/-</sup> and WT mice via ligation of the left anterior descending artery (LAD). Regional ischemia of the heart was confirmed by ECG, which showed ST elevation. Control group underwent sham operations without ligation of the LAD.

### **1.3 Echocardiography**

In Briefly, mice were anesthetized with 1% isoflurane at a rate of 1.0%/min. Using the Canadian Visualsonics Vevo 2100 color ultrasound diagnostic apparatus, equipped with a 17MHz transducer, scanning the left sternal border of the mouse, the right sternal border, the apex and the sternal fossa, collecting ultrasound images of the heart, and performing cardiac structures, each Measurement of valve blood flow and time parameters.

### **1.4 Adenovirus and plasmid constructs**

All these recombinant adenoviruses (*ad-Fat10*, *sh-Fat10*, *ad-Smad3*, *sh-Smad3*, *sh-Ub*, *Smad3-K378R-wt*, *Smad3-K378R-mut*) were designed by Gene-Chen (Shanghai Co., Ltd). The Flag-*Fat10*, GST-*Fat10*, deletion forms of Smad3(1-134, 135-229, 230-425 amino acids (a.a)), mutant forms Smad3(K333R, K341R, K378R, K409R) (lysine mutated to arginine) were also established from

Gene-Chen (Shanghai Co., Ltd). *Fat10* C-terminal diglycine motif-deficient plasmid ( $\Delta$ *Fat10*), and related adenovirus, *ad-Fat10*, *sh-Fat10*, *ad-Smad3* and *sh-Smad3* adenovirus were obtained from Gene Chem Company (*Shanghai, China*). UBE2Z, and UB6A interference fragment kits were purchased from GenePharma Company (*Shanghai, China*).

### **1.5 Isolation of primary cardiac fibroblasts**

Firstly, the hearts were placed in a 10cm dish with cold Dulbecco's modified Eagle's medium/Low glucose (DMEM, Hyclone, Logan, Utah, USA), and cut to approximately 1 mm<sup>3</sup> with scissors. The heart tissues were digested in 0.25% trypsin with gently shaking, all digestive fluid was collected in DMEM with 10% fetal bovine serum (FBS, ScienCell, San Diego, California, USA) and penicillin (100 IU/ml) and streptomycin (100 mg/ml). Then the cells were filtered through 75  $\mu$ m cell strainer and centrifuged at 1200 rpm for 5 min. Finally, the isolated cardiac cells were seeded in six-well plates for 90 min, the pre-seeding medium containing CM were removed and seeded in new six-well plates, while the CF which could be adhere to the culture plates. All CFs in this study were treated within three passage cultures.

### **1.6 Western blotting (WB), co-immunoprecipitation Co-IP, Immunofluorescent (IF) and GST pull-down assays**

WB, Co-IP, IF and GST pull-down assays were performed as previously described<sup>2,3</sup>. All antibodies listed in **Supplementary Table 1**.

### **1.7 Cell proliferation and Migration**

Edu cell proliferation kit (RiboBio, Guangzhou, China) was used to measure cells proliferation. Wound Healing Assay, transwell assay were used to detect cell migration as describe previously<sup>4</sup>.

## Supplementary Tables

**Supplementary Table 1.** The baseline of cardiac function of wild-type and *Fat10<sup>-/-</sup>* mice measured by echocardiography

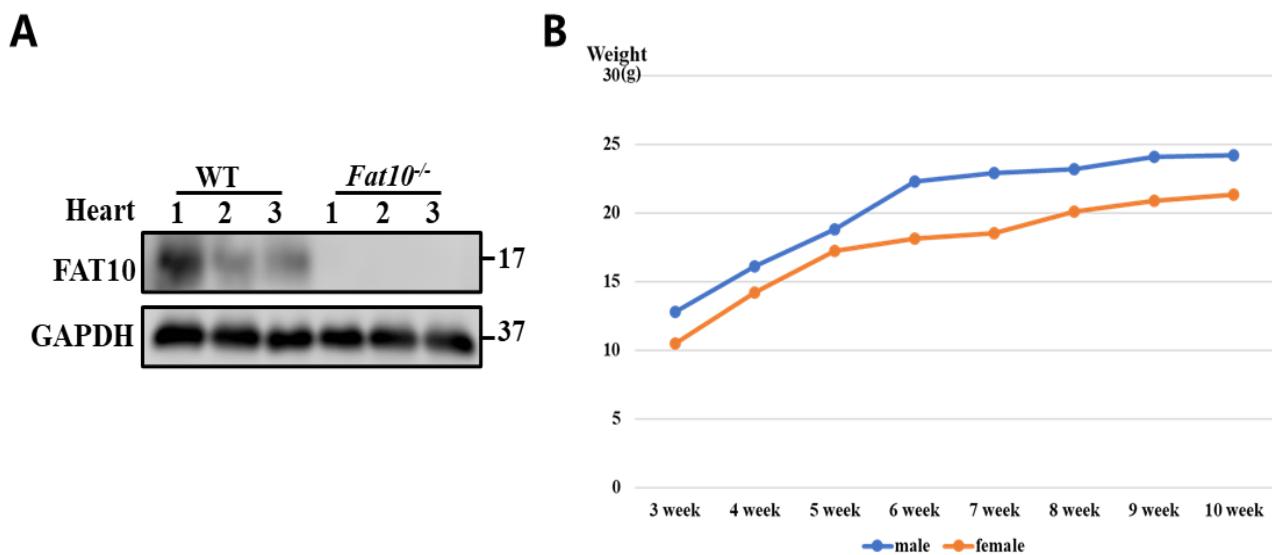
Cardiac function	<i>Wild-type</i> (n=6)	<i>Fat10<sup>-/-</sup></i> (n=6)	P
LVVol;d	36.87±1.73	37.3±1.51	0.66
LVVol;s	13.03±0.45	12.94±0.60	0.78
IVSd(mm)	0.71±0.06	0.70±0.05	0.95
IVSs(mm)	0.84±0.05	0.81±0.03	0.70
LVIDd(mm)	3.07±0.09	3.05±0.06	0.91
LVIDs(mm)	1.83±0.08	1.87±0.08	0.39
LVPWd(mm)	0.91±0.08	0.88±0.12	0.59
LVPWs(mm)	1.06±0.12	1.00±0.01	0.44

LVVol;d: left ventricular volume at diastolic phase; LVVol;s: left ventricular volume at systolic phase; SV: stroke volume; LVIDd: left ventricular internal diameter at diastolic phase; LVIDs: left ventricular internal diameter at systolic phase; IVSd: interventricular septal thickness at diastolic phase; IVSs: interventricular septal thickness at systolic phase; LVPWd: left ventricular posterior wall thickness at diastolic phase; LVPWs: left ventricular posterior wall thickness at systolic phase.

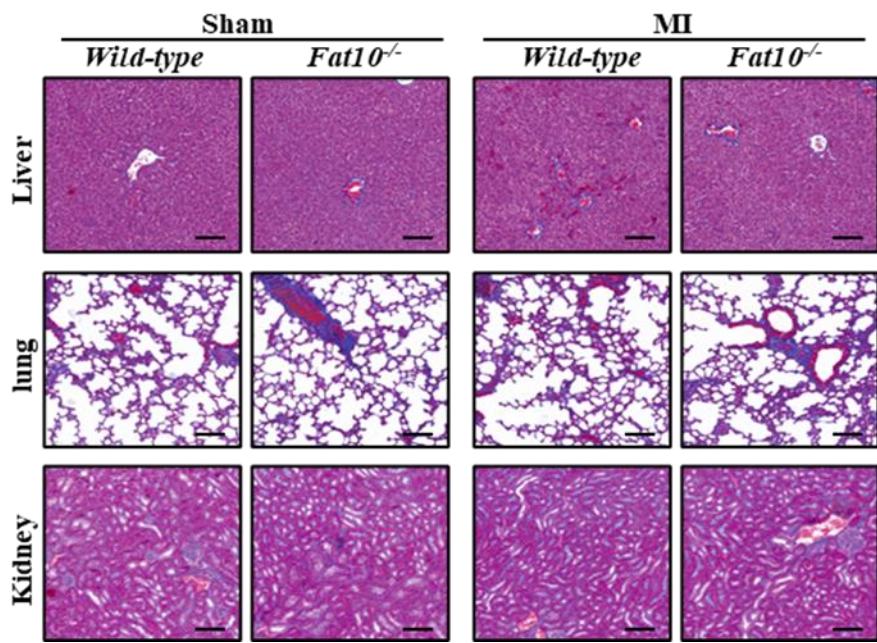
**Supplementary Table 2.** The following antibodies were used for western blotting and immunostaining

Target protein	Source (Catalog number)
FAT10	Abcam (ab168680)
Smad3	Abcam (ab40854)
Smad3	66516-1-Ig
UBD Fushon Protein	Proteintech (Ag27706)
Recombinant Human Smad3 protein	Abcam (ab89353)
Smooth Muscle Actin	Proteintech (14395-1-AP)
Collagen Type I	Proteintech (14695-1-AP)
Collagen Type III	Proteintech (22734-1-AP)
p-Smad3	Abcam (ab52903)
Smad2	Abcam (ab40855)
p-Smad2	Abcam (ab280888)
Smad4	Abcam (EP618Y)
Smad7	Proteintech (25840-1-AP)
GST	Proteintech (10000-0-AP)
Flag	Proteintech (66008-2-Ig)
HA	Proteintech (51064-2-AP)
His	Proteintech (66005-1-Ig)
GAPDH	Proteintech (60004-1-Ig)
MG132	Selleck (S2619)
CHX	Selleck (S3648)
TNF- $\alpha$ /IFN- $\gamma$	PeproTech

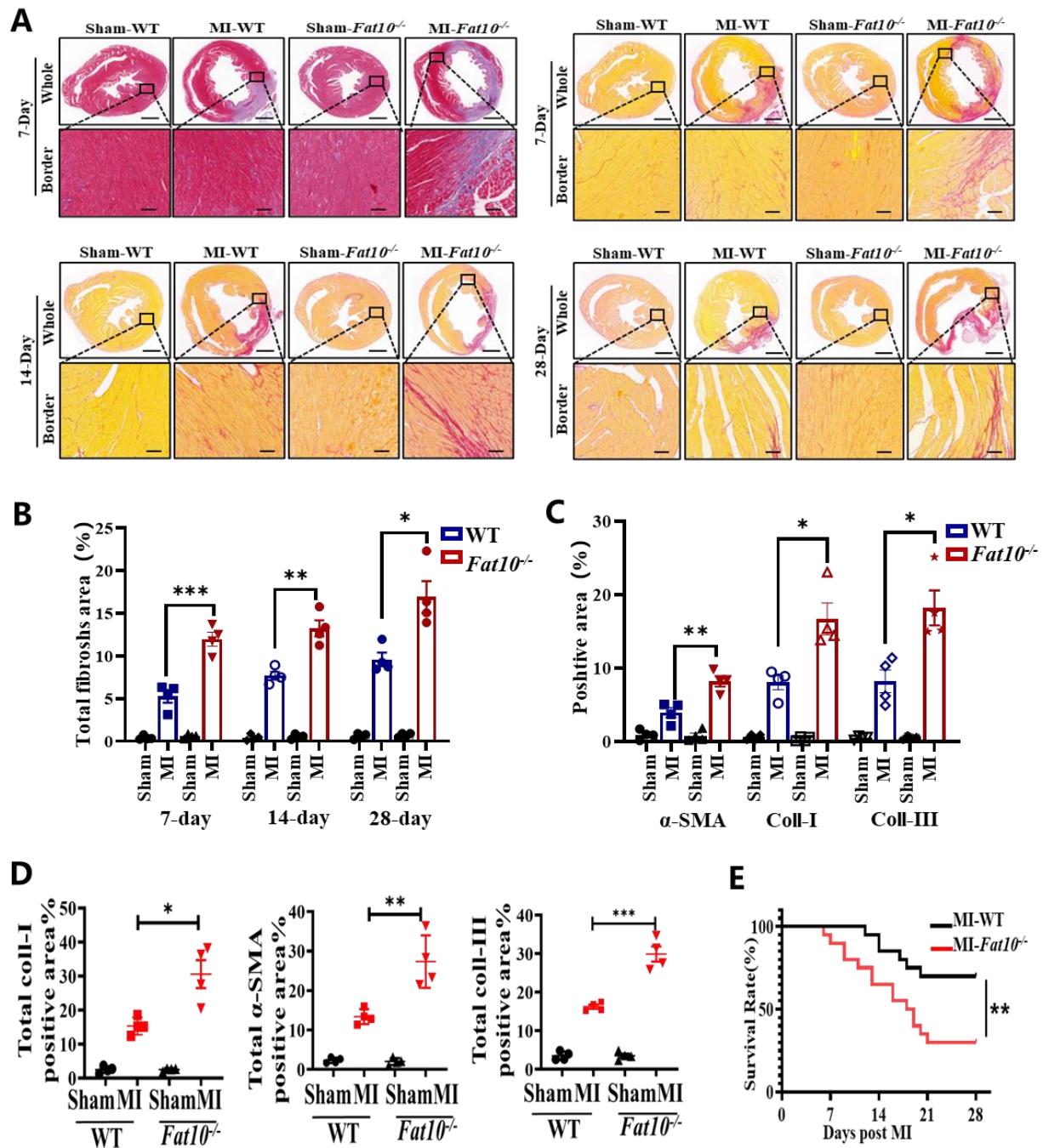
## Supplemental Figures



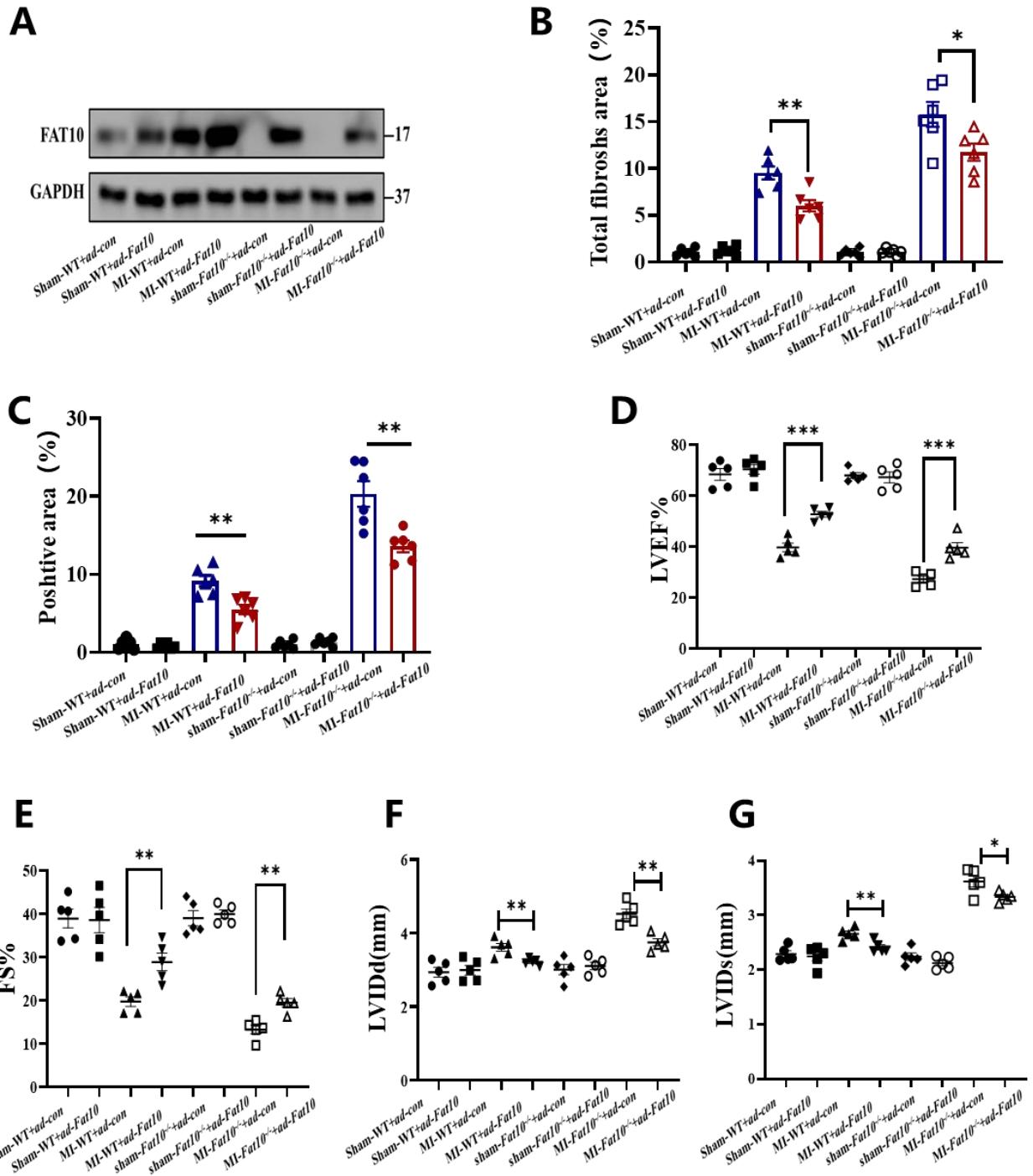
**Supplemental Figure S1. Validation and basic information of *Fat10*<sup>-/-</sup> mice.** (A) Western blotting was performed to detect the expression of FAT10 in heart tissues of WT and *Fat10*<sup>-/-</sup> mice. (B) Growth weight curves of *Fat10*<sup>-/-</sup> mice.



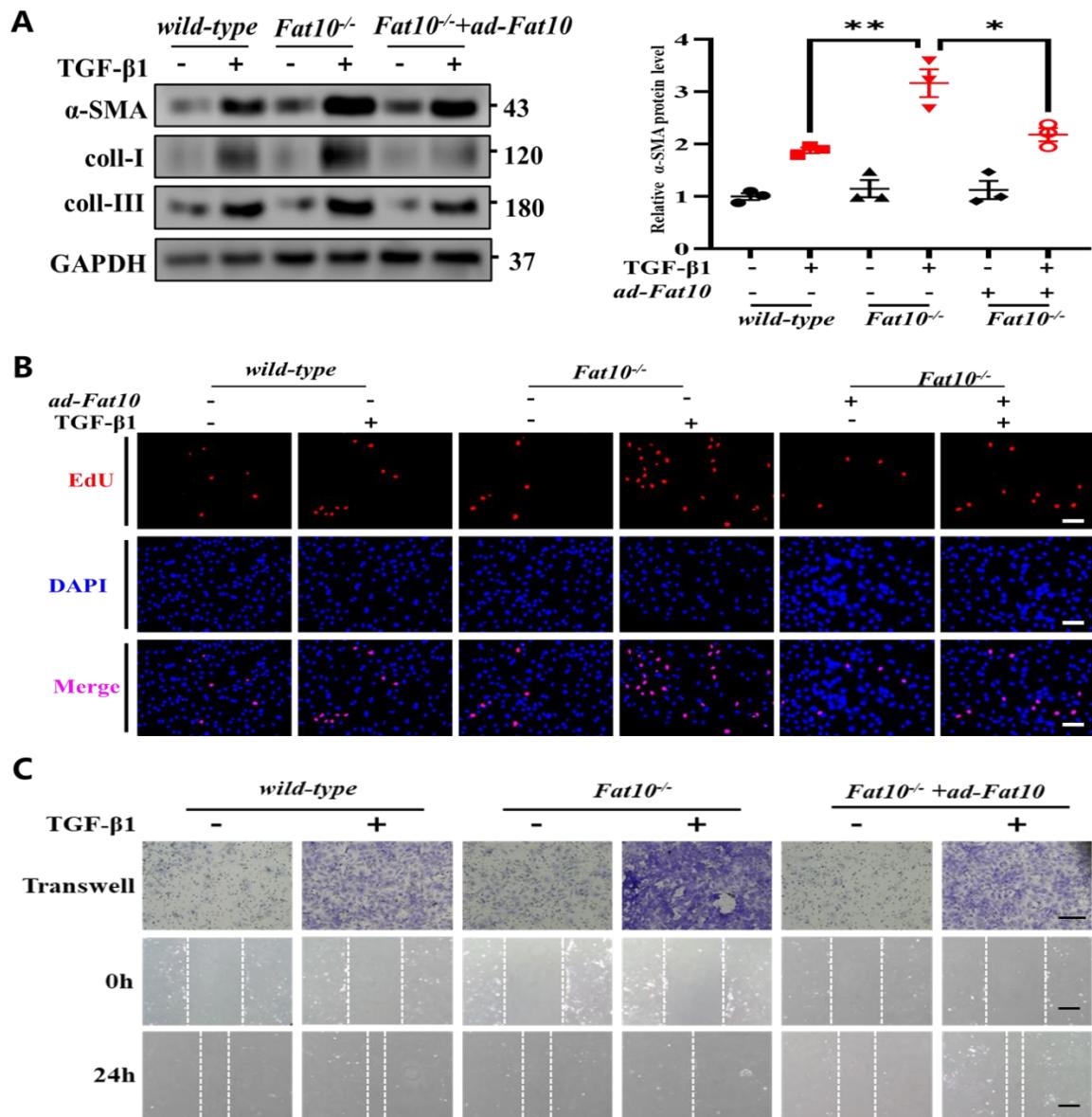
**Supplemental Figure S2.** Masson staining of liver, lung and kidney in each group.



**Supplemental Figure S3. FAT10 deficiency aggravates MI-induced cardiac fibrosis** (A) Masson and Sirius Red staining of whole heart fibrosis (up, bar 1mm) and infarct border fibrosis (down, bar 50μm). (B) Quantification of the total fibrotic area by Masson staining in 7, 14, 28day after being subject MI operation. (C) Quantification of α-SMA, collagen I and collagen III positive area by Immunofluorescence staining from WT and Fat10<sup>-/-</sup> mice after being subject 14day-sham operation (n=4 per group). (D) Quantification of α-SMA, collagen I and collagen III positive area by IHC from WT and Fat10<sup>-/-</sup> mice after being subject 14day-sham operation (E) Kaplan-Meier survival curves of mice in the 28 days after MI (n=20 per group). \*p<0.05, \*\*p<0.01, \*\*\*p<0.001 and ns=not significant. All data presented as mean±SEM.

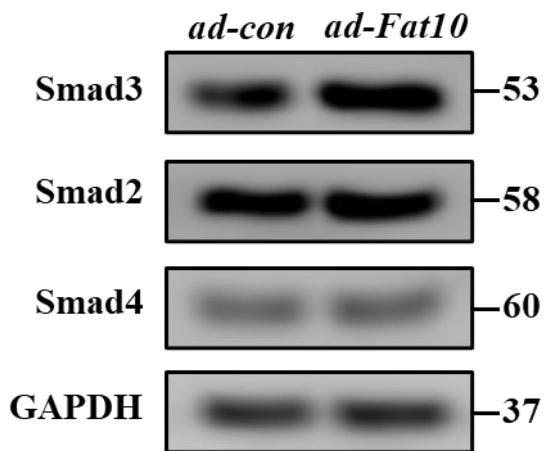


**Supplemental Figure S4. Overexpression of FAT10 alleviates the MI-induced cardiac fibrosis.** **(A)** Western blotting analyses of FAT10 expression in per group. **(B)** Quantification of the total fibrotic area by Masson staining in per group. **(C)** Quantification of collagen I by Immunofluorescence staining in per group. **(D-G)** Echocardiographic analysis to evaluate left ventricular ejection fraction (LVEF), fractional shortening (FS), left ventricular internal diameter at end-diastole (LVIDd) and left ventricular internal diameter at end-systole (LVIDs) in WT and Fat10<sup>-/-</sup> after sham and 2 weeks post-MI surgery. \*p < 0.05, \*\*p < 0.01, \*\*\*p < 0.001 and ns=not significant. All data presented as mean ± SEM; comparison by unpaired t-test.

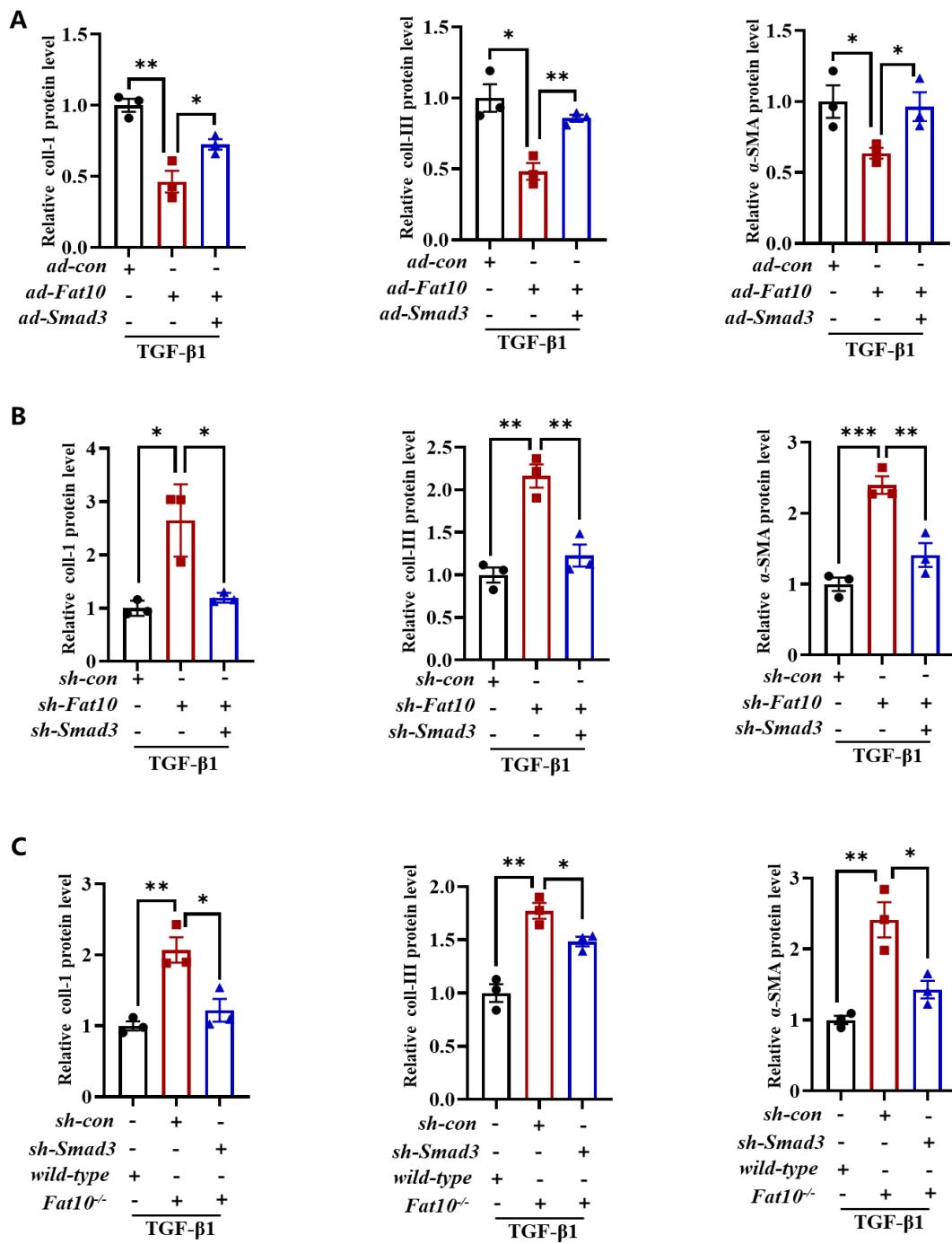


**Supplemental Figure S5. FAT10 inhibits the TGF- $\beta$ 1-induced fibrotic response in primary mouse CFs.**  
 Primary mouse CFs from the LV of WT mice and *Fat10*<sup>-/-</sup> mice and treated with TGF- $\beta$ 1. (A) The level of  $\alpha$ -SMA, collagen I and collagen III were detected by western blotting analysis. (B) Representative immunofluorescence images of EdU(red) and DAPI (blue)(left) (right) in the different experimental groups (bar 50 $\mu$ m). (C) Migration measured by transwell assay (bar 100 $\mu$ m) and wound-healing assay (bar 100 $\mu$ m).

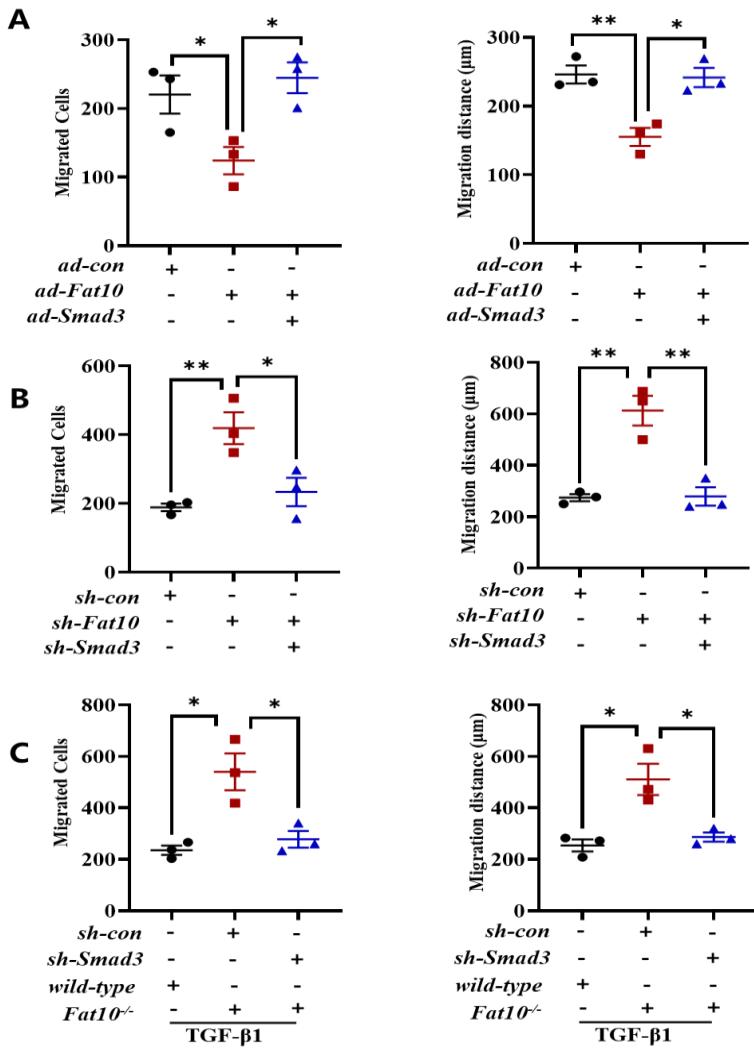
\*p<0.05, \*\*p<0.01, \*\*\*p<0.001 and ns=not significant. All data presented as mean $\pm$ SEM.



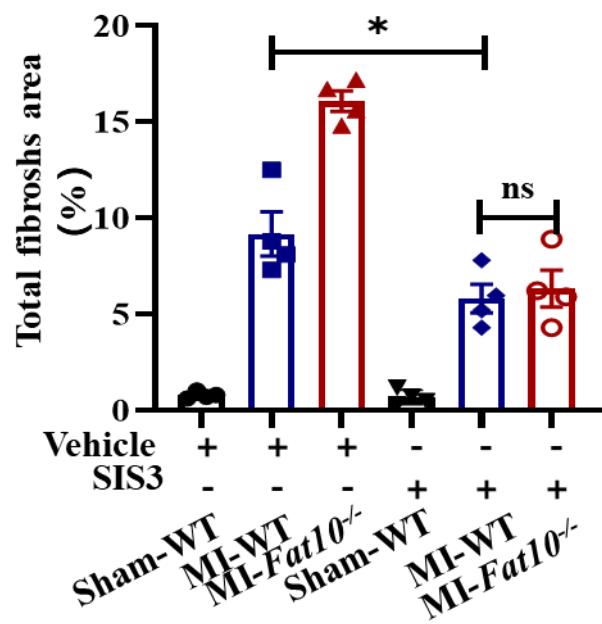
**Supplemental Figure S6. FAT10 did not affect Smad2 and Smad4 protein expression.** Western blotting analysis of Smad2, and Smad4 expression.



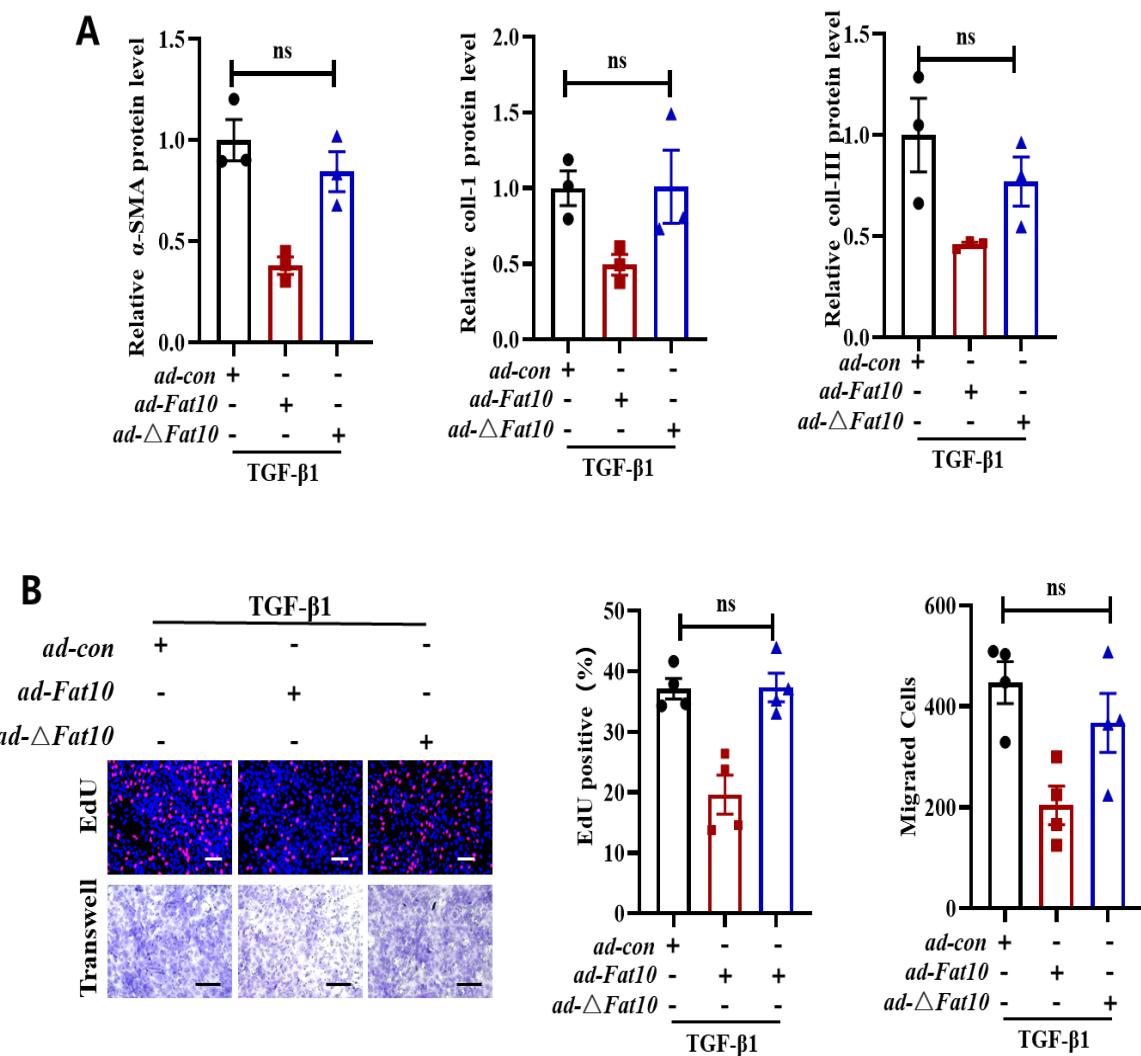
**Supplemental Figure S7. FAT10 regulates the expressions of cardiac fibrotic marker by affecting Smad3.** All group were transfected with adenovirus and then exposed to TGF- $\beta$ 1. **(A)** Quantitative assessment of the protein level of collagen I, collagen III and  $\alpha$ -SMA, NRCFs were co-transfected with *ad-Fat10* and *ad-Smad3* or *ad-con* (n=3 per group). **(B)** Quantitative assessment of the protein level of collagen I, collagen III and  $\alpha$ -SMA, NRCFs were co-transfected with *sh-Fat10* and *sh-Smad3* or *sh-con* (n=3 per group). **(C)** Quantitative assessment of the protein level of collagen I, collagen III and  $\alpha$ -SMA, *Fat10 $^{-/-}$*  and WT primary CF were transfected with *sh-Smad3* (n=3 per group). \*p<0.05, \*\*p<0.01, \*\*\*p<0.001 and ns=not significant. All data presented as mean $\pm$ SEM.



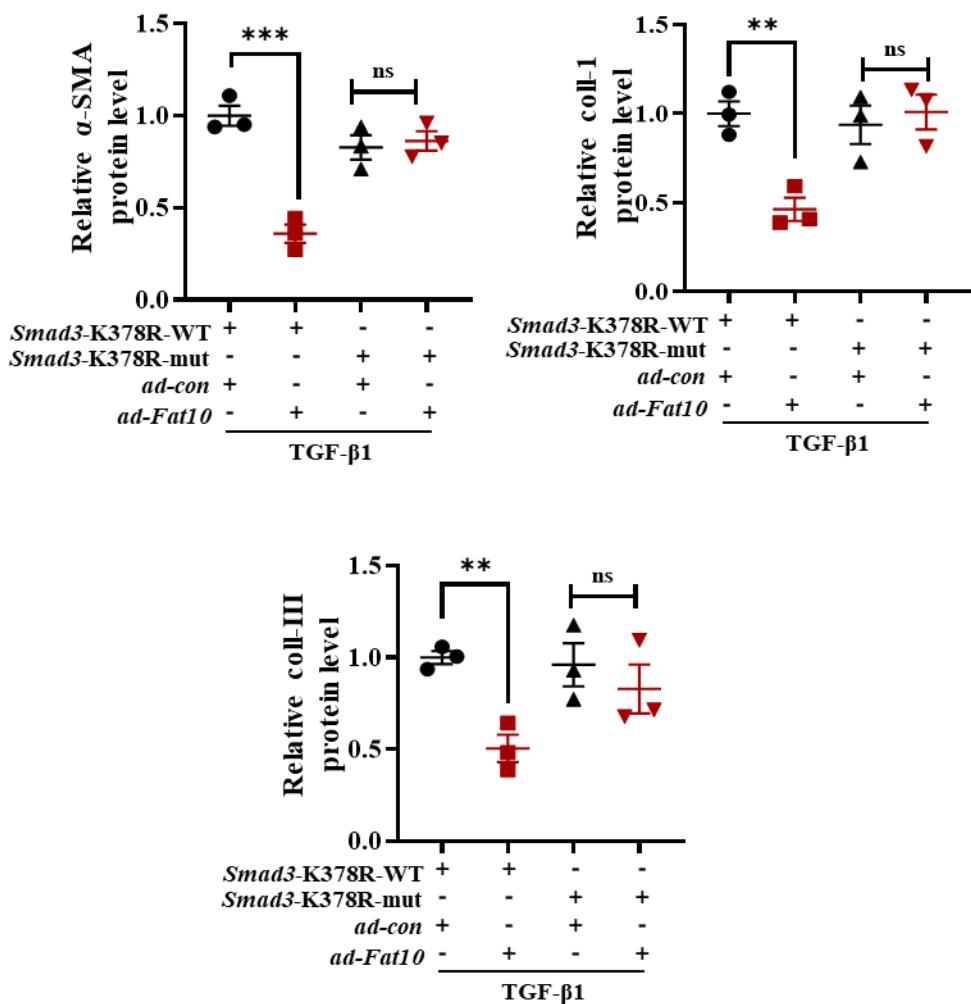
**Supplemental Figure S8. FAT10 regulates the CF migration by affecting Smad3.** All group were transfected with adenovirus and then exposed to TGF- $\beta$ 1. **(A)** The quantitative data of CF migration by transwell assay (left) and wound-healing assay (right), NRCFs were co-transfected with *ad-Fat10* and *ad-Smad3* or *ad-con* (n=3 per group). **(B)** The quantitative data of CF migration by Transwell assay (left) and wound-healing assay (right), NRCFs were co-transfected with *sh-Fat10* and *sh-Smad3* or *sh-con* (n=3 per group). **(C)** The quantitative data of CF migration by transwell assay (left) and wound-healing assay (right), *Fat10<sup>-/-</sup>* and WT primary CF were transfected with *sh-Smad3* (n=3 per group). \*p<0.05, \*\*p<0.01, \*\*\*p<0.001 and ns=not significant. All data presented as mean±SEM.



**Supplemental Figure S9.** Quantification of the total fibrotic area by Masson staining in per group.



**Supplemental Figure S10. FAT10 regulates the cardiac fibrotic marker via the K378 site of Smad3.** All group were transfected with adenovirus and then exposed to TGF- $\beta$ 1. **(A)** Quantitative assessment of the protein level of  $\alpha$ -SMA, collagen and collagen III when NRCFs after transfection with *ad-Fat10* or *ad-ΔFat10* ( $n=3$  per group). **(B)** Proliferation measured by EdU assay (up) and migration measured by transwell assay (down) ( $n=3$  per group).



**Supplemental Figure S11.** Quantitative assessment of the protein level of  $\alpha$ -SMA, collagen and collagen III when NRCPs after transfection with Smad3-K378R-wt or Smad3-K378R-mut in the presence of transfection with *ad-Fat10* (n=3 per group). \*p<0.05, \*\*p<0.01, \*\*\*p<0.001 and ns=not significant. All data presented as mean  $\pm$  SEM.

## References:

1. Nagai T, Honda S, Sugano Y, Matsuyama TA, Ohta-Ogo K, Asaumi Y, Ikeda Y, Kusano K, Ishihara M, Yasuda S, Ogawa H, Ishibashi-Ueda H, Anzai T. Decreased myocardial dendritic cells is associated with impaired reparative fibrosis and development of cardiac rupture after myocardial infarction in humans. *J Am Heart Assoc* 2014;3(3):e839.
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