

1 **IL-6 signaling accelerates iron overload by upregulating DMT1 in endothelial cells to**
2 **promote aortic dissection**

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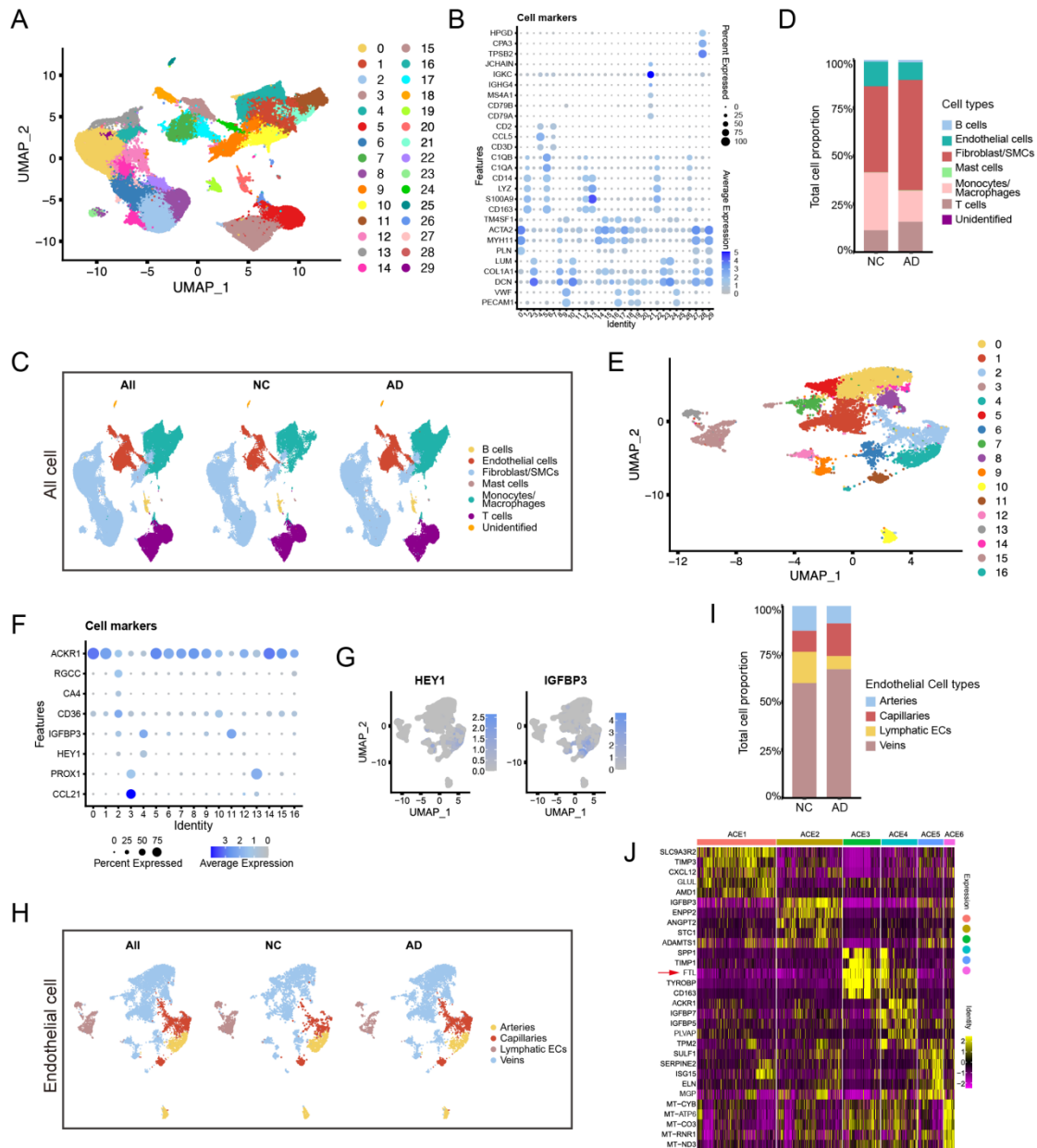
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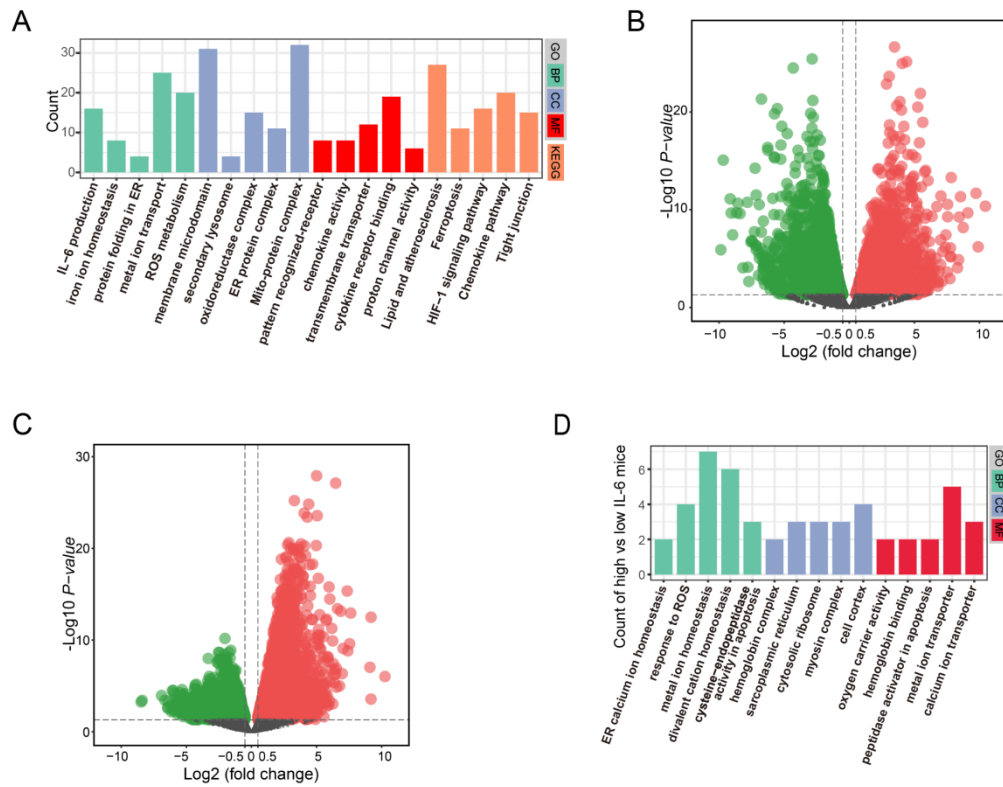
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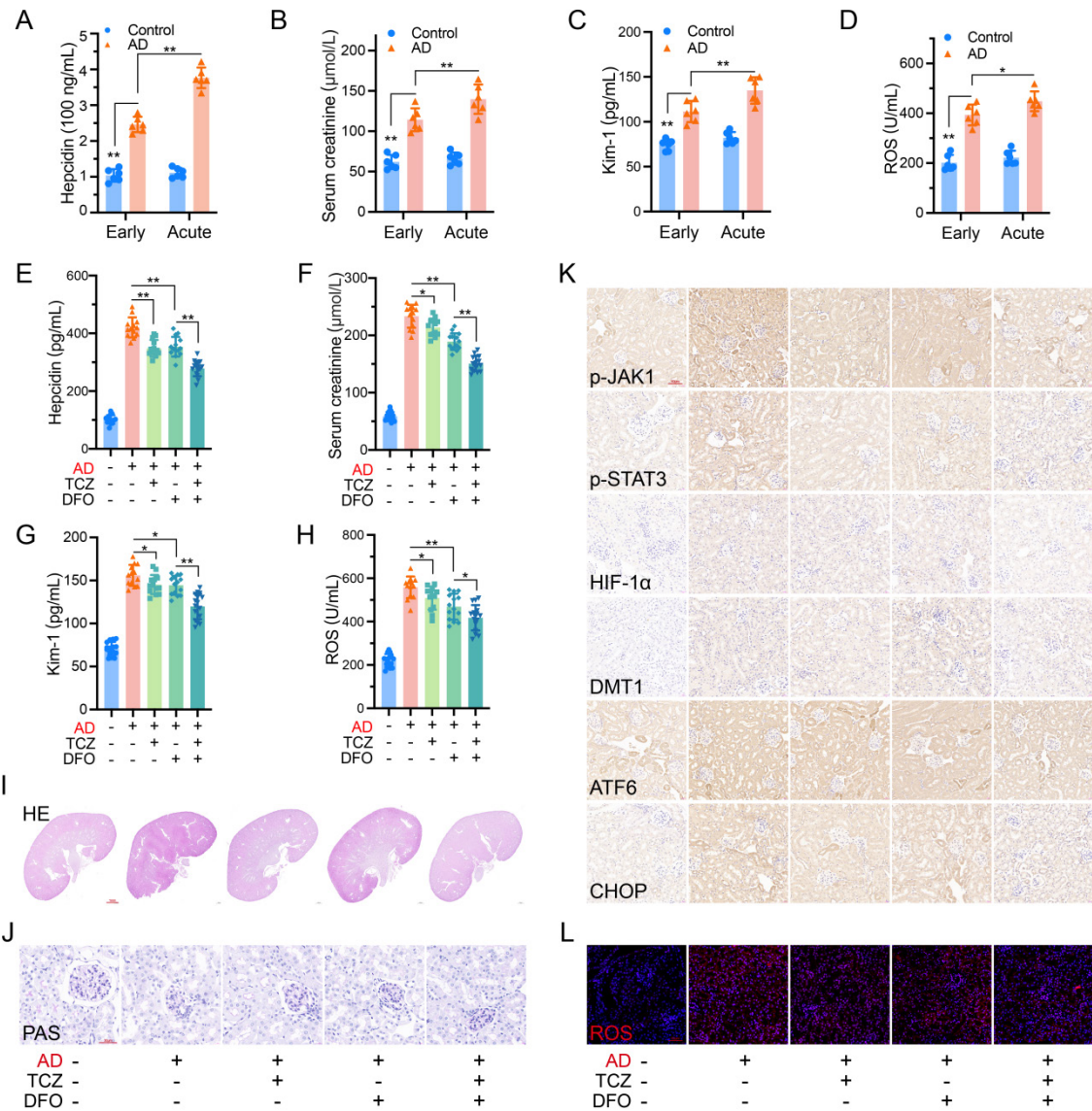
19 **Supplementary Table count:** 5



20 **Fig. S1.** Dimensionality reduction and clustering of aortic cells in scRNA-seq data. (A)
 21 UMAP plot of all cells and separate clusters. (B) Gene expression profiles of selected markers
 22 in separate clusters. (C) UMAP plot of seven cell types in the aortas. (D) Proportion of
 23 different cell types in the aortas. (E) Separate clusters of all endothelial cells (ECs). (F) Gene
 24 expression profiles of selected markers in separate clusters of ECs. (G) UMAP plot of
 25 the expression of aortic endothelial cell (AEC) marker genes (*HEY1* and *IGFBP3*). (H)
 26 UMAP of EC types in the aortas. (I) Proportion of different ECs. (J) Heatmap showing the
 27 top five genes in AECs.



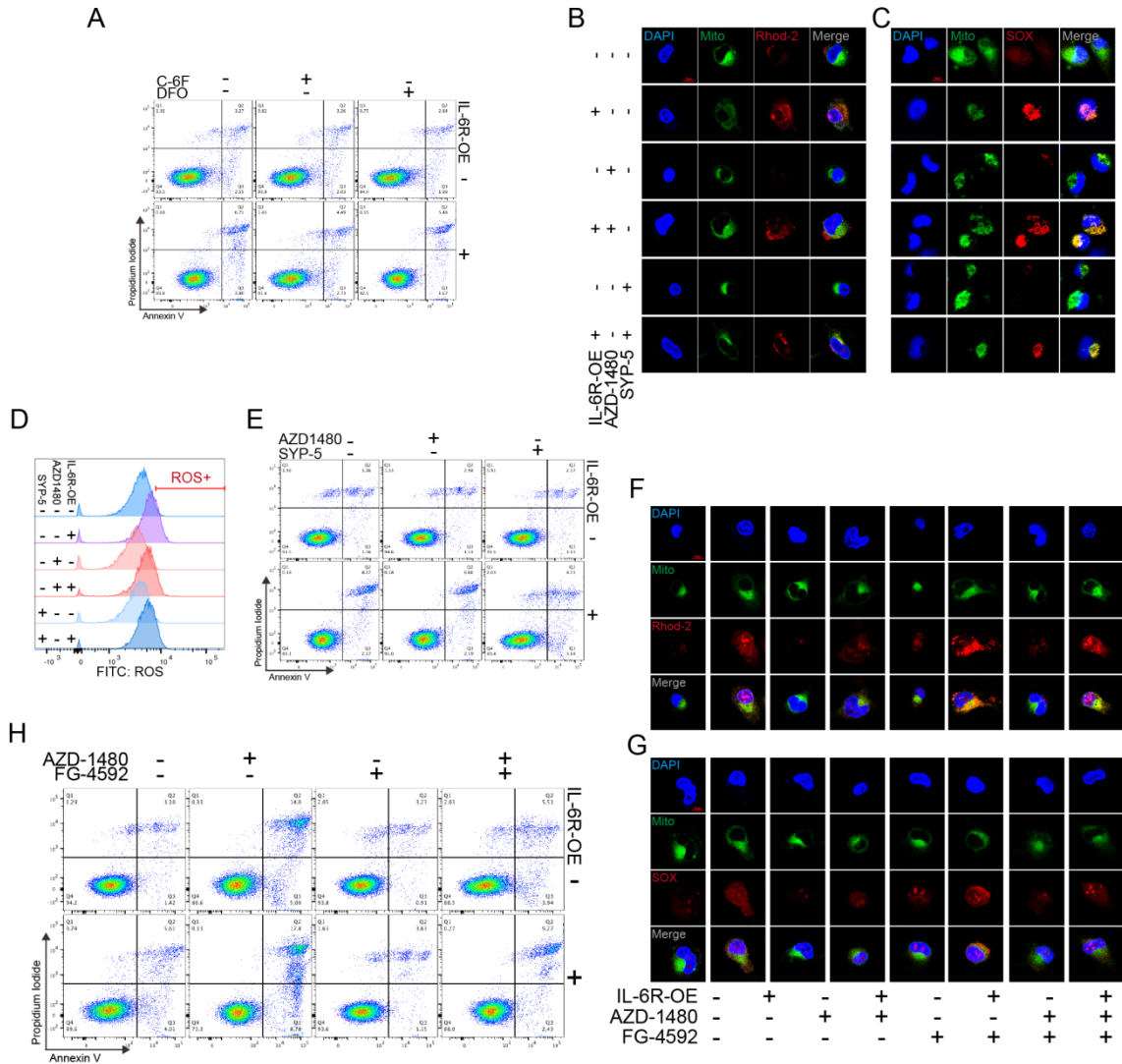
28 **Fig. S2.** Volcano plot and enrichment analysis of differentially expressed genes (DEGs) in
 29 scRNA-seq and bulk RNA-seq. **(A)** GO and KEGG analyses of DEGs in aortic endothelial
 30 cell 3 of scRNA-seq. **(B)** Volcano plot of bulk RNA-seq showing DEGs in aortas of dissection
 31 patients and transplant donors. **(C)** Volcano plot showing DEGs in the aortas of six control
 32 and six acute AD mice. **(D)** GO and KEGG analyses of DEGs between acute control and AD
 33 mouse aortas.



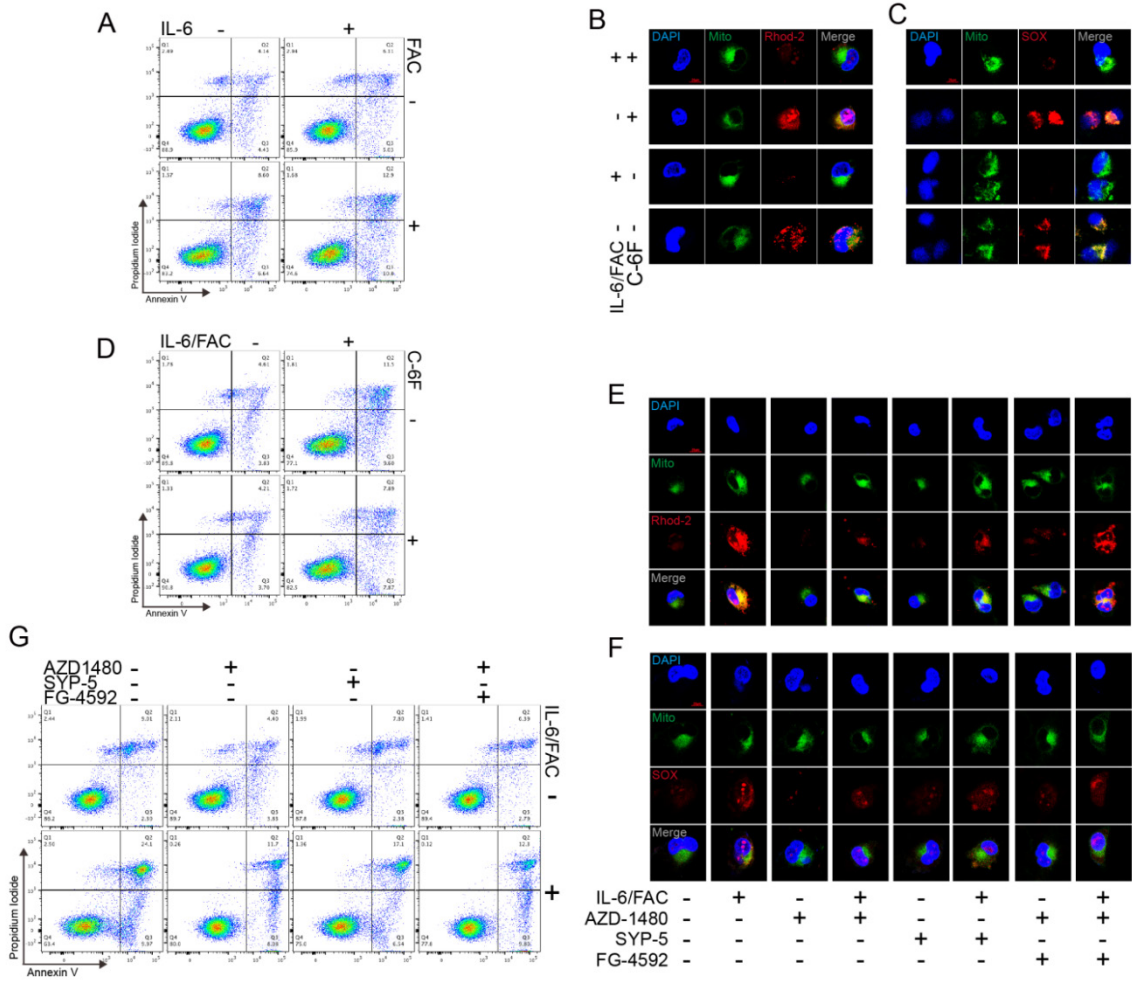
34 **Fig. S3.** Serum biomarker levels and renal section staining in mice with aortic dissection.

35 Contents of serum hepcidin (A), creatinine (B), Kim-1 (C), and ROS (D) in the first animal
 36 experiment. TCZ and DFO inhibited serum levels of hepcidin (E), creatinine (F), Kim-1 (G),
 37 and ROS (H). (I) HE staining of coronal sections of mouse kidneys. (J) PAS staining of renal
 38 sections. (K) IHC staining of renal sections. (L) ROS staining of renal tissues.

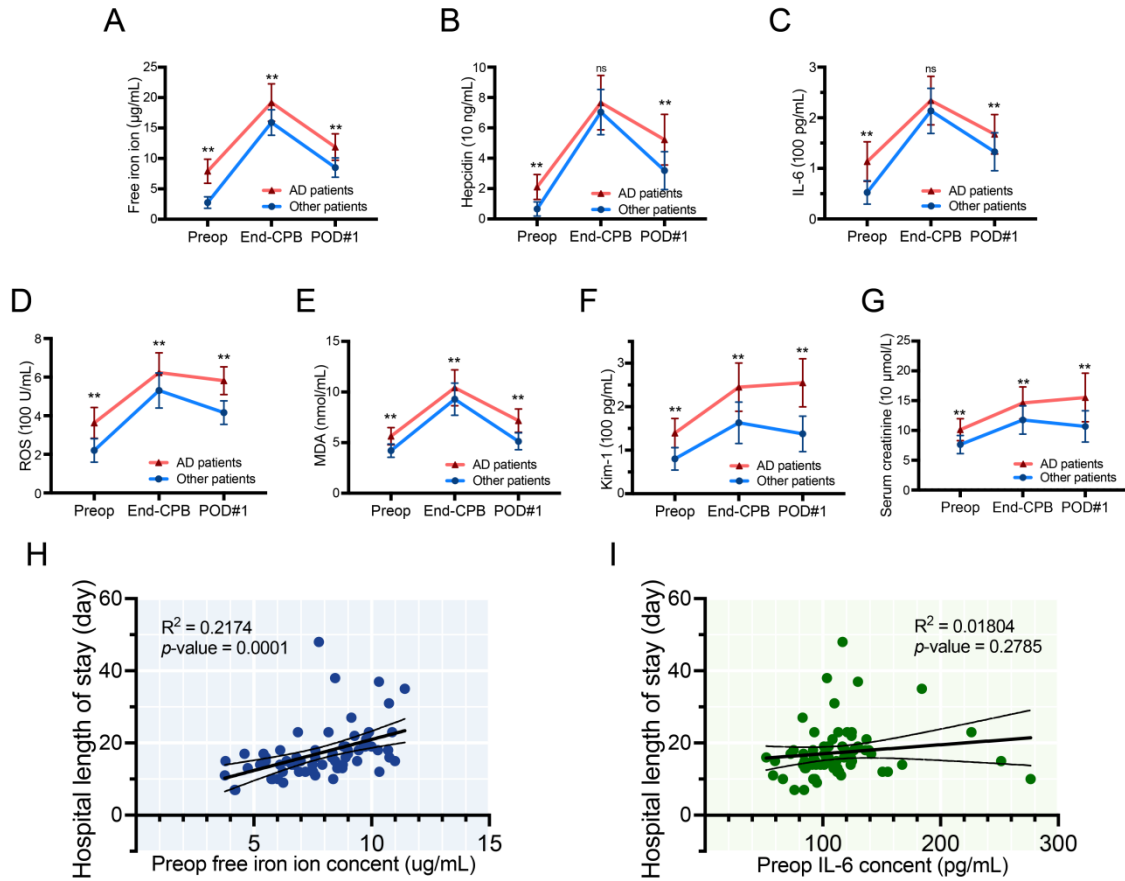
39 Data are presented as representative images or means \pm SDs. * $P < 0.05$, ** $P < 0.01$, ns, not
 40 significant.



41 **Fig. S4.** IL-6R overexpression induces ERS and apoptosis in HAECs. (A) C-6F and DFO
 42 reduced apoptosis in HAECs. (B) Confocal images of mito- Ca^{2+} fluorescence staining. (C)
 43 Confocal images of mitoSOX fluorescence staining. (D) AZD1480 and SYP-5 reduced ROS
 44 in HAECs. (E) AZD1480 and SYP-5 reduced apoptosis in HAECs. (F) Fluorescence images
 45 of mito- Ca^{2+} . (G) MitoSOX fluorescence staining. (H) Apoptosis of HAECs was detected by
 46 flow cytometry. Data are presented as representative images.



47 **Fig. S5.** Excessive IL-6 sensitized FAC-induced ERS and apoptosis in HAECs. (A) IL-6
 48 sensitized FAC-induced apoptosis in HAECs. (B) Confocal images of mito-Ca²⁺ fluorescence
 49 staining. (C) Confocal images of mitoSOX fluorescence staining. (D) C-6F reduced apoptosis
 50 induced by IL-6 and FAC in HAECs. (E) Fluorescence images of mito-Ca²⁺. (F) MitoSOX
 51 fluorescence staining. (G) Apoptosis of HAECs was detected by flow cytometry. Data are
 52 presented as representative images or means ± SDs.



53 **Fig. S6.** Changes in perioperative serum biomarkers and the correlation between biomarkers
54 and prognosis. Levels of serum iron ion (A), hepcidin (B), IL-6 (C), ROS (D), MDA (E),
55 Kim-1 (F), and creatinine (G) in aortic dissection (AD) patients and other patients requiring
56 aortic surgery at pre-operation (Preop), end of cardiopulmonary bypass (End-CPB), and
57 postoperation day one (POD#1). (H) Correlation between preoperative free iron ion and
58 hospital stay in AD patients. (I) Relationship between preoperative IL-6 and hospital stay in
59 dissection patients. Data are presented as representative images or means \pm SDs. * $P < 0.05$,
60 ** $P < 0.01$, ns, not significant.

61 **Table S1.** The primers sequence information.

Categories	Forward (5'--3')	Reverse (5'--3')
RT-PCR assay		
Human		
<i>FTH1</i>	-CCAGAACTACCACCAGGACTC-	-GAAGATTCGGCCACCTCGTT-
<i>FTL</i>	-CAGCCTGGTCAATTTGTACCT-	-GCCAATTCGCGGAAGAAGTG-
<i>HIF-1α</i>	-GAACGTCGAAAAGAAAAGTCTC G-	-CCTTATCAAGATGCGAACTCA CA-
<i>SLC39A8</i>	-ATGCTACCCAAATAACCAGCTC-	-ACAGGAATCCATATCCCCAAA CT-
<i>SOD2</i>	-GGAAGCCATCAAACGTGACTT-	-CCCGTTCCTTATTGAAACCAA GC-
<i>HMOX1</i>	-AAGACTGCGTTCCTGCTCAAC-	-AAAGCCCTACAGCAACTGTCTG -
<i>DMT1</i>	-CGGGTGCTGGAGCTATACAG-	-CGACAGTGTTGACATCAATGG C-
<i>PTPRC</i>	-ACCACAAGTTTACTAACGCAAG T-	-TTTGAGGGGGATTCCAGGTAA T-
<i>IL6R</i>	-CCCCTCAGCAATGTTGTTTGT-	CTCCGGGACTGCTAACTGG-
<i>LYN</i>	-TGCAGAGGGAATGGCATAACAT-	-TGA CT CGGAGACCAGAACATT AG-
<i>CSF1R</i>	-CTGCCAGATCGTGTGCTC-	-AGGTTGAGGGTCAGGACTTTT T-
<i>GAPDH</i>	-CGGGAAGCTTGTCATCAATGG-	-GGCAGAGATGATGACCCTTT-
Mouse		
<i>Ptprc</i>	-GTTTTTCGCTACATGACTGCACA-	-AGGTTGTCCA ACTGACATCTT TC-
<i>Il6r</i>	-GCCACCGTTACCCTGATTTG-	-TCCTGTGGTAGTCCATTCTCTG -

Categories	Forward (5'--3')	Reverse (5'--3')
<i>Lyn</i>	-ATTGTGGTGGCCTTATACCCT-	-ACCATCCCCATGCTCTTCTA-
<i>Csf1r</i>	-TGTCATCGAGCCTAGTGGC-	-GGTCCAAGGTCCAGTAGGG-
<i>Gapdh</i>	-AGGTCGGTGTGAACGGATTG-	-GGGGTCGTTGATGGCAACA-
ChIP assay		
DMT1-Site1	-AGGGACCCAGATGAAAGGTCTC TTT-	-GGTTCACGCCTGTAATCCCAG CA-
DMT1-Site2	-TGTTTAAGGTGTACAAGATGGA GG-	-AATCCTTGTAGTAGGCTGGCA C-
DMT1-Site3	-TCCCCTCCTCCCAGATTGAG-	-AGACATCAGAGCCACGCAAA-
Pulldown assay (biotin-labeling)		
DMT1-Site1-probe	-AGGGACCCAGATGAAAGGTCT-	-ACACCTTAAACAATTCCCTTC CATT-
DMT1-Site2-probe	-TGTTTAAGGTGTACAAGATGGA GG-	-AATCCTTGTAGTAGGCTGGCA C-
DMT1-Site3-probe	-AAACTTCTAGAAAAGTGCCTGC-	-TAACCCCAAGCCTGACAGTG-
Luciferase plasmids construction		
DMT1-WT -pGL3-Basic	-AGGTACCGAGCTTTACGCGTAA TTCTCCTGCCTGAGCCTCCCA-	-GCTTACTTAGATCGCAGATCTG GTGCAGAACTAGTTTACTTTC CCAATGC-
DMT1-Mut-1	-ACCACTGTGCCTGGCCGCAA-	-TTGCGGCCAGGCACAGTGGTT CTGTAATCCCAGCACTTTG-
DMT1-Mut-2	-GCTGGGATTGCAGAATTTTGTG CCAGCCTACTAC-	-AAAATTCTGCAATCCCAGCA CTTTGG-
DMT1-Mut-3	-CCAAGTCCGGAGTTTGCTCTGAT GTCTGCTCGTGG-	-AGAGCAAAGTCCGGACTTGGT GC-

62 **Abbreviations:** RT-PCR, reverse transcription-polymerase chain reaction assay; ChIP,
63 chromatin immunoprecipitation; WT, wild type plasmid; Mut, mutant plasmid.

64 **Table S2.** The antibodies used information.

Categories	Usage and dosage	Product codes	Sources
Primary antibodies			
FTH	WB 1:200 IHC 1:100 IF 1:50	sc-376594	Santa Cruz, California, USA
FTL	WB 1:200 IHC 1:100 IF 1:50	sc-74513	Santa Cruz, California, USA
IL-6R	WB 1:200 IHC 1:100 IF 1:50	sc-373708	Santa Cruz, California, USA
HIF-1 α	WB 1:1000 IHC 1:500 IF 1:100 ChIP 2 ug	#36169	CST, Massachusetts, USA
DMT1	WB 1:1000 IHC 1:200 IF 1:100	20507-1-AP	Proteintech, Illinois, USA
CD31	IF 1:100	11265-1-AP	Proteintech, Illinois, USA
p-JAK1	WB 1:1000 IHC 1:200	SAB4300123	SIGMA, Missouri, USA
p-STAT3	WB 1:1000 IHC 1:200	#9145	CST, Massachusetts, USA
ATF6	WB 1:1000 IHC 1:200	24169-1-AP	Proteintech, Illinois, USA
GRP78	WB 1:1000 IHC 1:200	11587-1-AP	Proteintech, Illinois, USA
CHOP	WB 1:1000 IHC 1:200	15204-1-AP	Proteintech, Illinois, USA
Caspase 12	WB 1:500	#58208	CST, Massachusetts, USA
Cleaved-caspase 3	WB 1:500	#9661	CST, Massachusetts, USA
GAPDH	WB 1:10000	10494-1-AP	Proteintech, Illinois, USA
Histone H3	WB 1:2000	17168-1-AP	Proteintech, Illinois, USA
Control Rabbit IgG	ChIP 2 ug	#2729	CST, Massachusetts, USA

Categories	Usage and dosage	Product codes	Sources
Secondary antibodies			
IRDye® 800CW Goat anti-Mouse IgG (H+L)	WB 1:20000	926-32210	LI-COR, Nebraska, USA
IRDye® 800CW Goat anti-Rabbit IgG (H+L)	WB 1:20000	926-32211	LI-COR, Nebraska, USA
Donkey anti-Mouse IgG (H+L) Alexa Fluor™ 488	IF 1:200	AB_2556542	Thermo Fisher Scientific, Massachusetts, USA
Goat anti-Mouse IgG (H+L) Alexa Fluor™ 594	IF 1:200	AB_2556547	Thermo Fisher Scientific, Massachusetts, USA
Donkey anti-RabbitIgG (H+L) Alexa Fluor™ 594	IF 1:200	AB_2556549	Thermo Fisher Scientific, Massachusetts, USA
Goat anti-Rabbit IgG (H+L) Alexa Fluor™ 488	IF 1:200	AB_2556544	Thermo Fisher Scientific, Massachusetts, USA

65 **Abbreviations:** WB, western blot; IHC, immunohistochemistry; IF, immunofluorescence;

66 ChIP, chromatin immunoprecipitation.

67 **Table S3.** The enzyme-linked immunosorbent assay kits used information.

Categories	Product codes	Sources
Human		
Free iron ion (Non-transferrin binds iron)	MM-60946H1	Mmbiology, Jiangsu, China
Interleukin-6	MM-0049H1	Mmbiology, Jiangsu, China
Malonaldehyde	MM-2037H1	Mmbiology, Jiangsu, China
Reactive oxygen species	MM-1893H1	Mmbiology, Jiangsu, China
Kidney injury molecule-1	MM-0647H1	Mmbiology, Jiangsu, China
Creatinine	MM-13833H1	Mmbiology, Jiangsu, China
Mouse		
Free iron ion (Non-transferrin binds iron)	MM-46281M1	Mmbiology, Jiangsu, China
Interleukin-6	MM-0163M1	Mmbiology, Jiangsu, China
Malonaldehyde	MM-0897M1	Mmbiology, Jiangsu, China
Reactive oxygen species	MM-43700M1	Mmbiology, Jiangsu, China
Kidney injury molecule-1	MM-0318M1	Mmbiology, Jiangsu, China
Creatinine	MM-0693M1	Mmbiology, Jiangsu, China

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69 **Table S4.** Clinical details of aortic dissection and other patients.

Variables	AD (n = 67)	Others (n = 70)	p-value
Preoperative variables			
Age (years)	50.0 ± 11.2	50.3 ± 13.0	0.813
Male	46 (68.7%)	57 (81.4%)	0.084
Height (cm)	169.6 ± 7.7	170.4 ± 7.6	0.230
Weight (kg)	74.0 ± 12.2	71.6 ± 11.6	0.244
Body mass index (kg/m ²)	25.7 ± 3.7	24.6 ± 3.4	0.060
Smoking	21 (31.3%)	33 (47.1%)	0.059
Hypertension	44 (65.7%)	35 (50.0%)	0.063
Left ventricular ejection fraction (%)	60.5 ± 4.7	58.1 ± 7.7	0.116
Free iron ion (ug/mL)	7.9 ± 2.0	2.7 ± 0.9	< 0.001
Interleukin-6 (pg/mL)	113.3 ± 39.1	52.4 ± 23.1	< 0.001
Serum creatinine (ug/mL)	101.2 ± 18.4	76.3 ± 15.2	< 0.001
Hepcidin (ng/mL)	21.1 ± 8.2	6.6 ± 4.6	< 0.001
Kidney injury molecule-1 (pg/mL)	139.5 ± 33.5	80.1 ± 25.8	< 0.001
Malondialdehyde (nmol/mL)	5.6 ± 0.8	4.2 ± 0.6	< 0.001
Reactive oxygen species (U/mL)	363.0 ± 79.4	220.2 ± 60.8	< 0.001
Operation-related variables			
Length of operation (hour)	7.5 ± 1.7	5.8 ± 1.1	< 0.001
Cardiopulmonary bypass time (min)	187.2 ± 46.6	136.9 ± 41.2	< 0.001
Cross-clamp time (min)	105.3 ± 28.5	89.8 ± 27.5	0.003
Assisted circulation time (min)	73.9 ± 28.4	39.0 ± 21.7	< 0.001
Postoperative variables			
Packed red cells on first 24h (units)	1.9 ± 2.2	0.7 ± 1.9	< 0.001
Frozen plasma on first 24h (100 mL)	0.9 ± 1.9	0.4 ± 1.3	0.067
Platelet concentrate on first 24h (units)	0.1 ± 0.3	0	0.006
Packed red cells (units)	4.3 ± 5.7	1.2 ± 3.6	< 0.001

Variables	AD (n = 67)	Others (n = 70)	p-value
Postoperative variables			
Fresh frozen plasma (100 mL)	1.6 ± 3.6	0.7 ± 2.6	0.049
Platelet concentrate (units)	0.4 ± 0.8	0.1 ± 0.7	< 0.001
Hospital length of stay (day)	17.3 ± 7.3	16.9 ± 6.3	0.799
ICU length of stay (hour)	98.8 ± 83.2	38.2 ± 45.6	< 0.001
Prolonged ICU of stay (≥ 3 days)	32 (47.8%)	7 (10.0%)	< 0.001
Mechanical ventilation (hour)	38.5 ± 48.4	27.6 ± 49.8	0.004
Mechanical ventilation (≥ 24 hours)	35 (52.2%)	8 (11.4%)	< 0.001
In-hospital mortality	2 (3.0%)	1 (1.4%)	0.534
Renal replacement therapy	5 (7.5%)	0	0.061
Acute kidney injury	28 (41.8%)	13 (18.6%)	0.003
Acute kidney injury			0.006
None	39 (58.2%)	57 (70.1%)	
Stage 1	15 (22.4%)	10 (14.3%)	
Stage 2	6 (9.0%)	3 (4.3%)	
Stage 3	7 (10.4%)	0	
Extracorporeal membrane oxygenation	0	2 (2.9%)	0.497
Stroke	7 (10.4%)	1 (1.4%)	0.059
Paraplegia	3 (4.5%)	0	0.228
Re-exploration for bleeding	2 (5.0%)	2 (2.9%)	0.228
Serious adverse events	42 (62.7%)	11 (15.7%)	< 0.001

70 **Footnote:** Variables are displayed as n (%) and mean ± standard deviation. The p-values with
71 statistical significance are bolded.

72 **Abbreviations:** ICU, intensive care unit.

73 **Table S5.** Univariable logistic regression of serious adverse events.

Variables	OR (95% CI)	<i>p</i>-value
Age	1.03 (0.99 - 1.08)	0.151
Male	1.29 (0.44 - 3.80)	0.649
Body mass index	0.99 (0.86 - 1.13)	0.869
Smoking	0.71 (0.25 - 2.04)	0.527
Hypertension	1.49 (0.53 - 4.18)	0.452
Left ventricular ejection fraction	1.01 (0.91 - 1.13)	0.830
Free iron ion	1.69 (1.24 - 2.31)	0.001
Interleukin-6	1.02 (1.00 - 1.04)	0.037
Serum creatinine	1.04 (1.00 - 1.08)	0.049
Hepcidin	1.04 (0.97 - 1.10)	0.266
Kidney injury molecule-1	1.00 (0.98 - 1.01)	0.584
Malondialdehyde	0.90 (0.50 - 1.61)	0.717
Reactive oxygen species	1.00 (0.99 - 1.00)	0.549
Cardiopulmonary bypass time	1.01 (1.00 - 1.02)	0.069
Cross-clamp time	1.01 (0.99 - 1.03)	0.170

74 **Footnote:** Preoperative and operation-related variables with *p*-values < 0.20 in univariable
 75 logistic regression were considered candidates for multivariable regression. The *p*-values with
 76 statistical significance are bolded.

77 **Abbreviations:** OR, odds ratio; CI, confidence interval.