

Review

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# Bridging Chronic Inflammation and Digestive Cancer: The Critical Role of Innate Lymphoid Cells in Tumor Microenvironments

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#### Abstract

The incidence and mortality of digestive system-related cancers have always been high and attributed to the heterogeneity and complexity of the immune microenvironment of the digestive system. Furthermore, several studies have shown that chronic inflammation in the digestive system is responsible for cancer incidence; therefore, controlling inflammation is a potential strategy to stop the development of cancer. Innate Lymphoid Cells (ILC) represent a heterogeneous group of lymphocytes that exist in contrast to T cells. They function by interacting with cytokines and immune cells in an antigen-independent manner. In the digestive system cancer, from the inflammatory phase to the development, migration, and metastasis of tumors, ILC have been found to interact with the immune microenvironment and either control or promote these processes. The conventional treatments for digestive tumors have limited efficacy, therefore, ILC-associated immunotherapies are promising strategies. This study reviews the characterization of different ILC subpopulations, how they interact with and influence the immune microenvironment as well as chronic inflammation, and their promotional or inhibitory role in four common digestive system tumors, including pancreatic, colorectal, gastric, and hepatocellular cancers. In particular, the review emphasizes the role of ILC in associating chronic inflammation with cancer and the potential for enhanced immunotherapy with cytokine therapy and adoptive immune cell therapy.

Keywords: Innate lymphoid cells; Inflammation; tumor immune microenvironment; cytokine; immunotherapy; Digestive system

## Introduction

# Tumor immune microenvironment and immunotherapy

Cancer is the second leading cause of death in the United States. In 2023, the estimated number of incidences and deaths by digestive system-related cancers was 348,840 and 172,010, respectively. Furthermore, the highest incidence and mortality rates were significantly attributed to colorectal cancer (CRC) and pancreatic cancer, respectively [1]. Therefore, there is an urgent need for earlier diagnosis as well as more effective treatment modalities. The in-depth investigation of tumor mechanisms and the development of new technologies will offer additional strategies to fight cancer [2-4].

The complexity of the tumor immune microenvironment, along with immune escape and resistance of tumor cells, significantly limits the clinical efficacy of conventional therapies like surgery, chemotherapy, and radiotherapy [5]. Targeting and precision are the new compasses of cancer treatment [6, 7]. Immunotherapies, including modified cytokines, over-the-counter cell therapies, immune checkpoint blockade, and cancer vaccines, have emerged as new options for cancer control. The effectiveness of these immunotherapies is often correlated with prolonged survival and reduced recurrence rates [8, 9]. Cytokines, secreted by inflammatory, tumor, or immune cells facilitate cellular interactions and communication as well as crucial mediators linking inflammation and cancer. Cytokine therapy aims to enhance growth-inhibitory and immunostimulatory cytokines such as interferon (IFN) and Interleukin-2 (IL-2), IL-7, IL-12, and IL-15. Furthermore, it also inhibits inflammatory and tumor-promoting cytokines like tumor necrosis factor (TNF), IL-1 $\beta$ , and IL-6 [10]. The cytokine therapies are continuously advancing, with IL-17, emerging as a key drug target for various chronic inflammatory diseases because of its modulatory effect on inflammation, resistance to pathogens [11], and therapeutic potential against numerous types of solid tumors [12, 13]. Innovations in cytokine construction methods and delivery modalities are further revitalizing cytokine therapy. The use of antibody-cytokine fusion molecules in conjunction with chemotherapy has proven effective in treating CRC [14, 15]. Moreover, the delivery capabilities of pluripotent stem cells and biomaterials for cytokines are under active development [16]. Recently, an innate lymphocyte closely related to cytokines has been discovered to be associated with the dynamic progression from inflammation to tumor.

# Role of ILC in progression from inflammation to cancer stage in the digestive system

Innate lymphoid cells (ILC) are a family of immune cells found throughout the mucosal tissues of the gastrointestinal tract, pancreas, and liver. They help the body build a first line of defense against external stimuli. Unlike T cells, ILC lack the antigen-specific T cell receptor (TCR) and functions primarily by secreting cytokines and interacting with other immune cells. Based on the secretion of characteristic cytokines and dependent transcription factors, ILC are divided into three groups, including group I ILC [comprising Natural killer (NK) cells and type 1 ILCs (ILC1s)], and groups ILC2s and ILC3s. NK cells and ILC1s are dependent on T-bet and Eomes, respectively, and characteristically express IFN-y and granzyme [17, 18], while ILC2s are dependent on GATA-3 and express IL-5, IL-9, IL-13 and amphiregulin (AREG) [19, 20]. The ILC3s group depends on RORyt and secretes IL-17 and IL-22 respectively depending on the phenotype [21]. ILC play a pivotal role in the regulation of the immune system and the maintenance of tissue homeostasis. ILC that secrete cytokines are involved in numerous immune pathways, including the control of immune responses to commensals and pathogens at the mucosal barrier, the enhancement of adaptive immunity, and the modulation of inflammatory responses. Comprehensive analysis of these cells revealed that they play different or even opposite roles in the dynamic procedure of inflammation to tumors in the digestive system [22-25]. This contradiction might be because ILC are closely connected to their surroundings and have a high degree of plasticity [16]. At different sites and stages, ILC are molded into distinct subpopulations that secrete different cytokines or exhibit specific interactions with surrounding cells.

Inflammation is primarily a defense mechanism of the body against external stimuli to maintain homeostasis. During acute inflammation, the inflammatory response initiates the repair by secreting inflammatory molecules and facilitating chemotaxis of inflammatory cells, such as macrophages and neutrophils, toward the site of injurv for detoxification. when However, inflammatory molecules are overexpressed or signaling pathways related to anti-inflammatory responses and inflammation reduction are impaired, external stimuli are not eliminated within a short time, and ultimately the duration of inflammation extends, leading to the transition to chronic inflammation. This transition has been found to regulate cancer via different mechanisms such as chemotaxis, tumorigenesis, proliferation, vascular network formation, as well as invasion and metastasis. It has been observed that in the digestive system, controlling chronic inflammation is closely with prevention, suppression associated of progression, and better prognosis in colorectal, pancreatic, gastric, and liver cancers [26-29]. Furthermore, chronic inflammation is an important target for cancer treatment and prevention [30]. In addition to damaging DNA, promoting cell proliferation, and modulating signaling pathways, the tumor-promoting effects of chronic inflammation also depend on the construction of a suppressive tumor immune microenvironment. With the accumulation of

immunosuppressive inflammatory factors [31], immunosuppressive cells are recruited [32], immune cells are then inhibited [33], and differentiated into immunosuppressive phenotype [34], ultimately the function of the inflammatory response is slowly shifted from defense and repair to cancer promotion. Currently, the studies on the digestive system-associated crosstalk between chronic inflammation and cancer have been focused on the effect of Helicobacter pylori (Hp) and related inflammation on gastric cancer, fibrosis in pancreatic and hepatocellular cancers, and the inflammatory response that damages the mucosal barrier of the tract and gastrointestinal promotes cellular chemotaxis [35-39]. Considerable research suggests that ILC are associated with the key nodes described above by producing cytokines or engaging in communication with specific cells. Chronic inflammation and its related cytokines, ILC, and the tumor immune microenvironment form a network. Effective control of ILC are beneficial for limiting the inflammatory response toward inhibiting digestive system tumors. This paper discusses the role of ILC in digestive system inflammation to cancer progression, specifically CRC, gastric, liver, and pancreatic cancers. It was observed that the interaction between ILC, cytokines, and other immune cells is the key underlying mechanism.

## The role of ILC subsets

## NK cells and ILC1s

The closely related ILC1s and NK cells together constitute the first group of ILC. These cells co-express IFN-y and granzyme, depend on transcription factors Eomes and T-bet for growth and development, and have similar surface markers and functional patterns. In contrast to ILC1s, NK cells do not depend on the transcription factor T-bet, instead, they require transcription factor Eomes for the maintenance of the identity and function [40]. NK cells participate in the circulation and exert their potent cytotoxicity, playing an important role in innate immunity. Contrary to earlier beliefs that ILC1s are tissue-resident and non-cytotoxic, primarily modulate T helper type 1 (Th1) cells by secreting cytokines such as IFN-y in response to Th1-like immune responses, recent studies have revealed that IL-15 mediates granzyme C expression and ILC1s cytotoxicity in a mouse breast cancer model, suggesting that ILC1s may also be cytotoxic [41]. ILC1s and NK cells have been progressively characterized, and their tissue specificity has also been identified [42-44].

IFN- $\gamma$  and TNF- $\alpha$  secreted by ILC1s are effective

against multiple exogenous infections, such as cytomegalovirus, *Toxoplasma gondii*, and vaccinia virus [17, 45, 46]. However, the excess production of IFN- $\gamma$  by ILC1s can exacerbate colitis in Crohn's disease mice [47]. Furthermore, intraepithelial lymphocytes contribute to the development of intestinal inflammation by supporting the survival of ILC1-like cells [48].

Whether NK cells specifically promote or inhibit cancer is closely correlated with their specific phenotype [49, 50]. The data indicate that NK cell infiltration significantly modulates in solid tumors [50]. Differing from CD56dim NK cells, which are predominantly circulating and have a potent cytolytic capacity with killer immunoglobulin-like receptors, CD56<sup>bright</sup> NK cells are primarily found in secondary lymphoid organs and tissues and play а pro-inflammatory role by secreting IFN-y, granulocyte-macrophage colony-stimulating factor (GM-CSF), TNF, and IL-10 [51]. In some tumors, NK cells exhibit a specific phenotype resembling the tissue-resident NK (trNK) cell population. Among them, decidual NK (dNK) cells have been observed to release pro-angiogenic factors and altered effector functions that could potentially induce cancer growth [52]. Another trNK-like tumor-infiltrating NK cell with antitumor functions has been identified as intraepithelial ILC1s (ieILC1s), which have been associated with malignancy suppression and can be stimulated by IL-12 and IL-15 to produce IFN-y [53].

Cytokines and a high degree of plasticity further complicate the role of group I ILC in tumorigenesis, especially due to the ease of conversion of NK to ILC1s or some other immunocompetent phenotype. IL-15 and transforming growth factor- $\beta$  (TGF- $\beta$ ) are strongly associated with the plasticity of group I ILC. For instance, in breast precancerous lesions as well as head and neck squamous cell carcinoma models, IL-15 mediates the production of granzymes by CD103+ cytotoxic ILC1s and promotes the proliferation of ieILC1s, which may inhibit cancer [54, 55]. Another important cytokine, TGF- $\beta$ , predominantly plays a pro-tumorigenic role via the NK cells to ILC1s transition [56]. TGF- $\beta$  can mediate the conversion of NK cells to intermediate type 1 innate lymphoid cell (intILC1s) populations and ILC1s populations, thereby enabling tumors to evade innate immune system surveillance [57]. This transition is particularly evident in CD56<sup>bright</sup> NK cells and can be augmented by IL-5, further highlighting the role of ILC's plasticity in their specific functions [58]. Furthermore, ILC1s can differentiate into other ILC subtypes and cause different outcomes, as evidenced in squamous cell carcinoma and hepatocellular carcinoma (HCC) models [59, 60].



Figure 1: ILC and Chronic Inflammation: Shaping Tumor Immunity and Therapy. ILC and chronic inflammation synergistically shape the tumor immune microenvironment, jointly promoting or inhibiting the proliferation, invasion, metastasis, and death of tumors. The progression of chronic inflammation to cancer is regulated by ILC. Furthermore, chronic inflammation also modulates the types of ILC and influences their function. Because of their high degree of plasticity and unique function in various phenotypes, targeting ILC with immunotherapy offers a promising strategy for cancer prevention and control.

NK cells and ILC1 exert specific effects on the body in different microenvironments. ILC1s have a more complex type and function than NK cells in the traditional sense of killing cells and are associated with but not limited to factors like IL-15 and TGF- $\beta$ . A more systematic phenotyping of these cells and the effect of cytokines on their plasticity will potentially offer insights into new treatments for cancer or inflammation.

## ILC2s

Transcription factor GATA-3 is a key factor that aids ILC2s in differentiation, maintenance, and function [61]. Similar to group 1 ILC, the phenotype and function of ILC2s are tissue-specific [62]. ILC2s are not only tissue-resident but can also participate in innate type 2 immune and inflammatory responses in the circulation [63]. As the counterpart of Th2 cells, ILC2s produce type 2 cytokines (IL-4, IL-5, IL-9, and IL-13) and are implicated in parasite immunity, airway inflammatory diseases, tissue repair, and tumor regulation. Furthermore, ILC2s are crucially associated with different mechanisms of immune defense against various parasites such as by pathogen efflux facilitating and migrating inflammation to lymphoid tissues [64, 65]. Moreover, ILC2s are closely associated with allergic airway diseases such as asthma, and their independent secretion of IL-5 and IL-13 confirms their important role in airway inflammation [66]. Transcription factor RORy induced ILC2 secretion of IL-17A, IL-17F, and IL-22 in chronic airway inflammation [67]. IL-2 signaling enhances ILC2 survival and proliferation and aids in the production of type 2 cytokines to promote eosinophilic pneumonia [68]. It has been observed that ILC2's functions are modulated by their specific environment. In addition, ILC2s are not only modulated by cytokines but certain lipid molecules have also been observed to play a significant role. IL-4 and IL-33 work synergistically in mediating ILC2 proliferation and promoting type 2 cytokine production [69]. Whereas, IFN- $\gamma$  and IL-27 negatively regulate tissue-resident ILC2s [70]. Studies have shown that prostaglandin E2 can inhibit ILC2 activation [71]. ILC2s are also instrumental in tissue repair. Increased levels of IL-33 in wounds increase ILC2 cells, thereby enhancing wound healing in mice [72].

The tumor microenvironment plays a crucial role in determining the differential impact of ILC2s on tumors. Moreover, IL-33 is a key mediator of most ILC2s-associated tumor immunity and ILC2s can even play an opposite role [73-76]. Other than cytokines, ILC2s rely on interactions with other immune cells in the tumor microenvironment. ILC2s have also been observed to interact with other immune cells, such as in airway inflammation, they can interact with T cells, DC, macrophages, mast cells, or basophils, whereas in cancer, they may interact with myeloid-derived suppressor cells (MDSC), regulatory T cells (Tregs), eosinophils, and neutrophils. Tregs and MDSC are recognized as key immunosuppressive cells that play a negative role in cancer defense. The suppressive function of Tregs is enhanced by ILC2s production of AREG protein, promoting EGFR-expressing tumor progression [77]. IL-13-producing ILC2s may further promote immunosuppression from MDSCs during advanced stages of breast cancer metastasis [78]. IL-33-induced amplification of ILC2s inhibits the activation of NK cells and their cytotoxic effects, thereby promoting tumor growth [79]. Overall, although various significant mechanisms of ILC2s have been extensively studied, many details still require further exploration to fully understand their bidirectional role in various diseases.

## ILC3s

ILC3s, which mirror Th17/Th22 cells, consist of several components: NCR+ ILC3s, NCR- ILC3s, and lymphoid tissue inducer (LTi)/LTi-like cells. In mice, NCR<sup>+</sup> ILC3s are NKp46<sup>+</sup>, whereas in humans, NCR<sup>+</sup> ILC3s are NKp44+. ILC3s in mice produce both IL-17 and IL-22, whereas in humans NKp44- ILC3s secretes IL-17 and NKp44<sup>+</sup> ILC3s produce IL-22. The development of NCR+ILC3s and LTi cells as well as cytokine production, significantly rely on RORyt [80]. Other than survival, RORyt and RORa are intrinsically linked with the functional maintenance and proliferation of ILC3s [81]. The Aryl hydrocarbon receptor (AHR) is another crucial transcription factor for ILC3s, which interacts with the Notch signaling pathway. A deficiency in AHR leads to fewer RORyt+ ILC with diminished IL-22 production, thus inhibiting protection against intestinal bacterial infections [82]. ThPOK is a novel key factor that regulates NKp46+ ILC3s differentiation and is significantly associated with the maintenance of ILC3s function and lineage stability [83]. LTi cells are integral in secondary lymphoid organ formation through Lymphotoxin  $\beta$ -receptor signaling, a feature absent in LTi-like cells [84]. Moreover, ILC3s can also undergo conversion to other ILC subpopulations. Recent research indicates that T-bet promotes the transformation of human ILC3s into functional NK cells [85].

ILC3s are associated with gastrointestinal homeostasis, inflammatory diseases, and cancer, and their function is dependent on bacteria. In the gastrointestinal tract, ILC3s are primarily involved in barrier repair and the inhibition of bacterial translocation, contributing to the maintenance of homeostasis. In addition, ILC3s strengthen the gastrointestinal barrier by secreting IL-13 [86]. They also eliminate circulating microorganisms by inducing the production of antimicrobial peptides in the liver, which aids in postoperative liver The high-fat regeneration [87]. diet-induced hepatocyte apoptosis can be mitigated by the upregulation of hepatic lipid metabolism following the macrophage-induced proliferation of ILC3s via IL-1 [88]. IL-22, secreted by ILC3s, is crucial in maintaining intestinal barrier function and facilitating interactions with bacteria. IL-22-mediated mucus multiplication provides an early defense against pathogens [89]. Short-chain fatty acids (SCFA) play a crucial role in intestinal mucosal homeostasis and ILC3s in the colon have numerous SCFA receptors on their surface. The congenital absence of these receptors impacts ILC3s' proliferation and IL-22 production, which reduces immunity against colony

infections and aids tissue repair [90, 91]. Whereas when the secretion of IL-22 by CD4+ T cells and ILC3s is enhanced by SCFA, intestinal infections can be better controlled [92]. The relationship between ILC3s and intestinal flora is bidirectional and contact with the intestinal flora is essential for the development and function of ILC3s. It has been observed that SCFAs are a major metabolite of microorganisms which has indicated that microorganisms can activate ILC3s through direct contact. Tryptophan, a digestive product of the intestinal microbiota and a ligand for the AHR, supplies energy to Lactobacillus spp. Furthermore, AHR is a key transcription factor that supports the function and differentiation of ILC3s. Microbiota can also activate ILC3s directly, such as by mediating IL-1 $\beta$  and IL-23.

The literature has indicated that inflammation crucially modulates the association between inflammatory bowel disease (IBD) and ILC3s. During inflammation, ILC3s play a protective role and support the intestinal barrier; however, chronic convert inflammation ILC3s can into а pro-inflammatory phenotype. In this state, they express additional inflammatory factors, creating a positive feedback loop that exacerbates intestinal inflammation [93]. Apart from its protective role against intestinal bacterial infections, IL-22 may also have detrimental effects on IBD. In an anti-CD40 antibodies-induced acute congenital colitis model, IL-22 secreted by ILC3s was found to aggravate inflammation [94].

ILC3s have different impacts on tumors, including direct contact, cytokine production, and the formation of tumor-associated tertiary lymphoid structures (TLS), attributed to their significant immune recruitment capacity. Furthermore, ILC3s have been shown to exert antitumor effects on HCC and melanoma cells via IFN-y release and Tumor necrosis factor-related apoptosis-inducing ligand (TRAIL)-dependent cytotoxicity stemming from direct recognition [95]. Whereas IL-22 from ILC3s has been linked with cancer promotion. In а colitis-associated carcinoma mouse model, IL-22 induced epithelial signal transducer and activator of transcription 3 (STAT-3) phosphorylation and proliferation, thereby facilitating cancer development [96]. The NLRP3 and NLRP6 inflammasomes are known to reduce IL-22 binding protein levels, leading to an uncontrolled increase in IL-22 and subsequent stimulation of colon cancer [97]. Moreover, ILC3s have also been observed to promote cancer via the IL-23/IL-17 axis. IL-23 activates ILC3s, contributing to intestinal carcinogenesis even in the absence of carcinogens [12]. IL-17 enables tumors to develop resistance to vascular endothelial growth factor

inhibitors, further promoting angiogenesis and tumor growth [98]. Similar to secondary lymphoid organs, LTi cells have been observed to mediate TLS formation and positively influence survival and prognosis [99]. LTi cell formation is associated with immune recruitment, which broadly influences the function of ILC3s. Upon activation by IL-12, ILC3s, various chemokine receptors, expressing can potentially inhibit tumor growth by recruiting numerous immune cells such as CD8+ T cells, activated MDSC, NK cells, and natural killer T (NKT) cells. However, this is countered by plasmacytoid ILC3 apoptosis pre-DC inducing via the CD95-mediated pathways [100]. This recruitment unintentionally mav also support cancer development. In addition, the recruitment of NKp44-ILC3s, Tregs, and MDSC by Chemokine ligand 21 (CCL21) promotes the establishment of tumor-supportive environment [101]. The specificity of ILC3s depends on the type of cancer, the stage of tumor development, and the characteristics of the immune microenvironment. Therefore, because of ILC3s' intricate crosstalk with various microbiomes and the environments they inhabit, they are a potential therapeutic target.

# ILC act as a bridge between inflammation and tumor

# Prolonged inflammation can also mark the beginning of cancer

Inflammation is a protective process of the body against external stimuli such as infection or injury, which repairs the damage and limits infection. Inflammatory responses are categorized into acute and chronic inflammation depending on their duration. Acute inflammation is characterized by the migration of macrophages and neutrophils to the site of injury and the secretion of chemokines and cytokines, which ultimately leads to the removal of exogenous substances and healing of the injury. This phase is characterized by a rapid and intense inflammatory response, and after the inflammatory cells have completed their mission, they are either eliminated or undergo apoptosis [102]. To facilitate this resolution, specialized pro-resolving mediators inhibit neutrophil tissue infiltration, mediate apoptosis, remove stimuli, and modulate cytokine secretion [103].

However, not all inflammation can be controlled and restored to homeostasis. Chronic inflammation is an excessive and deleterious response that initiates for self-defense and repair and is characterized by the accumulation of M2 macrophages, MDSC, Treg cells, and inflammation-suppressing cytokines, which establishes an immunosuppressive microenvironment. Although the involved mechanisms are still being investigated, the link between chronic inflammation and tumor development is indisputable. Chronic inflammation supports tumor development by damaging DNA, promoting cell proliferation, and modulating signaling pathways, thereby contributing to tumor progression, distant metastasis, and resistance to therapy. Furthermore, the DNA damage caused by inflammation further stimulates inflammation, creating a positive feedback loop that increases the frequency of mutations. In addition, inflammation-induced cell proliferation and migration support tumor development [104]. Inhibition of inflammatory response via peroxisome proliferator-activated receptors is a promising therapeutic strategy that inhibits tumor cell growth and proliferation [105]. The nuclear factor KB (NF-κB)-related signaling pathway is a bridge between inflammation and cancer and is associated with many cancer-related genes. Moreover, it has various roles, such as induction of pro-proliferative and anti-apoptotic genes, modulation of other signaling pathways, influencing genetic, epigenetic, and cellular metabolic changes, promotion of epithelial-to-mesenchymal transition, tumor invasion, and angiogenesis support, cancer metastasis, and even treatment tolerance and immunosuppression [106, 107]. The zinc finger transcription factor Miz1 is a newly identified tumor suppressor that inhibits liver tumorigenesis, possibly by isolating oncoprotein metadherin, thus preventing metadherin from activating NF-KB, and by shifting the macrophage phenotype toward a pro-inflammatory phenotype [108].

# ILC shape immune microenvironment to modulate the progression of inflammation to cancer

The link between inflammation and cancer extends beyond the above modalities, and their progression is significantly linked with induced immunosuppression. Chronic inflammation can create an immunosuppressive microenvironment either by increasing immunosuppressive inflammatory factors, enhancing the concentration of immunosuppressive cells, reducing the efficacy of immune cells, or by promoting the differentiation of otherwise neutral immune cells towards an immunosuppressive phenotype. This chronic inflammation-induced immunosuppressive microenvironment is either suppressed or enhanced by ILC, potentially because of its tight association with cytokines and immune cells. Cytokines specifically secreted by various ILC subpopulations have been indicated to be involved in the progression from chronic inflammation to cancer. It has been observed that NK cells and their secretion of IFN-y inhibit the progression from chronic pancreatitis to pancreatic cancer by inhibiting hepatic stellate cells (HSC) and the fibrosis they cause [39, 109, 110]. The IgA response due to ILC2s-induced IL-5 is important for the clearance of HP and the prevention of gastritis transformation to gastric cancer [35, 111]. ILC3s and their secretion of IL-22 have been shown to induce the progression of chronic inflammation to colon cancer and association with the perpetuation of colon cancer [96]. Moreover, other than cytokines, ILC significantly regulates the tumor immune microenvironment by interacting with immune cells. These crosstalks are multifaceted, specifically in immune cell recruitment, influencing immune cell activity and function, and shifting the direction of immune cell polarization.

Chemokines are widely upregulated as a specific type of cytokine in the inflammatory response. Furthermore, chemokines mediate the migration of immune cells involved in the development of anti-tumor immunity or induce a pro-tumor microenvironment, Moreover, they are also recognized as biomarkers with diagnostic effects that can predict efficacy and prognosis [112]. For instance, CCL15 can recruit monocytes, eosinophils, and neutrophils, is a prognostic marker for HCC, and is associated with metastasis [113-115]. In head and neck squamous cell carcinoma, CCL15 induces gefitinib resistance via NF-KB [116]. Increased expression of CCL-16 by centrosomal P4.1-associated protein underpins inflammatory pathways and HCC [117]. Chemokines recruit ILC and their recruitment of immune cells is also important for shaping the tumor immune microenvironment. CCL2 induces the recruitment of ILC3s to tumors and stimulates the production of CXCL13 by tumor stromal cells, which ultimately leads to the production of the cancer cell motility factor receptor activator of NF-KB ligand and promotes lymphatic metastasis of breast cancer cells [118]. However, in the breast cancer tumor microenvironment, CX3CL1 overexpression attracts NK cells, which then inhibits tumor growth and metastasis, and thus improves the efficacy of trastuzumab in the treatment of low-expressing human epidermal growth factor receptor 2 cancers [119]. IL-21-stimulated NK cells can secrete more chemokines and recruit activated T cells, thus enhancing the therapeutic effect of pancreatic cancer [120]. Similarly, the recruitment of DC by ILC2s has been observed to activate T-cell immunity and suppress pancreatic cancer [77]. CXCL2 derived from ILC2s recruits neutrophils and promotes HCC progression [121]. GM-CSF and IL-5 produced by

ILC2 exert anti-melanoma effects by recruiting eosinophils [122].

The effects of ILC on the activity and function of immune cells are also crucial for the tumor immune microenvironment. T cells and DCs, which are antigen-presenting, play an important role in fighting cancer and modulating inflammation. However, chronic infections and cancer lead to the production of depleted T cells with reduced proliferative capacity and effector function [123, 124]. Several studies have suggested that highly correlated ILC may be important in regulating T-cell immunity. NK cells have multiple pathways for DC recruitment, promotion of DC maturation, killing of immature DC, and inhibition of DC crossing over to enhance the antitumor effect of T cells [125-128]. IL-9 secreted by ILC2s correlates with T-cell activation and CRC suppression [129]. Furthermore, IL-13 secreted by ILC2s activates MDSCs and subsequently negatively regulates T-cell proliferation [130]. The T cells exerting anti-tumor effects as described above are CD8<sup>+</sup> T cells, while Treg cells are a specific type of immunosuppressive T cells, that also interact with ILC. The enhanced Treg cell function induced by ILC2s secreted AREG can markedly promote CRC progression [131]. In contrast, secretion of GM-CSF by ILC3s can reduce the number of Tregs in the gut [132]. It is well established that MDSC acts as one of the major players in suppressing tumor immunity [133]. In addition to attracting MDSC to the tumor immune microenvironment, ILC can also enhance MDSC function. ILC3s can be shaped into secreted ILCreg to further enhance the recruitment effect while secreting IL-17 and IL-22 for MDSC recruitment [134]. IL-13-producing ILC2s may further promote MDSC-mediated immunosuppression during advanced stages of breast cancer metastasis [78].

It has been observed that in addition to Treg and mast cells (immunosuppressive) or NK and CD8+ cytotoxic T lymphocytes (immune-promoting), the role of some immune cells, such as macrophages, neutrophils, and DC might be polarized, and depends on chronic inflammation [135]. Specifically, the conversion of macrophages to the M1 phenotype is associated with inhibition of tumor progression, while the shift to the M2 phenotype suggests tumor growth and invasion. This data reveals the possibility of using macrophages to regulate cancer development and regression [136, 137]. Different ILC subpopulations are involved in their polarization and may influence the direction of polarization. Studies have shown that NK cells are beneficial in controlling fibrosis, an important step in hepatitis leading to HCC, possibly by polarizing macrophages to an M1 phenotype during the hepatitis phase [138]. Furthermore,

adipose-resident ILC1 populations can drive the shift of macrophages toward a pro-inflammatory M1 phenotype [139]. Increased M2-type macrophages caused by ILC2s mediate an immunosuppressive microenvironment in gastric cancer [140]. GM-CSF derived from ILC3s promotes macrophage polarization toward the M1 phenotype and stimulates self-activation through a positive feedback loop, helping to maintain intestinal homeostasis and inhibit fibrosis [141]. In turn, macrophage polarization influences ILC function. IL-1ß secreted by M1-type macrophages determines the production of ILC2-derived IL-22, which contributes to the repair of inflammatory injury [142].

Overall, the bridge between inflammation and cancer is built by the constant interaction of ILC with cytokines and immune cells. Moreover, ILC themselves are shaped in their process of shaping the tumor immune microenvironment.

The following section will elaborate on how ILC regulate the progression of inflammation to cancer in the digestive system.

# Clues from digestive system inflammation to tumor-ILC

#### Pancreatic disease

Pancreatic adenocarcinoma, mainly pancreatic ductal adenocarcinoma (PDAC), is characterized by malignant features such as early metastasis, which poses a challenge for early diagnosis, and limits therapeutic efficacy. Despite the advent of chemotherapy, surgery, immunotherapy, targeted therapy, and combination therapy, its high incidence and the fact that it is still the fourth most common cause of cancer mortality underscore the importance of continued and in-depth study [1]. Chronic pancreatitis progressive, irreversible, is а multifactorial fibroinflammatory disease. Furthermore, it has been observed that chronic pancreatitis-mediated inflammatory response and metaplasia of alveolar cells to the ducts are crucial mediators of the transition from pancreatitis to pancreatic cancer. The previous literature was primarily focused on oxidative stress, cytokines, shared inflammatory signaling pathways, and the transformation of pancreatic stellate cells and alveolar cells [143-145]. However, the importance of ILC was observed after multiple ILC phenotypes were identified in pancreatic diseases.

The largest proportion of NK cells are active in both pancreatic cancer and pancreatitis. Activated CD56<sup>+</sup> NK cells are clustered in the pancreas of patients with chronic pancreatitis, and NK cells directly correlate with the benign outcomes of survival and recurrence in patients after pancreatic cancer surgery [146, 147]. NK cells can induce cancer stem cell (CSC) differentiation and inhibit pancreatic tumor growth and metastasis by secreting IFN-y and TNF- $\alpha$  [148]. IL-21 enhances the antibody-dependent cell-mediated cytotoxicity (ADCC) of NK cells and stimulates their secretion of IFN-y and chemokines for recruiting activated T cells, and its combination with cetuximab reduces tumor load in mice [120]. IL-2 is an important anti-tumor growth factor. The targeted immune cytokine L19-Interleukin-2 (L19-IL2) induces extensive necrosis and proliferation restriction in pancreatic cancer with the indispensable role of NK cells [149]. Furthermore, IL-15 has been observed to induce NK cell growth and proliferation and exert antitumor activity [150]. Moreover, NK cells and recombinant IL-15 (rIL-15) can inhibit fibrosis in chronic pancreatitis, a key target for pancreatitis and pancreatic cancer treatment, by inhibiting the atrophy alveolar cells, preventing tissue collagen of accumulation around blood vessels, and regulating pro-fibrotic genes [36, 151]. IL-15 in combination with CD40 agonists showed significant pancreatic cancer suppression, which was also associated with NK cells [152]. ILC2s are another major mediator in pancreatic cancer with a dual nature predominantly dependent on its activator, IL-33. On one hand, IL-33 induces ILC2s-mediated recruitment of DC and subsequent recruitment and activation of CD8<sup>+</sup> T cells, and on the other hand, ILC2s also have the ability to enhance the blockade of the programmed cell death protein-1 (PD-1) pathway. Therefore, immunotherapies co-targeting ILC2s and T cells warrants further research [74]. However, intra-tumoral fungi mediate IL-33 secretion by PDAC cells, and genetic defects in IL-33 or antifungal treatment inhibit pancreatic tumor growth and support prolonged mouse survival [73]. This paradox in turn may be related to the high plasticity of ILC2s. Recently, it was observed that under hypoxic conditions, ILC2s transform into immunosuppressive ILCreg. In a subcutaneous PDAC tumor mice model, the injection of hypoxia-treated ILC2 cells significantly increased tumor volume and subsequent peritoneal metastasis. Through RNA-seq sorting of ILC2s from hypoxic mice at various time points, researchers concluded that hypoxia induces reversible differentiation of ILC2s into high IL-10 expressing ILCreg, and this immunosuppressive effect can be counteracted by neoadjuvant AG (nab-paclitaxel + gemcitabine) [34]. In addition, IL-22 secreted by ILC3s mediates AKT signaling and promotes pancreatic cancer progression and metastasis, whereas concomitant IL-22/IL-22R or AKT blockade has also been shown to be effective [153]. However, the presence of intra-tumoral TLS

promotes better prognosis for many tumors, including pancreatic cancer, and targeting TLS formation might also be a novel idea for immunotherapy [154]. Previous data have confirmed that the levels of NCR<sup>+</sup> ILC3s in non-small cell lung cancer are positively correlated with disease stage and are associated with the density of TLS in the tumor [155]. Therefore, targeting ILC3s may inhibit the induction of TLS formation in pancreatic cancer.

Altogether, under the mediation of various triggers, ILC play a crucial role in the prolongation of chronic pancreatitis as well as the development and regression of pancreatic cancer. This is achieved through the secretion of cytokines, regulation of signaling pathways, enhancement of self-efficacy, or chemotaxis of other immune cells. These multifaceted interactions highlight the complexity and significance of ILC in the pathophysiology of pancreatic diseases.

#### **Intestinal diseases**

In the United States, both new cases and deaths related to CRC have been ranked third among all cancers [1]. Colon cancer is a disease that is highly influenced by external environmental and genetic factors. The molecular mechanisms of CRC have been partially elucidated, including the most common chromosomal instability, microsatellite instability due to DNA mismatch repair, and the CpG island methylation pathway. However, although understanding the mutation mechanism is crucial, there are various other factors involved. The association between CRC and inflammation has been increasingly recognized [156, 157]. Recent studies have indicated an increased risk of CRC in people with IBD as the inflammatory response has been observed throughout the development of CRC. Firstly, chronic inflammation initiates and promotes tumorigenesis by inducing DNA damage or epigenetic changes. Secondly, inflammation is secondary to tumors, and factors like hypoxiainduced cell death or disruption of the epithelial barrier, as well as invasion of microbial products, contribute to cancer development. Finally, pro-tumorigenic inflammation is often caused by treatment-induced necrotic cells that produce patterns damage-associated molecular [158]. Furthermore, the role of ILC in intestinal inflammation and their crosstalk with intestinal microorganisms have been extensively studied, suggesting that a deeper understanding of ILC could lead to novel approaches in CRC treatment.



Figure 2: Crosstalk Between ILC and Other Immune Cells: Steering Tumor Progression. ILC prefer to be thoroughly integrated into the immune microenvironment in which they are embedded rather than working alone. The crosstalk between ILC and other immune cells is essentially linked with ILC-mediated tumor immune microenvironment construction. By recruiting immune cells, influencing their proliferation and activity, and polarizing neutral immune cells, ILC play an important role in controlling the direction of tumor progression.

The NK cells and ieILC1s make up the largest proportion of innate lymphocytes that infiltrate CRC cells [156]. NK cells have a suppressive effect on intestinal inflammation and CRC; however, the definitive mechanism for this effect remains unclear and might be achieved via other immune cells [159-161]. However, in some cases, the inhibitory function of NK cells against CRC is limited or even reversed, which may be attributed to the specialized phenotype of NK and ILC1 cells and their function being influenced by the tumor microenvironment [162]. The studies have observed that after co-culture with tumor cells, the degranulation capacity and ability to release cytotoxic molecules and IFN-y were decreased in NK cells [163]. NK Cells transform to a dNK phenotype and promote CRC progression by secreting pro-angiogenic cytokines [164]. Moreover, in the intestinal inflammatory environment, ILC1s extracellular matrix remodeling, cause which contributes to fibrosis and tumor growth [165]. In addition, the loss of ILC1s due to T-bet depletion provides a defense against severe colitis in mice [24]. Overall, the current research is not sufficient to draw specific generalizations about the role of group I ILC in CRC.

ILC2s have a dual role in both intestinal inflammation and intestinal tumors. These cells are crucial in protecting the intestinal tract. It has been observed that in the fight against amebic colitis, IL-33-mediated protection requires the presence of ILC2s [20]. Mice lacking ILC2s acquired a higher tumor load, and high expression of ILC2s signature genes in tumors has been associated with a better prognosis in CRC [166]. ILC2s are more abundant in CRC tissues than in adjacent tissues, and their IL-9 derivatives activate CD8+T cells and inhibit CRC growth [129]. Furthermore, AREG may play a role in the interaction between ILC2s and intestinal inflammation. Neurohormone U (NMU) mediates the production of AREG in ILC2s and is associated with the intestinal barrier, inflammatory protection, and anti-parasite activities [167]. However, previous studies have shown that high levels of AREG positively correlate with CRC malignancy. AREG derived from ILC2s suppresses tumor immunity by enhancing the function of Tregs [131]. AREG function has been observed to change as inflammation progresses to cancer, which requires comprehensive research. STAT6 is a molecule that is positively associated with intestinal tumor formation in mice and the progression and poor prognosis of humans. Its level is downregulated in the epithelium after it neutralizes ILC2s-derived IL-13 to modulate intestinal permeability and DS-induced colitis [168]. IL-25 promotes ILC2s in tumors, maintaining tumor-infiltrating MDSC to form an immunosuppressive tumor microenvironment [169]. Moreover, as an activator of ILC2s, IL-33 has been associated with CRC. The blockade of the IL-33/ST2 signaling axis relieves the inhibition of colon cancer [170]. Conversely, IL-33 contributes to intestinal polyposis and tumorigenesis through immune and wound-healing responses mediated by tumor epithelial cells [76]. The IL-33 receptor ST2 is highly expressed in colonic Treg cells and enhances their function [171]. Overall, analyzing the targeting axis from ILC2 activators to products and their branches is necessary to further assess the role of ILC2s in CRC.

As the most prevalent ILC in the intestinal tract, ILC3s play a crucial role in maintaining the intestinal barrier, resisting pathogen infection, and cooperating with other immune cells. It has been noted that IL-22 secreted by ILC3s is crucial for the intestinal barrier and resistance to pathogens; however, they may have negative effects on intestinal inflammation such as IBD. This could be related to epithelial inflammation caused by endoplasmic reticulum stress response, necessitating the fine-tuned regulation of IL-22 to ensure beneficial outcomes [172]. In colon cancer, IL-22 is mainly produced by NCR+ILC3s and then induces STAT-3 phosphorylation and proliferation by interacting with IL-22R on epithelial cells [96]. During intestinal inflammation, IL23-reactive ILC3s increase and secrete IL-17, IL-22, and IFN-y to mediate innate colitis [173]. Furthermore, the signal transduction of microbial products drives IL-23 to stimulate tumor-induced inflammation and IL-17 response, thereby promoting tumor development, which may also be related to ILC3s [12]. The plasticity of ILC3s supports its diverse roles. When ILC3s infiltrate into the tertiary lymphoid structure of the tumor, they differentiate into an ILC1s phenotype, and the subsequent loss of major histocompatibility complex class II and reduced communication with T cells may be key to their reduced immunity [174]. TGF- $\beta$  signal transduction initiates the transformation from ILC3s to ILCreg [175]. Therefore, constructing a more detailed ILC-cytokine-tumor immune microenvironment network is essential to identify suitable therapeutic targets.

## Gastritis and gastric cancer

Gastric cancer, predominantly adenocarcinoma, is a prevalent malignancy in the digestive system. It ranks as the sixth most commonly diagnosed cancer and the fourth leading cause of cancer-related deaths [176]. The risk factors for gastric cancer include HP, family history, poor diet, alcoholism, smoking, and Epstein-Barr virus infection [177]. The HP-induced chronic non-atrophic gastritis leads to intestinal epithelial hyperplasia, which increases the risk of gastric carcinogenesis [35]. Eradication of HP infection, particularly in first-degree relatives with a family history of gastric cancer, significantly reduces its incidence [178].

Multiple ILC subgroups, especially ILC2s, engage in crosstalk with HP, which diversely influences the progression of gastric cancer. Reduced abundance of NK cells has been observed in the tumor microenvironment of gastric cancer, which correlates with a worse prognosis [179]. Additionally, NK cell function is often compromised or even reversed in gastric cancer. Persistent IFN-y responses from NK and T cells post-HP infection may paradoxically enhance the risk of both gastric cancer and ulcers [180]. HP-derived lipopolysaccharide promotes the proliferation of low cytotoxic IL-10-secreting NK cells, thus impeding the elimination of HP and fostering chronic infection [181]. Other HP components, such as HopQ outer membrane adhesin and pro-inflammatory peptides, further impair NK cell functionality, adversely affecting gastric cancer prevention [182, 183]. Moreover, the impact of ILC2s on HP can vary with the disease's duration. ILC2s are a predominant ILC subset in the stomach, which are quickly induced by HP infection, peaking at two weeks. Furthermore, ILC activation by IL-33, originating from commensal microbes, produces IL-5, which stimulates IgA release. This ILC2-dependent IgA response is critical for eliminating pathogenic HP, thus safeguarding the stomach [111]. However, chronic HP infection suppresses IL-33 production, crucial for the effective activation of both ILC2s and T cells in the gastric mucosa and for inducing a Th2 response. Whereas exogenous IL-33 supplementation induces M2 polarization macrophage and transmural inflammation [22]. Lastly, Connexin43, essential for gastric mucosal gap junctions, shows a positive correlation with the progression from inflammation to gastric cancer. Its significance in HP-associated gastric cancer is linked to an upsurge in the transcription factor GATA-3, an ILC2s marker [184].

ILC is not only associated with HP, but also affects gastric cancer by interacting with inflammatory molecules, hormones, chemotaxis, and cell polarization. This multi-faceted interaction highlights the complexity of gastric cancer pathology. Gastric cancer cell-derived prostaglandin E2 inhibits NK cell proliferation and promotes apoptosis [185]. IFN-y induces high 927 gene expression in gastric cancer cells with enhanced invasion and metastasis and reduced susceptibility to NK cells [186]. Several cytokine therapies aiming to target and mitigate the adverse effects of NK cells have been explored. IL-2 helps NK cells restore Herceptin-mediated ADCC damage in gastric cancer patients [187]. Recombinant mouse IL-15 (rmIL-15) facilitates NK cell proliferation and produces more IFN-y to improve survival and inhibit liver metastasis in an inhibited gastric cancer mouse model [188]. NKG2D extracellular domain assembles with human IL-15 to form a fusion protein, dsNKG2D-IL-15, which helps NK cells infiltrate gastric cancer tissues and inhibit the growth of gastric cancer in mice [189]. Tumor cell-expressed CX3CL1 recruits NK cells and is associated with a better prognosis in gastric adenocarcinoma [190]. Cytokine therapy is significant for reversing NK cell dysfunction in gastric cancer, which may be a novel therapy for this cancer. It has been identified that ILC2s are positively associated with the development of gastric cancer. Spasmolytic polypeptide-expressing metaplasia (SPEM) is associated with the pathological progression of intestinal-type gastric cancer. This association highlights the potential therapeutic targets in SPEM-related pathways. ILC2 depletion inhibits post-injury chemotaxis by inhibiting the reprogramming of principal cells into SPEM, which begins after acute mural cell loss [25]. Endogenous intragastric glucocorticoids are necessary to inhibit the transcription of pro-inflammatory genes and the development of SPEM in the stomach [191]. Recent studies suggest that this inhibition may be synergistic with the action of androgens on ILC2s. The study confirms that androgens prevent gastric inflammation and chemotaxis by blocking the production of pro-inflammatory cytokines of ILC2s, which is supported by the prevention of SPEM development through the depletion of ILC2s [192]. Increased ILC2s in the periphery of gastric cancer patients and their resulting dysregulation of Th1/Th2 responses as well as increased MDSC and M2 phenotype macrophages constitute an immunosuppressive microenvironment in gastric cancer [140]. Compared to the healthy individuals, the levels of ILC3s and IL22 were observed to be significantly increased in gastritis, and precancerous lesion patients, suggesting that ILC3s may also be associated with gastritis and gastric cancer [193]. Polymorphisms in the IL-17 gene may also be associated with gastric cancer risk [194]. However, experimental studies on ILC3s and gastric cancer are still lacking.

Overall, ILC and gastric cancer crosstalk involve HP infection, chemotaxis, microenvironmental shaping of ILC, and cytokine networks. Future research focusing on these interactions could pave the way for novel therapeutic strategies. More adequate studies could help to stifle gastric cancer at the primary stage or inhibit stage progression.

## Chronic hepatitis and liver cancer

Although liver cancer does not have a significantly increased incidence, it has the lowest five-year survival rate of all cancers and it ranks fifth in cancer-related deaths among men [1]. HCC is more closely related to chronic inflammation than other cancers, and most HCCs are associated with chronic liver inflammation and its mediated cirrhosis [195-197]. This strong link to inflammation highlights the critical need for targeted therapeutic strategies. Hepatic fibrosis as a pathological state is common in various viral hepatitis, non-alcoholic steatohepatitis (NASH), and alcoholic liver disease. The effectiveness of vaccines and antiviral treatment for some hepatitis is significant, but not sufficient. This gap necessitates the development of additional treatment options. Since liver fibrosis is generally considered irreversible, the main goal is to inhibit its progression. Immunotherapy for whole-body treatment has been recognized and promoted [198]. As an emerging immune cell, the role of ILC in HCC has also been widely investigated. This section will focus on the role of ILC in liver inflammation, fibrosis, and interaction with the tumor microenvironment.

NK cells, which constitute approximately half of the lymphocytes in the liver [199], are thought to have the ability to inhibit hepatic fibrosis. HSC activation promotes the proliferation of fibrotic myofibroblasts is the initiation as well as the key process of fibrosis. In the early stage of liver fibrosis, activated HSC is recognized by NK cells, which subsequently mediate apoptosis through the TRAIL/FasL pathway, granule cytotoxicity, and the production of IFN-y leading to HSC death [39, 109, 110]. This demonstrates the potential therapeutic value of NK cells in early-stage fibrosis. Moreover, as fibrosis progressed to a later stage, HSC inversely inhibited the antifibrotic capacity of NK cells in a TGF- $\beta$ -dependent form [23]. NK-derived IFN-y facilitates macrophage polarization to the M1 phenotype and prevents the progression of NASH to fibrosis [138]. A recent study showed that CD8+ tissue-resident memory T cells induced hepatic stellate cell apoptosis to regress liver fibrosis in NASH [200]. In addition, group 1 ILC inhibit the proliferation of CD8<sup>+</sup> T cells by affecting the availability of IL-2 in long-term interaction with CD8+ T cells, which may be another target for inhibiting liver fibrosis [201]. In the early HCC stage, inflammatory cytokines (such as TNF- $\alpha$  and IFN- $\gamma$ ) induce CD56<sup>+</sup> NK cells to infiltrate and kill cancer cells by stimulating tumors chemokines production (such as CXCL10, CCL5, and CCL2) [202]. The role of NK cells in HCC highlights their importance in both anti-fibrotic and anti-cancer responses. Specific localization of NK cells in HCC progression correlates with the respective phenotype

of each subtype. The immune microenvironment of HCC is inextricably linked to the phenotypic shaping of liver-specific ILC. In NK cells IL-10 may excessively express NKG2A, which is positively correlated with NK cell depletion and dysfunction [203]. In vitro studies have indicated that high TGF- $\beta$  concentrations mediate the differentiation of NK-like cells to ILC1s as well as the loss of its cytotoxicity, whereas cytokine gradient mediates tumor ILC composition [204]. Some immune cells can also affect NK cells. The activation receptor for NK, NKp30, reduces significantly after co-culture with MDSC, which inhibits both NK cytotoxicity and IFN-y release [205]. In addition, patients on immune checkpoint inhibitors indicated increased KLRF1<sup>high</sup> NK-like ILC subgroup, which was associated with progression- and relapse-free survival, and warrants further research to assess the exact mechanism [206]. CD49a+Eomes+ NK cells, typical dNK cells, can promote HCC development by promoting angiogenesis and reducing cytotoxicity, as also observed in CRC [207].

ILC2s promote chronic hepatitis, which may be associated with two cytokines, IL-33 and IL-13. Hepatocyte damage releases large amounts of IL-33 to promote ILC2s activation and expansion, while IL-13 secreted by ILC2s promotes hepatic fibrosis in a STAT6-dependent manner or by activating HSC [208]. This indicates the critical role of ILC2s in the progression of liver diseases. The enrichment of ILC2s in human HCC has been associated with a higher recurrence and shorter survival rates. The reduction of e-cadherin in tumor cells leads to loss of killer cell lectin-like receptor G1 (KLRG1) expression in ILC2s, and this KLRG1- ILC2s subgroup secretes more CXCL2 recruitment neutrophils to form an immunosuppressive microenvironment [121]. This suggests that targeting ILC2s could be a strategic approach to combat HCC. Furthermore, B cells participate in the ILC2s reprogramming, and the derived ICOSL signal induces IL-13 secretion via ICOS+ ILC2s, which promotes the progression of hepatitis and liver cancer [209]. Recently, ILC3s have also been implicated in liver fibrosis as they enhance liver fibrosis by producing IL-22 and IL-17A when co-cultured with human HSC in vitro, and their secretion of IL-22 can block the anti-fibrotic effect of IFN-γ [13]. In addition, high levels of IL-23 can help NCR- ILC3s to secrete IL-17, suggesting poor clinical prognosis of HCC and weakened CD8+ T cell immunity [60].

Overall, ILC and cytokines should be focused in the early stage of the disease to shape its characteristic phenotype. This approach could be a more effective intervention for managing and treating liver diseases.



**Figure 3: ILC in the Digestive System Tumor Microenvironment.** During inflammation, different subsets of ILC interact with various factors including cytokines, immune cells, and tumor cells, significantly impacting the progression of digestive system tumors. This interaction highlights ILC's close relation to key targets critical for the transformation from digestive system inflammation to cancer, such as HSC and HP. ILC modulate the development and outcome of digestive system cancer through multiple pathways. However, it's important to note that the phenotype and function of ILC are also reciprocally shaped by the immune microenvironment. NK cells can inhibit the progression of chronic pancreatitis and PDAC by secreting IFN-y to induce CSC differentiation, blocking fibrosis, promoting necrosis, and limiting proliferation in pancreatic cancer cells. Furthermore, they may also exacerbate gastritis and gastric carcinogenesis due to their weak cytotoxicity and mediation of a sustained IFN-y response. This process is enhanced by HP and its secreted products, but targeted cytokine therapies may help NK cells regain their inhibitory capacity against gastric cancer. Moreover, NK cells promote hepatitis and fibrosis associated with HCC by inhibiting HSC. The conversion of NK cells to dNK mediates the progression of HCC and CRC. ILC1s may promote hepatic fibrosis by decreasing the killing of HSC by T cells. ILC2s inhibit the progression of gastrit to gastric cancer by killing HP while also mediating gastric cancer progression through SPEM and interaction with immunosuppressive cells. AREG secreted by ILC2s induces Tregs-mediated promotion of CRC. Transitioning from ILC2s to ILCreg promotes the progression of PDAC. ILC3s mediate colorectal inflammation by secreting IL-17 and IL-22 and contribute to cancer development. Their transition to ILCreg may also

## Discussion

# ILC subpopulations are significant potential targets for cancer immunotherapy

ILC have an innate ability to communicate with cytokines and surrounding cells. This not only them with external associated stimuli and homeostasis but also with inflammation and tumor inevitably, given the complex and interconnected inflammation and tumor microenvironment. This dynamic interaction is crucial for understanding the role of ILC in health and disease. However, this communication ability is a two-way street. ILC not only kill pathogens, recruit and polarize immune cells, influence epithelial metaplasia, and even directly act on tumor stem cells, but are also shaped as they shape the immune microenvironment they house. This reciprocal influence is key to their functionality. For example, after the influence of IL-15

and TGF- $\beta$ , NK transforms into ieILC1s, which inhibits cancer, and ILC1s (which has weak cytotoxicity) or anoxic environment mediates ILC2s into ILCreg phenotype, which promotes pancreatic cancer. Therefore, strategic modulation of ILC phenotypes could be a significant focus in cancer research.

#### Key players in the progression of inflammation and cancer in the digestive System

Inflammation should be exclusionary and reparative; however, chronic inflammatory responses are detrimental to tumor control. Chronic inflammation can influence tumor development, distant metastasis, and resistance to therapy at the genetic level, through cellular interactions, and especially through the construction of the immune microenvironment. The spread of chronic inflammation to cancer is thought to be a relatively

reversible and progressive process. The digestive system is a mucosal immune system that has extensive contact with the outside environment and comprises а large and complex immune microenvironment. Positive correlations have been demonstrated between HP-associated gastritis, NASH, chronic pancreatitis, intestinal inflammation, and cancers of various organs. These findings indicate intervention is crucial in that early these prevent inflammatory processes to cancer development. How to reverse or inhibit the progression of this process is crucial for early detection and treatment of cancer. Cytokine therapies involving ILC have provided additional options for immunotherapy in pancreatic and gastric cancers, such as dsNKG2D-IL-15, rIL-15, and L19-IL-2; however, ILC have higher efficiency and function. Further exploration into the diverse roles of ILC in cancer could identify new avenues for treatment.

ILC has been shown to pervade the digestive system and is associated with several key nodes from inflammation to cancer. First, ILC can inhibit the onset and progression of inflammation in the digestive system. This emphasizes the preventative role of ILC in disease progression. NK cells, synergistically with IL-15, can inhibit fibrosis in chronic pancreatitis by suppressing alveolar cell atrophy, preventing tissue collagen accumulation around blood vessels, and down-regulate pro-fibrotic genes. Furthermore, it has been observed that ILC2s produce IL-5 to mediate the IgA response in the early stages of HP infection, thereby eliminating pathogenic HP and thus inhibiting the development of gastritis. Moreover, androgens have been observed to prevent gastric inflammation and chemotaxis by blocking the production of proinflammatory cytokines of ILC2s. NK cells inhibit hepatitis progression and fibrosis by mediating apoptosis through the TRAIL/FasL pathway, granule cytotoxicity, and the production of IFN-y, leading to HSC death. This highlights the therapeutic potential of leveraging ILC in inflammation control. However, as mentioned, the inflammatory environment also shapes or inhibits ILC function, and some ILC have opposite roles. Chronic HP infection inhibits NK cell toxicity, induces a long-term NK cell-associated IFN-y response, and increases the risk of gastric cancer or inhibits IL-33, thereby limiting the activation of ILC2s that can clear it. When the fibrosis of the liver progressed to a later stage, HSC could inversely inhibit the anti-fibrotic capacity of NK cells via the TGF-β-dependent form. The specific role of different ILC in various stages of inflammation at key targets is crucial to prevent the progression of chronic digestive inflammation to tumors.

In addition to interacting with inflammation, ILC are involved in other pathways throughout digestive cancers. ILC can directly affect cancer cell proliferation and survival through their cytotoxicity, ADCC effects, and promotion of CSC differentiation. In addition, ILC can also affect signaling pathways such as IL-33/ST2, AREG-EGFR, AKT, STAT-3, *etc.* Moreover, ILC recruits, polarizes and functionally affects CD8<sup>+</sup> T cells, Tregs, MDSC, neutrophils, and macrophages, which helps to shape the tumor immune microenvironment. ILC are also involved in angiogenesis and tumor migration. Almost all of these processes are associated with cytokines secreted by ILC.

#### **Current limitations and future directions**

Although ILC have demonstrated considerable clinical potential, the majority of currently available information on ILC is derived from in vitro experiments and preclinical studies. Targeting ILC may have associated side effects and limitations; however, whether the results of preclinical studies are translated clinically and how these therapies can be personalized according to the tumor microenvironment or patient-specific factors should be studied. The practical translation of the above potential functions of ILC into clinically applicable immunotherapeutic regimens can be performed in the following ways. The first is to identify how to enhance the specific features of ILC per requirement. Cvtokines, chemokines, microorganisms, and hormones offer many possibilities, which can be divided among different ILC groups. Another question is how to restore the suppressed function of ILC. This may require a more in-depth study of the specific mechanisms by which ILC interacts with surrounding immune cells, stromal cells, and microorganisms. Understanding these interactions is crucial for therapeutic development. Furthermore, when clarifying the pros and cons of ILC, it should be remembered that it performs different functions at different time points and can have ambiguous double-edged effects at the same time point. This complexity necessitates careful consideration in immunotherapy research. Therefore, the possible adverse effects of ILC should be considered when immunotherapy studies are conducted. This could, however, be solved by considering the plasticity of ILC. The role of ILC is inextricably linked to phenotype; therefore, more comprehensive studies of the typing and role of tissue-resident and peripheral blood ILC are needed. Further research in this area could lead to breakthroughs in targeted therapy. Finally, and most importantly, how ILC-related immunotherapies can be targeted should be assessed.

#### Table 1: Effect of ILC on cancer.

Cancer	ILC subsets	Effects	Possible mechanism	Contributing factors	Inhibiting factors	References
Pancreatic cancer	NK	Suppression	NK cells induce CSC differentiation and inhibit pancreatic tumor growth and metastasis by secreting IFN-γ and TNF-α, ADCC, recruiting T cells, extensive necrosis and limited proliferation, and prevention of fibrosis.	IL-21, IL-15, IL-2		[36, 120, 148, 149]
Pancreatic cancer	ILC2s	Suppression	DC recruitment, CD8+T cell recruitment and activation, PD-1 pathway blocking enhancement	IL-33		[74]
Pancreatic cancer	ILC2s	Promotion	Transformation into ILCreg	Hypoxia, IL-33	AG	[34, 73]
Pancreatic cancer	ILC3s	Promotion	Production of IL-22 mediates AKT signaling.			[153]
CRC	NK	Promotion	Reduction of cytotoxicity, transformation to dNK, and promotion of angiogenesis	Tumor cells		[164]
CRC	ILC1s	Promotion	ILC1s drives intestinal epithelial and matrix remodeling.			[165]
CRC	ILC2s	Promotion	Production of AREG and enhancement of Tregs function, production of IL-13-mediated STAT6 activation, maintenance of tumor invasion MDSC	IL-25, IL-33	NMU	[131, 167-169]
CRC	ILC3s	Promotion	Production of IL-17, IL-22, and IFN-γ, transition to ILCreg, decreased communication with T cells	TGF-β		[173-175]
Gastric cancer	NK	Promotion	Persistent IFN- $\gamma$ response, low cytotoxicity, and production of IFN- $\gamma$ can contribute to gastric cancer cell invasive and metastatic capacity and reduce susceptibility to NK cells.	HP and its products, prostaglandin E2	IL-2,IL-15,CX3C L1	[180, 182, 183, 185-190]
Gastric cancer	ILC2s	Promotion	SPEM, dysregulation of Th1/Th2 responses together with increased MDSC and M2 phenotype macrophages.		Endogenous intragastric glucocorticoids, androgens	[25, 140, 192]
Gastric cancer	ILC2s	Suppression	ILC2s produce IL-5 and mediate IgA response to eradicate HP.	IL-33	HP	[22, 111]
HCC	NK	Suppression	NK cells promote HSC cell death and IFN- $\gamma$ production and help macrophages polarize towards the M1 phenotype.	IFN-γ, TNF-α	HSC, IL-10, TGF-β, MDSC	[23, 39, 109, 110, 138, 202, 203, 205]
HCC	NK	Promotion	Transformation to dNK			[207]
HCC	ILC2s	Promotion	Secretion of IL-13, CXCL2 and recruitment of neutrophils	B cell		[121, 208, 209]
HCC	ILC3s	Promotion	Production of IL-22 and IL-17A, prevention of IFN- $\gamma$ from antifibrosis, proliferation reduction, and increased apoptosis of CD8+T cells.			[13, 60]

In addition to engineered cytokines, the genetic engineering of ILC is also worth exploring, which can be achieved *via* CAR-engineered NK (CAR-NK) cells. While optimizing manufacturing processes, scientists should also consider how to enhance CAR-NK cells' infiltration into solid tumors, cytotoxicity, and prolonged and reliable tumor surveillance. Moreover, CAR-NK cells have a better safety profile compared to engineered T cells. In the autologous setting, there is a reduced likelihood of cytokine release syndrome and neurotoxicity, while in the allogeneic setting, there is a reduced risk of graft-versus-host disease.

## Conclusion

In summary, ILC are valuable for tumor control as they influence inflammation in the digestive system and shape the immune microenvironment. Furthermore, they predominantly function by interacting with other cells and cytokines. Moreover, their high degree of plasticity and heterogeneity offers possibilities for the directional shaping of their relevant functions by influencing the environment in which they live. The construction of a comprehensive and detailed cytokine-ILC-immune microenvironment network is necessary for further understanding digestive disorders and to provide additional immunotherapeutic possibilities.

## Abbreviations

ILC: Innate Lymphoid Cells; CRC: Colorectal Cancer; Hp: Helicobacter pylori; STAT-3: Signal Transducer and Activator of Transcription 3; IL-2: Interleukin-2; TNF: Tumor Necrosis Factor; TCR: T cell receptor; NK: Natural killer; ILC1s: Type 1 innate lymphoid cells: IFN: Interferon; AREG: Amphiregulin; NF- $\kappa$ B: Nuclear factor  $\kappa$ B (NF- $\kappa$ B); Th1: T helper type 1; TRAIL: Tumour necrosis factor-related apoptosis-inducing ligand; trNK: Tissue-resident NK; dNK: Decidual NK; ieILC1s: Intraepithelial ILC1s; TGF-β: Transforming growth factor-*β*; PDAC: Pancreatic ductal adenocarcinoma; PD-1: Programmed cell death protein-1; DC: Dendritic cells; MDSC: Myeloid-derived suppressor cells; KLRG1: Killer cell lectin-like receptor G1; CXCL2: C-X-C chemokine ligand 2; LTi: Lymphoid tissue inducer; AHR: Aryl hydrocarbon receptor; SCFA: Short-chain fatty acids; IBD: Inflammatory bowel disease; TLS: Tumor-associated tertiary lymphoid structures; NKT: Natural killer T; CCL21: Chemokine ligand 21; NASH: Non-alcoholic steatohepatitis; HCC: Hepatocellular carcinoma; HSC: Hepatic stellate cell; CSC: Cancer stem cell; ADCC: Antibody-dependent cell-mediated cytotoxicity; L19-IL2: L19-Interleukin-2; rIL-15: Recombinant IL-15; ILCreg: Regulatory innate lymphoid cells; AG: AG (nab-paclitaxel + gemcitabine); NMU: Neurohormone U; rmIL-15: Recombinant mouse IL-15; SPEM: Spasmolytic polypeptide-expressing metaplasia; CAR-NK: CAR-engineered NK; Tregs: regulatory T cells.

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#### **Author contributions**

GS and QW prepared the figure and drafted this manuscript. GS, QW, ZL, JX, XH, ZW, PZ, and SZ conceived the idea and analyzed the literature. MX, XW, and XH edited and revised the manuscript. All authors contributed to the article and approved the submitted version. All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

#### **Competing Interests**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

#### References

- Siegel RL, Miller KD, Wagle NS, Jemal A. Cancer statistics, 2023. CA Cancer J Clin. 2023; 73: 17-48.
- Tian H, Zhang T, Qin S, Huang Z, Zhou L, Shi J, et al. Enhancing the therapeutic efficacy of nanoparticles for cancer treatment using versatile targeted strategies. J Hematol Oncol. 2022; 15: 132.
- Liu Z, Shi M, Ren Y, Xu H, Weng S, Ning W, et al. Recent advances and applications of CRISPR-Cas9 in cancer immunotherapy. Mol Cancer. 2023; 22: 35.
- Liu P, Zhu H, Zhu H, Zhang X, Feng A, Zhu X, et al. Predicting Survival for Hepatic Arterial Infusion Chemotherapy of Unresectable Colorectal Liver Metastases: Radiomics Analysis of Pretreatment Computed Tomography. J Transl Int Med. 2022; 10: 56-64.
- Jiang X, Zou W, Liao Z. Chemical Therapy for Chronic Pancreatitis: An Assumption or an Alternative? J Transl Int Med. 2022; 10: 304-7.
- Sun K, Lv H, Chen B, Nie C, Zhao J, Wang S, et al. Dawning precision treatment for gastric cancer: The latest biomarkers. J Transl Int Med. 2021; 9: 228-30.
- Xie J, Luo X, Deng X, Tang Y, Tian W, Cheng H, et al. Advances in artificial intelligence to predict cancer immunotherapy efficacy. Front Immunol. 2022; 13: 1076883.
- Gauci ML, Lanoy E, Champiat S, Caramella C, Ammari S, Aspeslagh S, et al. Long-Term Survival in Patients Responding to Anti-PD-1/PD-L1 Therapy and

Disease Outcome upon Treatment Discontinuation. Clin Cancer Res. 2019; 25: 946-56.

- Grossmann KF, Margolin K. Long-term survival as a treatment benchmark in melanoma: latest results and clinical implications. Ther Adv Med Oncol. 2015; 7: 181-91.
- 10. Taniguchi K, Karin M. IL-6 and related cytokines as the critical lynchpins between inflammation and cancer. Semin Immunol. 2014; 26: 54-74.
- Mills KHG. IL-17 and IL-17-producing cells in protection versus pathology. Nat Rev Immunol. 2023; 23: 38-54.
- Grivennikov SI, Wang K, Mucida D, Stewart CA, Schnabl B, Jauch D, et al. Adenoma-linked barrier defects and microbial products drive IL-23/IL-17-mediated tumour growth. Nature. 2012; 491: 254-8.
- Wang S, Li J, Wu S, Cheng L, Shen Y, Ma W, et al. Type 3 innate lymphoid cell: a new player in liver fibrosis progression. Clin Sci (Lond). 2018; 132: 2565-82.
- Puca E, Schmitt-Koopmann C, Furter M, Murer P, Probst P, Dihr M, et al. The targeted delivery of interleukin-12 to the carcinoembryonic antigen increases the intratumoral density of NK and CD8(+) T cell in an immunocompetent mouse model of colorectal cancer. J Gastrointest Oncol. 2020; 11: 803-11.
- Millul J, Krudewig C, Zana A, Dakhel Plaza S, Puca E, Villa A, et al. Immunotherapy with Immunocytokines and PD-1 Blockade Enhances the Anticancer Activity of Small Molecule-Drug Conjugates Targeting Carbonic Anhydrase IX. Mol Cancer Ther. 2021; 20: 512-22.
- Mehralizadeh H, Nazari A, Oruji F, Roostaie M, Hosseininozari G, Yazdani O, et al. Cytokine sustained delivery for cancer therapy; special focus on stem cell- and biomaterial- based delivery methods. Pathol Res Pract. 2023; 247: 154528.
- Klose CSN, Flach M, Möhle L, Rogell L, Hoyler T, Ebert K, et al. Differentiation of type 1 ILCs from a common progenitor to all helper-like innate lymphoid cell lineages. Cell. 2014; 157: 340-56.
- Huang C, Bi J. Expression Regulation and Function of T-Bet in NK Cells. Front Immunol. 2021; 12: 761920.
- Tindemans I, Serafini N, Di Santo JP, Hendriks RW. GATA-3 function in innate and adaptive immunity. Immunity. 2014; 41: 191-206.
- Uddin MJ, Leslie JL, Burgess SL, Oakland N, Thompson B, Abhyankar M, et al. The IL-33-ILC2 pathway protects from amebic colitis. Mucosal Immunol. 2022; 15: 165-75.
- Schroeder JH, Howard JK, Lord GM. Transcription factor-driven regulation of ILC1 and ILC3. Trends Immunol. 2022; 43: 564-79.
- Buzzelli JN, Chalinor HV, Pavlic DI, Sutton P, Menheniott TR, Giraud AS, et al. IL33 Is a Stomach Alarmin That Initiates a Skewed Th2 Response to Injury and Infection. Cell Mol Gastroenterol Hepatol. 2015; 1: 203-21.e3.
- Shi J, Zhao J, Zhang X, Cheng Y, Hu J, Li Y, et al. Activated hepatic stellate cells impair NK cell anti-fibrosis capacity through a TGF-β-dependent emperipolesis in HBV cirrhotic patients. Sci Rep. 2017; 7: 44544.
- Schroeder JH, Roberts LB, Meissl K, Lo JW, Hromadová D, Hayes K, et al. Sustained Post-Developmental T-Bet Expression Is Critical for the Maintenance of Type One Innate Lymphoid Cells *In Vivo*. Front Immunol. 2021; 12: 760198.
- Meyer AR, Engevik AC, Madorsky T, Belmont E, Stier MT, Norlander AE, et al. Group 2 Innate Lymphoid Cells Coordinate Damage Response in the Stomach. Gastroenterology. 2020; 159: 2077-91.e8.
- Michels N, van Aart C, Morisse J, Mullee A, Huybrechts I. Chronic inflammation towards cancer incidence: A systematic review and meta-analysis of epidemiological studies. Crit Rev Oncol Hematol. 2021; 157: 103177.
- Shui Y, Li M, Su J, Chen M, Gu X, Guo W. Prognostic and clinicopathological significance of systemic immune-inflammation index in pancreatic cancer: a meta-analysis of 2,365 patients. Aging (Albany NY). 2021; 13: 20585-97.
- Yuan K, Ye J, Liu Z, Ren Y, He W, Xu J, et al. Complement C3 overexpression activates JAK2/STAT3 pathway and correlates with gastric cancer progression. J Exp Clin Cancer Res. 2020; 39: 9.
- Stockdale AJ, Kreuels B, Henrion MYR, Giorgi E, Kyomuhangi I, de Martel C, et al. The global prevalence of hepatitis D virus infection: Systematic review and meta-analysis. J Hepatol. 2020; 73: 523-32.
- Xie H, Ruan G, Ge Y, Zhang Q, Zhang H, Lin S, et al. Inflammatory burden as a prognostic biomarker for cancer. Clin Nutr. 2022; 41: 1236-43.
- Yao D, Dong M, Dai C, Wu S. Inflammation and Inflammatory Cytokine Contribute to the Initiation and Development of Ulcerative Colitis and Its Associated Cancer. Inflamm Bowel Dis. 2019; 25: 1595-602.
- Erdman SE, Poutahidis T. Cancer inflammation and regulatory T cells. Int J Cancer. 2010; 127: 768-79.
- Wherry EJ, Blattman JN, Murali-Krishna K, van der Most R, Ahmed R. Viral persistence alters CD8 T-cell immunodominance and tissue distribution and results in distinct stages of functional impairment. J Virol. 2003; 77: 4911-27.
- Ye L, Jin K, Liao Z, Xiao Z, Xu H, Lin X, et al. Hypoxia-reprogrammed regulatory group 2 innate lymphoid cells promote immunosuppression in pancreatic cancer. EBioMedicine. 2022; 79: 104016.
- Ernst PB, Gold BD. The disease spectrum of Helicobacter pylori: the immunopathogenesis of gastroduodenal ulcer and gastric cancer. Annu Rev Microbiol. 2000; 54: 615-40.
- Manohar M, Kandikattu HK, Verma AK, Mishra A. IL-15 regulates fibrosis and inflammation in a mouse model of chronic pancreatitis. Am J Physiol Gastrointest Liver Physiol. 2018; 315: G954-g65.
- Raabe J, Kaiser KM, ToVinh M, Finnemann C, Lutz P, Hoffmeister C, et al. Identification and characterization of a hepatic IL-13-producing ILC3-like

population potentially involved in liver fibrosis. Hepatology. 2023; 78: 787-802.

- Yang YM, Kim SY, Seki E. Inflammation and Liver Cancer: Molecular Mechanisms and Therapeutic Targets. Semin Liver Dis. 2019; 39: 26-42.
- Jeong WI, Park O, Gao B. Abrogation of the antifibrotic effects of natural killer cells/interferon-gamma contributes to alcohol acceleration of liver fibrosis. Gastroenterology. 2008; 134: 248-58.
- Wong P, Foltz JA, Chang L, Neal CC, Yao T, Cubitt CC, et al. T-BET and EOMES sustain mature human NK cell identity and antitumor function. J Clin Invest. 2023; 133: e162530.
- Nixon BG, Chou C, Krishna C, Dadi S, Michel AO, Cornish AE, et al. Cytotoxic granzyme C-expressing ILC1s contribute to antitumor immunity and neonatal autoimmunity. Science Immunology. 2022; 7: eabi8642.
- Gordon SM, Chaix J, Rupp LJ, Wu J, Madera S, Sun JC, et al. The transcription factors T-bet and Eomes control key checkpoints of natural killer cell maturation. Immunity. 2012; 36: 55-67.
- Krämer B, Nalin AP, Ma F, Eickhoff S, Lutz P, Leonardelli S, et al. Single-cell RNA sequencing identifies a population of human liver-type ILC1s. Cell Rep. 2023; 42: 111937.
- Corvino D, Kumar A, Bald T. Plasticity of NK cells in Cancer. Frontiers in Immunology. 2022; 13: 888313.
- Weizman OE, Adams NM, Schuster IS, Krishna C, Pritykin Y, Lau C, et al. ILC1 Confer Early Host Protection at Initial Sites of Viral Infection. Cell. 2017; 171: 795-808.e12.
- Nabekura T, Shibuya A. ILC1: guardians of the oral mucosa against enemy viruses. Immunity. 2021; 54: 196-8.
- Bernink JH, Peters CP, Munneke M, te Velde AA, Meijer SL, Weijer K, et al. Human type 1 innate lymphoid cells accumulate in inflamed mucosal tissues. Nat Immunol. 2013; 14: 221-9.
- Nazmi A, Hoek KL, Greer MJ, Piazuelo MB, Minato N, Olivares-Villagómez D. Innate CD8αα+ cells promote ILC1-like intraepithelial lymphocyte homeostasis and intestinal inflammation. PLoS One. 2019; 14: e0215883.
- de Jonge K, Ebering A, Nassiri S, Maby-El Hajjami H, Ouertatani-Sakouhi H, Baumgaertner P, et al. Circulating CD56(bright) NK cells inversely correlate with survival of melanoma patients. Sci Rep. 2019; 9: 4487.
- Nersesian S, Schwartz SL, Grantham SR, MacLean LK, Lee SN, Pugh-Toole M, et al. NK cell infiltration is associated with improved overall survival in solid cancers: A systematic review and meta-analysis. Transl Oncol. 2021; 14: 100930.
- Cooper MA, Fehniger TA, Turner SC, Chen KS, Ghaheri BA, Ghayur T, et al. Human natural killer cells: a unique innate immunoregulatory role for the CD56(bright) subset. Blood. 2001; 97: 3146-51.
- Hanna J, Goldman-Wohl D, Hamani Y, Avraham I, Greenfield C, Natanson-Yaron S, et al. Decidual NK cells regulate key developmental processes at the human fetal-maternal interface. Nat Med. 2006; 12: 1065-74.
- 53. Fuchs A, Vermi W, Lee JS, Lonardi S, Gilfillan S, Newberry RD, et al. Intraepithelial type 1 innate lymphoid cells are a unique subset of IL-12- and IL-15-responsive IFN- $\gamma$ -producing cells. Immunity. 2013; 38: 769-81.
- Dadi S, Chhangawala S, Whitlock BM, Franklin RA, Luo CT, Oh SA, et al. Cancer Immunosurveillance by Tissue-Resident Innate Lymphoid Cells and Innate-like T Cells. Cell. 2016; 164: 365-77.
- Moreno-Nieves UY, Tay JK, Saumyaa S, Horowitz NB, Shin JH, Mohammad IA, et al. Landscape of innate lymphoid cells in human head and neck cancer reveals divergent NK cell states in the tumor microenvironment. Proc Natl Acad Sci U S A. 2021; 118: e2101169118.
- 56. Cortez VS, Ulland TK, Cervantes-Barragan L, Bando JK, Robinette ML, Wang Q, et al. SMAD4 impedes the conversion of NK cells into ILC1-like cells by curtailing non-canonical TGF- $\beta$  signaling. Nat Immunol. 2017; 18: 995-1003.
- Gao Y, Souza-Fonseca-Guimaraes F, Bald T, Ng SS, Young A, Ngiow SF, et al. Tumor immunoevasion by the conversion of effector NK cells into type 1 innate lymphoid cells. Nat Immunol. 2017; 18: 1004-15.
- Hawke LG, Mitchell BZ, Ormiston ML. TGF-β and IL-15 Synergize through MAPK Pathways to Drive the Conversion of Human NK Cells to an Innate Lymphoid Cell 1-like Phenotype. J Immunol. 2020; 204: 3171-81.
- Koh J, Kim HY, Lee Y, Park IK, Kang CH, Kim YT, et al. IL23-Producing Human Lung Cancer Cells Promote Tumor Growth via Conversion of Innate Lymphoid Cell 1 (ILC1) into ILC3. Clin Cancer Res. 2019; 25: 4026-37.
- Liu Y, Song Y, Lin D, Lei L, Mei Y, Jin Z, et al. NCR(-) group 3 innate lymphoid cells orchestrate IL-23/IL-17 axis to promote hepatocellular carcinoma development. EBioMedicine. 2019; 41: 333-44.
- 61. Klein Wolterink RG, Serafini N, van Nimwegen M, Vosshenrich CA, de Bruijn MJ, Fonseca Pereira D, et al. Essential, dose-dependent role for the transcription factor Gata3 in the development of IL-5+ and IL-13+ type 2 innate lymphoid cells. Proc Natl Acad Sci U S A. 2013; 110: 10240-5.
- Mjösberg JM, Trifari S, Crellin NK, Peters CP, van Drunen CM, Piet B, et al. Human IL-25- and IL-33-responsive type 2 innate lymphoid cells are defined by expression of CRTH2 and CD161. Nat Immunol. 2011; 12: 1055-62.
- Bartemes KR, Kephart GM, Fox SJ, Kita H. Enhanced innate type 2 immune response in peripheral blood from patients with asthma. J Allergy Clin Immunol. 2014; 134: 671-8.e4.
- Varela F, Symowski C, Pollock J, Wirtz S, Voehringer D. IL-4/IL-13-producing ILC2s are required for timely control of intestinal helminth infection in mice. Eur J Immunol. 2022; 52: 1925-33.

- Tamadaho RSE, Osei-Mensah J, Arndts K, Debrah LB, Debrah AY, Layland LE, et al. Reduced Type 2 Innate Lymphocyte Cell Frequencies in Patent Wuchereria bancrofti-Infected Individuals. Pathogens. 2023; 12: 665.
- Halim TY, Krauss RH, Sun AC, Takei F. Lung natural helper cells are a critical source of Th2 cell-type cytokines in protease allergen-induced airway inflammation. Immunity. 2012; 36: 451-63.
- Hsu AT, Gottschalk TA, Tsantikos E, Hibbs ML. The Role of Innate Lymphoid Cells in Chronic Respiratory Diseases. Front Immunol. 2021; 12: 733324.
- Roediger B, Kyle R, Tay SS, Mitchell AJ, Bolton HA, Guy TV, et al. IL-2 is a critical regulator of group 2 innate lymphoid cell function during pulmonary inflammation. J Allergy Clin Immunol. 2015; 136: 1653-63.e7.
- Bal SM, Bernink JH, Nagasawa M, Groot J, Shikhagaie MM, Golebski K, et al. IL-1β, IL-4 and IL-12 control the fate of group 2 innate lymphoid cells in human airway inflammation in the lungs. Nat Immunol. 2016; 17: 636-45.
- Moro K, Kabata H, Tanabe M, Koga S, Takeno N, Mochizuki M, et al. Interferon and IL-27 antagonize the function of group 2 innate lymphoid cells and type 2 innate immune responses. Nat Immunol. 2016; 17: 76-86.
- Maric J, Ravindran A, Mazzurana L, Björklund Å K, Van Acker A, Rao A, et al. Prostaglandin E(2) suppresses human group 2 innate lymphoid cell function. J Allergy Clin Immunol. 2018; 141: 1761-73.e6.
- Li Y, Lin S, Xiong S, Xie Q. Recombinant Expression of Human IL-33 Protein and Its Effect on Skin Wound Healing in Diabetic Mice. Bioengineering (Basel). 2022; 9: 734.
- Alam A, Levanduski E, Denz P, Villavicencio HS, Bhatta M, Alhorebi L, et al. Fungal mycobiome drives IL-33 secretion and type 2 immunity in pancreatic cancer. Cancer Cell. 2022; 40: 153-67.e11.
- Moral JA, Leung J, Rojas LA, Ruan J, Zhao J, Sethna Z, et al. ILC2s amplify PD-1 blockade by activating tissue-specific cancer immunity. Nature. 2020; 579: 130-5.
- Guillerey C, Stannard K, Chen J, Krumeich S, Miles K, Nakamura K, et al. Systemic administration of IL-33 induces a population of circulating KLRG1(hi) type 2 innate lymphoid cells and inhibits type 1 innate immunity against multiple myeloma. Immunol Cell Biol. 2021; 99: 65-83.
- Maywald RL, Doerner SK, Pastorelli L, De Salvo C, Benton SM, Dawson EP, et al. IL-33 activates tumor stroma to promote intestinal polyposis. Proc Natl Acad Sci U S A. 2015; 112: E2487-96.
- Wang S, Zhang Y, Wang Y, Ye P, Li J, Li H, et al. Amphiregulin Confers Regulatory T Cell Suppressive Function and Tumor Invasion via the EGFR/GSK-3β/Foxp3 Axis. J Biol Chem. 2016; 291: 21085-95.
- Ito A, Akama Y, Satoh-Takayama N, Saito K, Kato T, Kawamoto E, et al. Possible Metastatic Stage-Dependent ILC2 Activation Induces Differential Functions of MDSCs through IL-13/IL-13Rα1 Signaling during the Progression of Breast Cancer Lung Metastasis. Cancers (Basel). 2022; 14: 3267.
- Long A, Dominguez D, Qin L, Chen S, Fan J, Zhang M, et al. Type 2 Innate Lymphoid Cells Impede IL-33-Mediated Tumor Suppression. J Immunol. 2018; 201: 3456-64.
- Spits H, Artis D, Colonna M, Diefenbach A, Di Santo JP, Eberl G, et al. Innate lymphoid cells--a proposal for uniform nomenclature. Nat Rev Immunol. 2013; 13: 145-9.
- Fiancette R, Finlay CM, Willis C, Bevington SL, Soley J, Ng STH, et al. Reciprocal transcription factor networks govern tissue-resident ILC3 subset function and identity. Nat Immunol. 2021; 22: 1245-55.
- Lee JS, Cella M, McDonald KG, Garlanda C, Kennedy GD, Nukaya M, et al. AHR drives the development of gut ILC22 cells and postnatal lymphoid tissues via pathways dependent on and independent of Notch. Nat Immunol. 2011; 13: 144-51.
- Gao X, Shen X, Liu K, Lu C, Fan Y, Xu Q, et al. The Transcription Factor ThPOK Regulates ILC3 Lineage Homeostasis and Function During Intestinal Infection. Front Immunol. 2022; 13: 939033.
- Zhong C, Zheng M, Zhu J. Lymphoid tissue inducer-A divergent member of the ILC family. Cytokine Growth Factor Rev. 2018; 42: 5-12.
- Kiekens L, Wahlen S, Persyn E, De Vos Z, Taghon T, Vandekerckhove B, et al. T-BET drives the conversion of human type 3 innate lymphoid cells into functional NK cells. Front Immunol. 2022; 13: 975778.
- Sonnenberg GF, Fouser LA, Artis D. Border patrol: regulation of immunity, inflammation and tissue homeostasis at barrier surfaces by IL-22. Nat Immunol. 2011; 12: 383-90.
- Jakob MO, Spari D, Sànchez Taltavull D, Salm L, Yilmaz B, Doucet Ladevèze R, et al. ILC3s restrict the dissemination of intestinal bacteria to safeguard liver regeneration after surgery. Cell Rep. 2023; 42: 112269.
- Hamaguchi M, Okamura T, Fukuda T, Nishida K, Yoshimura Y, Hashimoto Y, et al. Group 3 Innate Lymphoid Cells Protect Steatohepatitis From High-Fat Diet Induced Toxicity. Frontiers in Immunology. 2021; 12: 648754.
- Zheng Y, Valdez PA, Danilenko DM, Hu Y, Sa SM, Gong Q, et al. Interleukin-22 mediates early host defense against attaching and effacing bacterial pathogens. Nat Med. 2008; 14: 282-9.
- Sepahi A, Liu Q, Friesen L, Kim CH. Dietary fiber metabolites regulate innate lymphoid cell responses. Mucosal Immunol. 2021; 14: 317-30.
- Chun E, Lavoie S, Fonseca-Pereira D, Bae S, Michaud M, Hoveyda HR, et al. Metabolite-Sensing Receptor Ffar2 Regulates Colonic Group 3 Innate Lymphoid Cells and Gut Immunity. Immunity. 2019; 51: 871-84.e6.
- Yang W, Yu T, Huang X, Bilotta AJ, Xu L, Lu Y, et al. Intestinal microbiota-derived short-chain fatty acids regulation of immune cell IL-22 production and gut immunity. Nat Commun. 2020; 11: 4457.

- Zeng B, Shi S, Ashworth G, Dong C, Liu J, Xing F. ILC3 function as a double-edged sword in inflammatory bowel diseases. Cell Death Dis. 2019; 10: 315.
- Eken A, Singh AK, Treuting PM, Oukka M. IL-23R+ innate lymphoid cells induce colitis via interleukin-22-dependent mechanism. Mucosal Immunol. 2014; 7: 143-54.
- Siegler JJ, Correia MP, Hofman T, Prager I, Birgin E, Rahbari NN, et al. Human ILC3 Exert TRAIL-Mediated Cytotoxicity Towards Cancer Cells. Front Immunol. 2022; 13: 742571.
- Kirchberger S, Royston DJ, Boulard O, Thornton E, Franchini F, Szabady RL, et al. Innate lymphoid cells sustain colon cancer through production of interleukin-22 in a mouse model. J Exp Med. 2013; 210: 917-31.
- Huber S, Gagliani N, Zenewicz LA, Huber FJ, Bosurgi L, Hu B, et al. IL-22BP is regulated by the inflammasome and modulates tumorigenesis in the intestine. Nature. 2012; 491: 259-63.
- Chung AS, Wu X, Zhuang G, Ngu H, Kasman I, Zhang J, et al. An interleukin-17-mediated paracrine network promotes tumor resistance to anti-angiogenic therapy. Nat Med. 2013; 19: 1114-23.
- Schumacher TN, Thommen DS. Tertiary lymphoid structures in cancer. Science. 2022; 375: eabf9419.
- Nussbaum K, Burkhard SH, Ohs I, Mair F, Klose CSN, Arnold SJ, et al. Tissue microenvironment dictates the fate and tumor-suppressive function of type 3 ILCs. J Exp Med. 2017; 214: 2331-47.
- 101. Shields JD, Kourtis IC, Tomei AA, Roberts JM, Swartz MA. Induction of lymphoidlike stroma and immune escape by tumors that express the chemokine CCL21. Science. 2010; 328: 749-52.
- 102. Feng L, Guo J, Wang S, Liu X, Ge N, Wang G, et al. Endoscopic Transmural Drainage and Necrosectomy in Acute Necrotizing Pancreatitis: A Review. J Transl Int Med. 2021; 9: 168-76.
- Panigrahy D, Gilligan MM, Serhan CN, Kashfi K. Resolution of inflammation: An organizing principle in biology and medicine. Pharmacol Ther. 2021; 227: 107879.
- Kay J, Thadhani E, Samson L, Engelward B. Inflammation-induced DNA damage, mutations and cancer. DNA Repair (Amst). 2019; 83: 102673.
- 105. Yuvaraj S, Kumar BRP. Peroxisome Proliferator-activated Receptor-γ As A Novel and Promising Target For Treating Cancer Via Regulation of Inflammation: A Brief Review. Mini Rev Med Chem. 2022; 22: 3-14.
- Hoesel B, Schmid JA. The complexity of NF-κB signaling in inflammation and cancer. Mol Cancer. 2013; 12: 86.
- 107. Mirzaei S, Saghari S, Bassiri F, Raesi R, Zarrabi A, Hushmandi K, et al. NF-κB as a regulator of cancer metastasis and therapy response: A focus on epithelial-mesenchymal transition. J Cell Physiol. 2022; 237: 2770-95.
- Zhang W, Zhangyuan G, Wang F, Jin K, Shen H, Zhang L, et al. The zinc finger protein Miz1 suppresses liver tumorigenesis by restricting hepatocyte-driven macrophage activation and inflammation. Immunity. 2021; 54: 1168-85.e8.
- 109. Jin H, Jia Y, Yao Z, Huang J, Hao M, Yao S, et al. Hepatic stellate cell interferes with NK cell regulation of fibrogenesis via curcumin induced senescence of hepatic stellate cell. Cell Signal. 2017; 33: 79-85.
- 110. Radaeva S, Sun R, Jaruga B, Nguyen VT, Tian Z, Gao B. Natural killer cells ameliorate liver fibrosis by killing activated stellate cells in NKG2D-dependent and tumor necrosis factor-related apoptosis-inducing ligand-dependent manners. Gastroenterology. 2006; 130: 435-52.
- 111. Satoh-Takayama N, Kato T, Motomura Y, Kageyama T, Taguchi-Atarashi N, Kinoshita-Daitoku R, et al. Bacteria-Induced Group 2 Innate Lymphoid Cells in the Stomach Provide Immune Protection through Induction of IgA. Immunity. 2020; 52: 635-49.e4.
- 112. Ozga AJ, Chow MT, Luster AD. Chemokines and the immune response to cancer. Immunity. 2021; 54: 859-74.
- Li Y, Yu HP, Zhang P. CCL15 overexpression predicts poor prognosis for hepatocellular carcinoma. Hepatol Int. 2016; 10: 488-92.
- 114. Liu LZ, Zhang Z, Zheng BH, Shi Y, Duan M, Ma LJ, et al. CCL15 Recruits Suppressive Monocytes to Facilitate Immune Escape and Disease Progression in Hepatocellular Carcinoma. Hepatology. 2019; 69: 143-59.
- 115. Itatani Y, Kawada K, Fujishita T, Kakizaki F, Hirai H, Matsumoto T, et al. Loss of SMAD4 from colorectal cancer cells promotes CCL15 expression to recruit CCR1+ myeloid cells and facilitate liver metastasis. Gastroenterology. 2013; 145: 1064-75.e11.
- 116. Yin X, Han S, Song C, Zou H, Wei Z, Xu W, et al. Metformin enhances gefitinib efficacy by interfering with interactions between tumor-associated macrophages and head and neck squamous cell carcinoma cells. Cell Oncol (Dordr). 2019; 42: 459-75.
- 117. Chen RY, Yen CJ, Lin YJ, Wang JM, Tasi TF, Huang YC, et al. CPAP enhances and maintains chronic inflammation in hepatocytes to promote hepatocarcinogenesis. Cell Death Dis. 2021; 12: 983.
- Irshad S, Flores-Borja F, Lawler K, Monypenny J, Evans R, Male V, et al. RORyt(+) Innate Lymphoid Cells Promote Lymph Node Metastasis of Breast Cancers. Cancer Res. 2017; 77: 1083-96.
- Dreyer TF, Kuhn S, Stange C, Heithorst N, Schilling D, Jelsma J, et al. The Chemokine CX3CL1 Improves Trastuzumab Efficacy in HER2 Low-Expressing Cancer In Vitro and In Vivo. Cancer Immunol Res. 2021; 9: 779-89.
- 120. McMichael EL, Jaime-Ramirez AC, Guenterberg KD, Luedke E, Atwal LS, Campbell AR, et al. IL-21 Enhances Natural Killer Cell Response to Cetuximab-Coated Pancreatic Tumor Cells. Clin Cancer Res. 2017; 23: 489-502.

- Xu X, Ye L, Zhang Q, Shen H, Li S, Zhang X, et al. Group-2 Innate Lymphoid Cells Promote HCC Progression Through CXCL2-Neutrophil-Induced Immunosuppression. Hepatology. 2021; 74: 2526-43.
- 122. Jacquelot N, Seillet C, Wang M, Pizzolla A, Liao Y, Hediyeh-Zadeh S, et al. Blockade of the co-inhibitory molecule PD-1 unleashes ILC2-dependent antitumor immunity in melanoma. Nat Immunol. 2021; 22: 851-64.
- McLane LM, Abdel-Hakeem MS, Wherry EJ. CD8 T Cell Exhaustion During Chronic Viral Infection and Cancer. Annu Rev Immunol. 2019; 37: 457-95.
- Thommen DS, Schumacher TN. T Cell Dysfunction in Cancer. Cancer Cell. 2018; 33: 547-62.
- 125. Böttcher JP, Bonavita E, Chakravarty P, Blees H, Cabeza-Cabrerizo M, Sammicheli S, et al. NK Cells Stimulate Recruitment of cDC1 into the Tumor Microenvironment Promoting Cancer Immune Control. Cell. 2018; 172: 1022-37.e14.
- 126. Mocikat R, Braumüller H, Gumy A, Egeter O, Ziegler H, Reusch U, et al. Natural killer cells activated by MHC class I(low) targets prime dendritic cells to induce protective CD8 T cell responses. Immunity. 2003; 19: 561-9.
- 127. Morandi B, Mortara L, Chiossone L, Accolla RS, Mingari MC, Moretta L, et al. Dendritic cell editing by activated natural killer cells results in a more protective cancer-specific immune response. PLoS One. 2012; 7: e39170.
- Iyori M, Zhang T, Pantel H, Gagne BA, Sentman CL. TRAIL/DR5 plays a critical role in NK cell-mediated negative regulation of dendritic cell cross-priming of T cells. J Immunol. 2011; 187: 3087-95.
- 129. Wan J, Wu Y, Huang L, Tian Y, Ji X, Abdelaziz MH, et al. ILC2-derived IL-9 inhibits colorectal cancer progression by activating CD8(+) T cells. Cancer Lett. 2021; 502: 34-43.
- Trabanelli S, Chevalier MF, Martinez-Usatorre A, Gomez-Cadena A, Salomé B, Lecciso M, et al. Tumour-derived PGD2 and NKp30-B7H6 engagement drives an immunosuppressive ILC2-MDSC axis. Nat Commun. 2017; 8: 593.
- 131. Zaiss DM, van Loosdregt J, Gorlani A, Bekker CP, Gröne A, Sibilia M, et al. Amphiregulin enhances regulatory T cell-suppressive function via the epidermal growth factor receptor. Immunity. 2013; 38: 275-84.
- 132. Mortha A, Chudnovskiy A, Hashimoto D, Bogunovic M, Spencer SP, Belkaid Y, et al. Microbiota-dependent crosstalk between macrophages and ILC3 promotes intestinal homeostasis. Science. 2014; 343: 1249288.
- Tcyganov E, Mastio J, Chen E, Gabrilovich DI. Plasticity of myeloid-derived suppressor cells in cancer. Curr Opin Immunol. 2018; 51: 76-82.
- Tumino N, Fiore PF, Pelosi A, Moretta L, Vacca P. Myeloid derived suppressor cells in tumor microenvironment: Interaction with innate lymphoid cells. Semin Immunol. 2022; 61-64: 101668.
- 135. Liu C, Xie J, Lin B, Tian W, Wu Y, Xin S, et al. Pan-Cancer Single-Cell and Spatial-Resolved Profiling Reveals the Immunosuppressive Role of APOE+ Macrophages in Immune Checkpoint Inhibitor Therapy. Adv Sci (Weinh). 2024; 11: e2401061.
- Xia Y, Rao L, Yao H, Wang Z, Ning P, Chen X. Engineering Macrophages for Cancer Immunotherapy and Drug Delivery. Adv Mater. 2020; 32: e2002054.
- Pan Y, Yu Y, Wang X, Zhang T. Tumor-Associated Macrophages in Tumor Immunity. Front Immunol. 2020; 11: 583084.
- Tosello-Trampont AC, Krueger P, Narayanan S, Landes SG, Leitinger N, Hahn YS. NKp46(+) natural killer cells attenuate metabolism-induced hepatic fibrosis by regulating macrophage activation in mice. Hepatology. 2016; 63: 799-812.
- 139. O'Sullivan TE, Rapp M, Fan X, Weizman OE, Bhardwaj P, Adams NM, et al. Adipose-Resident Group 1 Innate Lymphoid Cells Promote Obesity-Associated Insulin Resistance. Immunity. 2016; 45: 428-41.
- 140. Bie Q, Zhang P, Su Z, Zheng D, Ying X, Wu Y, et al. Polarization of ILC2s in peripheral blood might contribute to immunosuppressive microenvironment in patients with gastric cancer. J Immunol Res. 2014; 2014: 923135.
- 141. Castro-Dopico T, Fleming A, Dennison TW, Ferdinand JR, Harcourt K, Stewart BJ, et al. GM-CSF Calibrates Macrophage Defense and Wound Healing Programs during Intestinal Infection and Inflammation. Cell Rep. 2020; 32: 107857.
- 142. Yang FC, Chiu PY, Chen Y, Mak TW, Chen NJ. TREM-1-dependent M1 macrophage polarization restores intestinal epithelium damaged by DSS-induced colitis by activating IL-22-producing innate lymphoid cells. J Biomed Sci. 2019; 26: 46.
- 143. Xue R, Jia K, Wang J, Yang L, Wang Y, Gao L, et al. A Rising Star in Pancreatic Diseases: Pancreatic Stellate Cells. Front Physiol. 2018; 9: 754.
- Pinho AV, Chantrill L, Rooman I. Chronic pancreatitis: a path to pancreatic cancer. Cancer Lett. 2014; 345: 203-9.
- 145. Le Cosquer G, Maulat C, Bournet B, Cordelier P, Buscail E, Buscail L. Pancreatic Cancer in Chronic Pancreatitis: Pathogenesis and Diagnostic Approach. Cancers (Basel). 2023; 15: 761.
- 146. Hunger RE, Mueller C, Z'Graggen K, Friess H, Büchler MW. Cytotoxic cells are activated in cellular infiltrates of alcoholic chronic pancreatitis. Gastroenterology. 1997; 112: 1656-63.
- 147. Iannone F, Porzia A, Peruzzi G, Birarelli P, Milana B, Sacco L, et al. Effect of surgery on pancreatic tumor-dependent lymphocyte asset: modulation of natural killer cell frequency and cytotoxic function. Pancreas. 2015; 44: 386-93.
- 148. Jewett A, Kos J, Fong Y, Ko MW, Safaei T, Perišić Nanut M, et al. NK cells shape pancreatic and oral tumor microenvironments; role in inhibition of tumor growth and metastasis. Semin Cancer Biol. 2018; 53: 178-88.
- 149. Wagner K, Schulz P, Scholz A, Wiedenmann B, Menrad A. The targeted immunocytokine L19-IL2 efficiently inhibits the growth of orthotopic pancreatic cancer. Clin Cancer Res. 2008; 14: 4951-60.

- 150. Rautela J, Huntington ND. IL-15 signaling in NK cell cancer immunotherapy. Curr Opin Immunol. 2017; 44: 1-6.
- 151. Ze X, Zou W, Li Z. Translational research in anti-pancreatic fibrosis drug discovery and development. J Transl Int Med. 2021; 9: 225-7.
- 152. Van Audenaerde JR, Marcq E, von Scheidt B, Davey AS, Oliver AJ, De Waele J, et al. Novel combination immunotherapy for pancreatic cancer: potent anti-tumor effects with CD40 agonist and interleukin-15 treatment. Clin Transl Immunology. 2020; 9: e1165.
- Xuan X, Zhou J, Tian Z, Lin Y, Song J, Ruan Z, et al. ILC3 cells promote the proliferation and invasion of pancreatic cancer cells through IL-22/AKT signaling. Clin Transl Oncol. 2020; 22: 563-75.
- Zou X, Guan C, Gao J, Shi W, Cui Y, Zhong X. Tertiary lymphoid structures in pancreatic cancer: a new target for immunotherapy. Front Immunol. 2023; 14: 1222719.
- 155. Carrega P, Loiacono F, Di Carlo E, Scaramuccia A, Mora M, Conte R, et al. NCR(+)ILC3 concentrate in human lung cancer and associate with intratumoral lymphoid structures. Nat Commun. 2015; 6: 8280.
- Shah SC, Itzkowitz SH. Colorectal Cancer in Inflammatory Bowel Disease: Mechanisms and Management. Gastroenterology. 2022; 162: 715-30.e3.
- 157. Xie H, Ruan G, Wei L, Zhang H, Ge Y, Zhang Q, et al. Comprehensive comparative analysis of prognostic value of serum systemic inflammation biomarkers for colorectal cancer: Results from a large multicenter collaboration. Front Immunol. 2022; 13: 1092498.
- Schmitt M, Greten FR. The inflammatory pathogenesis of colorectal cancer. Nat Rev Immunol. 2021; 21: 653-67.
- 159. Huang YW, Lin CW, Pan P, Shan T, Echeveste CE, Mo YY, et al. Black Raspberries Suppress Colorectal Cancer by Enhancing Smad4 Expression in Colonic Epithelium and Natural Killer Cells. Front Immunol. 2020; 11: 570683.
- 160. Sconocchia G, Eppenberger S, Spagnoli GC, Tornillo L, Droeser R, Caratelli S, et al. NK cells and T cells cooperate during the clinical course of colorectal cancer. Oncoimmunology. 2014; 3: e952197.
- Bank U, Deiser K, Plaza-Sirvent C, Osbelt L, Witte A, Knop L, et al. c-FLIP is crucial for IL-7/IL-15-dependent NKp46(+) ILC development and protection from intestinal inflammation in mice. Nat Commun. 2020; 11: 1056.
- 162. Loyon R, Jary M, Salomé B, Gomez-Cadena A, Galaine J, Kroemer M, et al. Peripheral Innate Lymphoid Cells Are Increased in First Line Metastatic Colorectal Carcinoma Patients: A Negative Correlation With Th1 Immune Responses. Front Immunol. 2019; 10: 2121.
- 163. Rocca YS, Roberti MP, Arriaga JM, Amat M, Bruno L, Pampena MB, et al. Altered phenotype in peripheral blood and tumor-associated NK cells from colorectal cancer patients. Innate Immun. 2013; 19: 76-85.
- 164. Bruno A, Bassani B, D'Urso DG, Pitaku I, Cassinotti E, Pelosi G, et al. Angiogenin and the MMP9-TIMP2 axis are up-regulated in proangiogenic, decidual NK-like cells from patients with colorectal cancer. Faseb j. 2018; 32: 5365-77.
- Jowett GM, Norman MDA, Yu TTL, Rosell Arévalo P, Hoogland D, Lust ST, et al. ILC1 drive intestinal epithelial and matrix remodelling. Nat Mater. 2021; 20: 250-9.
- 166. Huang Q, Jacquelot N, Preaudet A, Hediyeh-Zadeh S, Souza-Fonseca-Guimaraes F, McKenzie ANJ, et al. Type 2 Innate Lymphoid Cells Protect against Colorectal Cancer Progression and Predict Improved Patient Survival. Cancers (Basel). 2021; 13: 559.
- 167. Tsou AM, Yano H, Parkhurst CN, Mahlakõiv T, Chu C, Zhang W, et al. Neuropeptide regulation of non-redundant ILC2 responses at barrier surfaces. Nature. 2022; 611: 787-93.
- 168. Lin Y, Li B, Yang X, Liu T, Shi T, Deng B, et al. Non-hematopoietic STAT6 induces epithelial tight junction dysfunction and promotes intestinal inflammation and tumorigenesis. Mucosal Immunol. 2019; 12: 1304-15.
- 169. Jou E, Rodriguez-Rodriguez N, Ferreira AF, Jolin HE, Clark PA, Sawmynaden K, et al. An innate IL-25-ILC2-MDSC axis creates a cancer-permissive microenvironment for Apc mutation-driven intestinal tumorigenesis. Sci Immunol. 2022; 7: eabn0175.
- O'Donnell C, Mahmoud A, Keane J, Murphy C, White D, Carey S, et al. An antitumorigenic role for the IL-33 receptor, ST2L, in colon cancer. Br J Cancer. 2016; 114: 37-43.
- 171. Schiering C, Krausgruber T, Chomka A, Fröhlich A, Adelmann K, Wohlfert EA, et al. The alarmin IL-33 promotes regulatory T-cell function in the intestine. Nature. 2014; 513: 564-8.
- 172. Yan J, Yu J, Liu K, Liu Y, Mao C, Gao W. The Pathogenic Roles of IL-22 in Colitis: Its Transcription Regulation by Musculin in T Helper Subsets and Innate Lymphoid Cells. Front Immunol. 2021; 12: 758730.
- 173. Geremia A, Arancibia-Cárcamo CV, Fleming MP, Rust N, Singh B, Mortensen NJ, et al. IL-23-responsive innate lymphoid cells are increased in inflammatory bowel disease. J Exp Med. 2011; 208: 1127-33.
- Goc J, Lv M, Bessman NJ, Flamar AL, Sahota S, Suzuki H, et al. Dysregulation of ILC3s unleashes progression and immunotherapy resistance in colon cancer. Cell. 2021; 184: 5015-30.e16.
- 175. Wang S, Qu Y, Xia P, Chen Y, Zhu X, Zhang J, et al. Transdifferentiation of tumor infiltrating innate lymphoid cells during progression of colorectal cancer. Cell Res. 2020; 30: 610-22.
- 176. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA Cancer J Clin. 2021; 71: 209-49.

- 177. Machlowska J, Baj J, Sitarz M, Maciejewski R, Sitarz R. Gastric Cancer: Epidemiology, Risk Factors, Classification, Genomic Characteristics and Treatment Strategies. Int J Mol Sci. 2020; 21: 4012.
- 178. Choi IJ, Kim CG, Lee JY, Kim YI, Kook MC, Park B, et al. Family History of Gastric Cancer and Helicobacter pylori Treatment. N Engl J Med. 2020; 382: 427-36.
- 179. Li B, Jiang Y, Li G, Fisher GA, Jr., Li R. Natural killer cell and stroma abundance are independently prognostic and predict gastric cancer chemotherapy benefit. JCI Insight. 2020; 5: e136570.
- 180. Yang CA, Scheibenbogen C, Bauer S, Kleinle C, Wex T, Bornschein J, et al. A frequent Toll-like receptor 1 gene polymorphism affects NK- and T-cell IFN-γ production and is associated with Helicobacter pylori-induced gastric disease. Helicobacter. 2013; 18: 13-21.
- Rudnicka K, Miszczyk E, Matusiak A, Walencka M, Moran AP, Rudnicka W, et al. Helicobacter pylori-driven modulation of NK cell expansion, intracellular cytokine expression and cytotoxic activity. Innate Immun. 2015; 21: 127-39.
- 182. Betten A, Bylund J, Christophe T, Boulay F, Romero A, Hellstrand K, et al. A proinflammatory peptide from Helicobacter pylori activates monocytes to induce lymphocyte dysfunction and apoptosis. J Clin Invest. 2001; 108: 1221-8.
- 183. Gur C, Maalouf N, Gerhard M, Singer BB, Emgård J, Temper V, et al. The Helicobacter pylori HopQ outermembrane protein inhibits immune cell activities. Oncoimmunology. 2019; 8: e1553487.
- 184. Liu X, Cao K, Xu C, Hu T, Zhou L, Cao D, et al. GATA-3 augmentation down-regulates Connexin43 in Helicobacter pylori associated gastric carcinogenesis. Cancer Biol Ther. 2015; 16: 987-96.
- 185. Li T, Zhang Q, Jiang Y, Yu J, Hu Y, Mou T, et al. Gastric cancer cells inhibit natural killer cell proliferation and induce apoptosis via prostaglandin E2. Oncoimmunology. 2016; 5: e1069936.
- 186. Yang Y, Lee JH, Kim KY, Song HK, Kim JK, Yoon SR, et al. The interferon-inducible 9-27 gene modulates the susceptibility to natural killer cells and the invasiveness of gastric cancer cells. Cancer Lett. 2005; 221: 191-200.
- 187. Kono K, Takahashi A, Ichihara F, Sugai H, Fujii H, Matsumoto Y. Impaired antibody-dependent cellular cytotoxicity mediated by herceptin in patients with gastric cancer. Cancer Res. 2002; 62: 5813-7.
- 188. Wang W, Jin J, Dai F, Long Z, Liu X, Cai H, et al. Interleukin-15 suppresses gastric cancer liver metastases by enhancing natural killer cell activity in a murine model. Oncol Lett. 2018; 16: 4839-46.
- 189. Chen Y, Chen B, Yang T, Xiao W, Qian L, Ding Y, et al. Human fused NKG2D-IL-15 protein controls xenografted human gastric cancer through the recruitment and activation of NK cells. Cell Mol Immunol. 2017; 14: 293-307.
- 190. Hyakudomi M, Matsubara T, Hyakudomi R, Yamamoto T, Kinugasa S, Yamanoi A, et al. Increased expression of fractalkine is correlated with a better prognosis and an increased number of both CD8+ T cells and natural killer cells in gastric adenocarcinoma. Ann Surg Oncol. 2008; 15: 1775-82.
- Busada JT, Ramamoorthy S, Cain DW, Xu X, Cook DN, Cidlowski JA. Endogenous glucocorticoids prevent gastric metaplasia by suppressing spontaneous inflammation. J Clin Invest. 2019; 129: 1345-58.
- 192. Busada JT, Peterson KN, Khadka S, Xu X, Oakley RH, Cook DN, et al. Glucocorticoids and Androgens Protect From Gastric Metaplasia by Suppressing Group 2 Innate Lymphoid Cell Activation. Gastroenterology. 2021; 161: 637-52.e4.
- 193. Fu W, Wang W, Zhang J, Zhao Y, Chen K, Wang Y, et al. Dynamic change of circulating innate and adaptive lymphocytes subtypes during a cascade of gastric lesions. J Leukoc Biol. 2022; 112: 931-8.
- 194. Long ZW, Yu HM, Wang YN, Liu D, Chen YZ, Zhao YX, et al. Association of IL-17 polymorphisms with gastric cancer risk in Asian populations. World J Gastroenterol. 2015; 21: 5707-18.
- 195. Ringelhan M, Pfister D, O'Connor T, Pikarsky E, Heikenwalder M. The immunology of hepatocellular carcinoma. Nat Immunol. 2018; 19: 222-32.
- Wang G, Li J, Bojmar L, Chen H, Li Z, Tobias GC, et al. Tumour extracellular vesicles and particles induce liver metabolic dysfunction. Nature. 2023; 618: 374-82.
- 197. Wang X, He Q, Zhou C, Xu Y, Liu D, Fujiwara N, et al. Prolonged hypernutrition impairs TREM2-dependent efferocytosis to license chronic liver inflammation and NASH development. Immunity. 2023; 56: 58-77.e11.
- Llovet JM, Castet F, Heikenwalder M, Maini MK, Mazzaferro V, Pinato DJ, et al. Immunotherapies for hepatocellular carcinoma. Nat Rev Clin Oncol. 2022; 19: 151-72.
- Mikulak J, Bruni E, Oriolo F, Di Vito C, Mavilio D. Hepatic Natural Killer Cells: Organ-Specific Sentinels of Liver Immune Homeostasis and Physiopathology. Front Immunol. 2019; 10: 946.
- 200. Koda Y, Teratani T, Chu PS, Hagihara Y, Mikami Y, Harada Y, et al. CD8(+) tissue-resident memory T cells promote liver fibrosis resolution by inducing apoptosis of hepatic stellate cells. Nat Commun. 2021; 12: 4474.
- 201. Fumagalli V, Venzin V, Di Lucia P, Moalli F, Ficht X, Ambrosi G, et al. Group 1 ILCs regulate T cell-mediated liver immunopathology by controlling local IL-2 availability. Sci Immunol. 2022; 7: eabi6112.
- 202. Chew V, Chen J, Lee D, Loh E, Lee J, Lim KH, et al. Chemokine-driven lymphocyte infiltration: an early intratumoural event determining long-term survival in resectable hepatocellular carcinoma. Gut. 2012; 61: 427-38.
- 203. Sun C, Xu J, Huang Q, Huang M, Wen H, Zhang C, et al. High NKG2A expression contributes to NK cell exhaustion and predicts a poor prognosis of patients with liver cancer. Oncoimmunology. 2017; 6: e1264562.

- 204. Heinrich B, Gertz EM, Schäffer AA, Craig A, Ruf B, Subramanyam V, et al. The tumour microenvironment shapes innate lymphoid cells in patients with hepatocellular carcinoma. Gut. 2022; 71: 1161-75.
- 205. Hoechst B, Voigtlaender T, Ormandy L, Gamrekelashvili J, Zhao F, Wedemeyer H, et al. Myeloid derived suppressor cells inhibit natural killer cells in patients with hepatocellular carcinoma via the NKp30 receptor. Hepatology. 2009; 50: 799-807.
- 206. Heinrich B, Ruf B, Subramanyam V, Myojin Y, Lai CW, Craig AJ, et al. Checkpoint Inhibitors Modulate Plasticity of Innate Lymphoid Cells in Peripheral Blood of Patients With Hepatocellular Carcinoma. Front Immunol. 2022; 13: 849958.
- 207. Zecca A, Barili V, Rizzo D, Olivani A, Biasini E, Laccabue D, et al. Intratumor Regulatory Noncytotoxic NK Cells in Patients with Hepatocellular Carcinoma. Cells. 2021; 10: 614.
- McHedlidze T, Waldner M, Zopf S, Walker J, Rankin AL, Schuchmann M, et al. Interleukin-33-dependent innate lymphoid cells mediate hepatic fibrosis. Immunity. 2013; 39: 357-71.
- 209. He Y, Luo J, Zhang G, Jin Y, Wang N, Lu J, et al. Single-cell profiling of human CD127(+) innate lymphoid cells reveals diverse immune phenotypes in hepatocellular carcinoma. Hepatology. 2022; 76: 1013-29.