

1 **SERINC2-mediated serine metabolism promotes cervical cancer progression and drives T cell**
2 **exhaustion**

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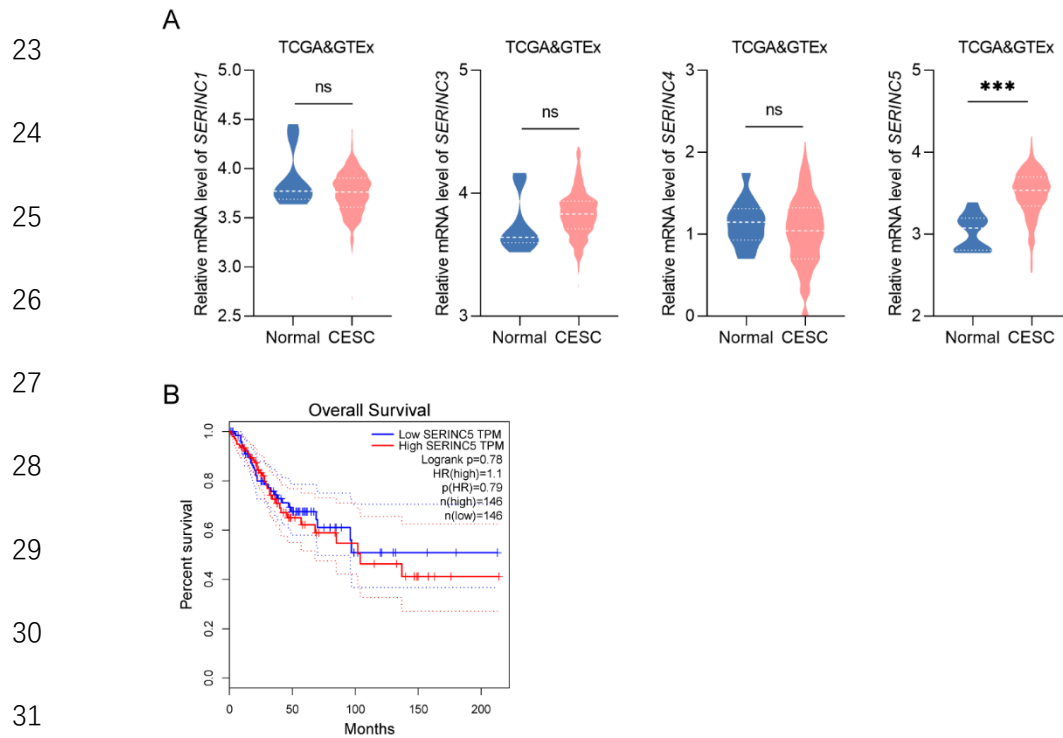
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32 **Supplementary Fig.1** Expression profile and prognosis analysis of SERINC family genes. **(A)**

33 Expression of *SERINC1*, *SERINC3*, *SERINC4* and *SERINC5* using TCGA and GTEx database. **(B)**

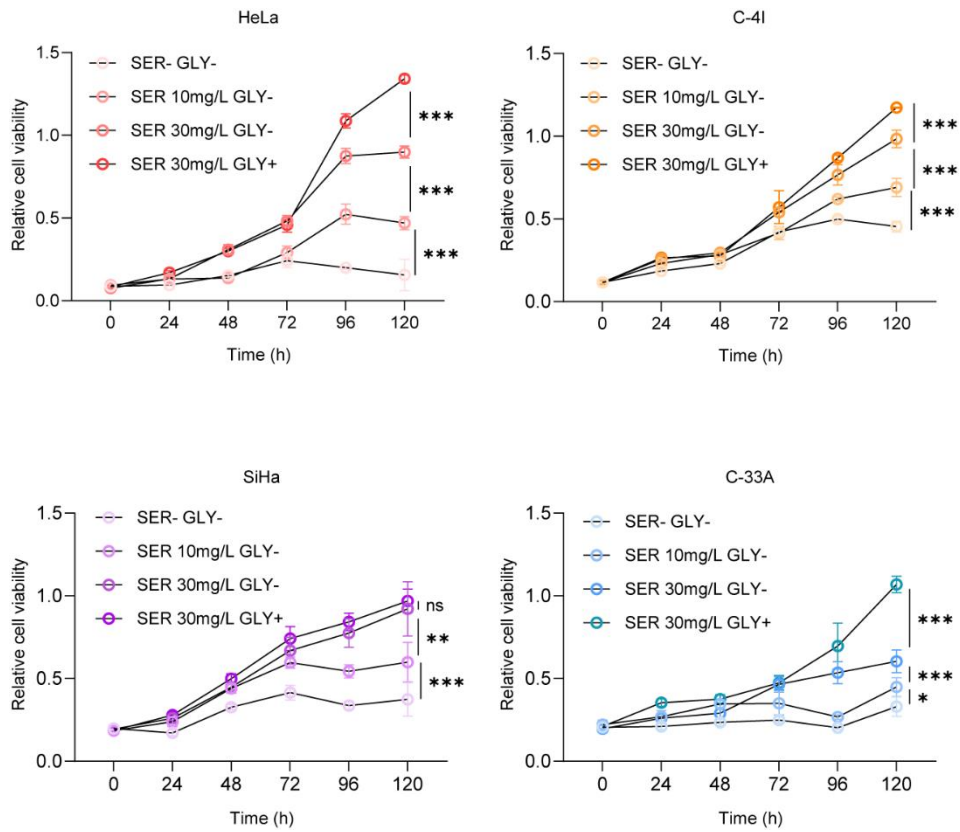
34 Kaplan-Meier analysis of OS in patients with high or low *SERINC5* expression using GEPIA online

35 server. ns: not significant, $*P < 0.05$, $**P < 0.01$, $***P < 0.001$

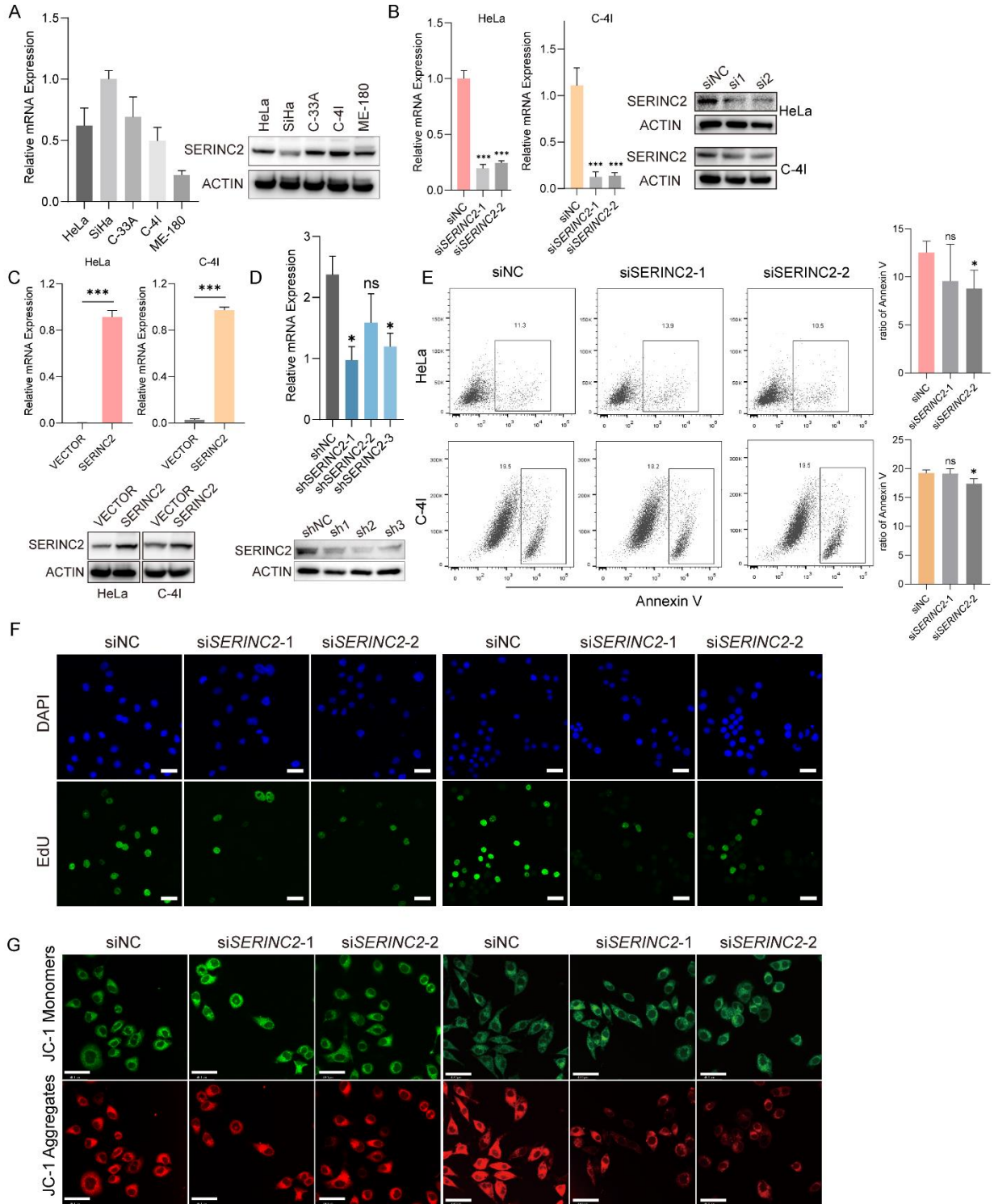
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Supplementary Fig.2 The growth of cervical cancer cells depended on serine. CCK-8 analysis of HeLa, C-4I, C-33A and SiHa cell viability under different serine concentration (serine 0mg/L, 10mg/L, 30mg/L, without glycine and serine 30mg/L with glycine 10mg/L) with the analysis. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$



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83 **Supplementary Fig.3** SERINC2 cell line expression, knockdown/overexpressed efficiency and
84 functional assay. **(A)** SERINC2 expression in HeLa, C-4I, C-33A, SiHa and ME-180 cells at mRNA and
85 protein level. **(B)** Validation of SERINC2 knockdown efficiency at mRNA and protein level in HeLa and
86 C-4I cells. **(C)** Validation of SERINC2 overexpression efficiency at mRNA and protein level in HeLa
87 and C-4I cells. **(D)** Validation and statistical analysis of knockdown efficiency at mRNA and protein
88 expression level of shNC/shSERINC2 HeLa cells. **(E)** Apoptosis assay staining the effect of
89 siNC/siSERINC2 HeLa and C-4I cells with analysis. **(F)** Representative EdU staining of SERINC2
90 knockdown HeLa and C-4I cells in separate channel. Scale bar = 50µm. **(G)** Representative JC-1 staining
91 images of SERINC2 knockdown HeLa and C-4I cells in separate channel. Scale bar = 50µm.* $P < 0.05$,
92 ** $P < 0.01$, *** $P < 0.001$

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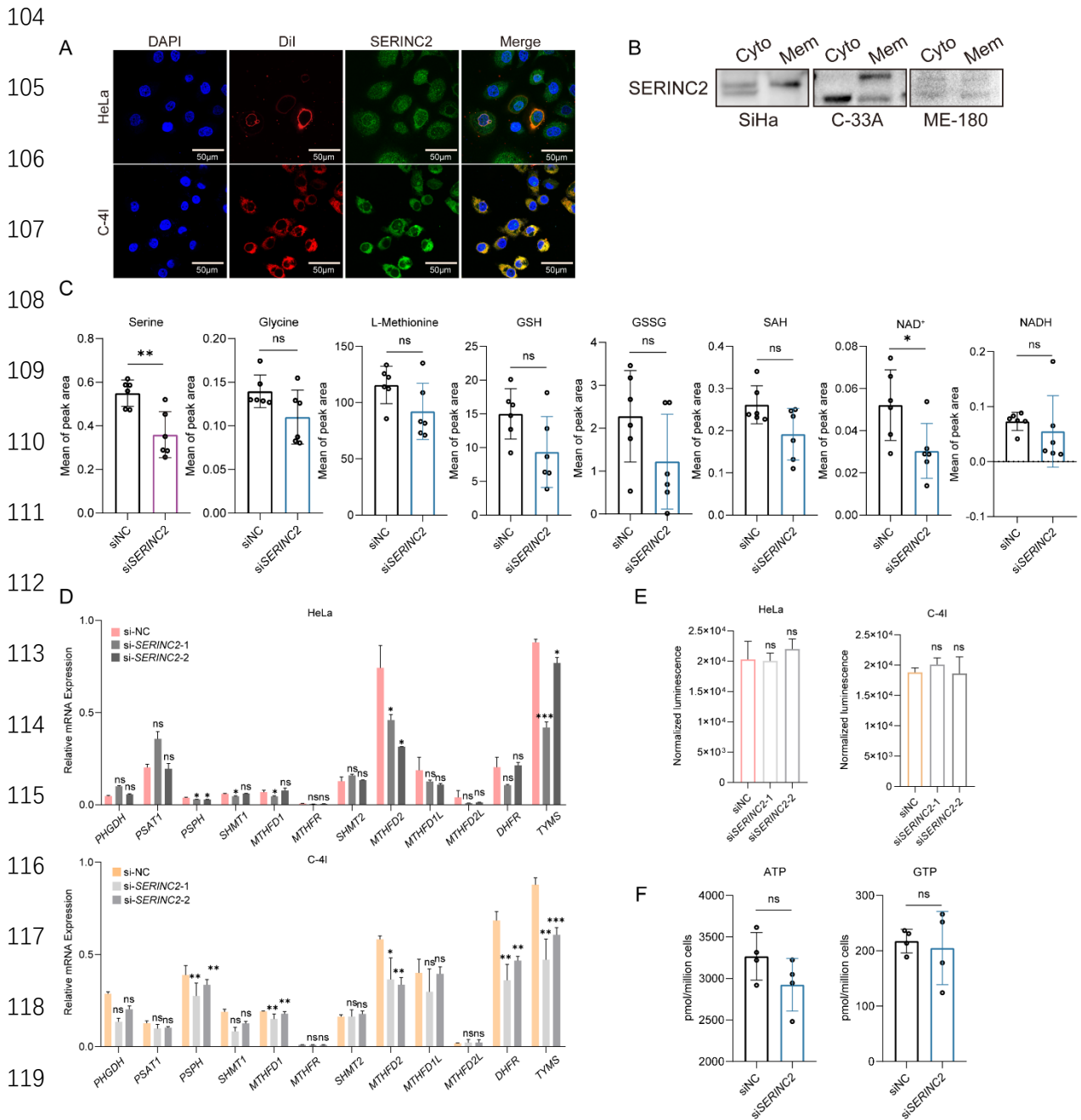
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120 **Supplementary Fig.4** SERINC2 didn't involve in SGOC metabolism. **(A)** IF staining of SERINC2
 121 expression and localization in HeLa and C-4I cells. DiI: lipophilic carbocyanine fluorescent dye used to
 122 target cell membrane. scale bar: 50 μ m. **(B)** Western blotting using protein extracts from cytoplasm and
 123 membrane to determine SERINC2 expression in SiHa, C-33A and ME-180 cells. **(C)** Western blotting
 124 was applied to detect change in the AKT-mTOR pathway after SERINC2 knockdown. Puromycin
 125 infiltration assay was used to detect the synthesis of nascent protein after SERINC2 knockdown. **(D)**

126 mRNA protein level expression of ATF in SERINC2 knockdown HeLa, C-4I cells under normal complete
127 medium conditions and serine/glycine deprived cultured HeLa and C-4I cells. The serine withdrawal
128 time was 24h. **(E)** Relative quantification of serine derived SGOC metabolites after knockdown of
129 SERINC2 using LC-MS. **(F)** mRNA level of SGOC metabolism enzyme after SERINC2 knockdown.
130 **(G)** Intracellular ATP levels of SERINC2-knockdown HeLa and C-4I cells by ATP assay kit. **(H)**
131 Determination of intracellular ATP and GTP level using LC-MS after SERINC downregulation in HeLa
132 cells. ns: not significant, * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$

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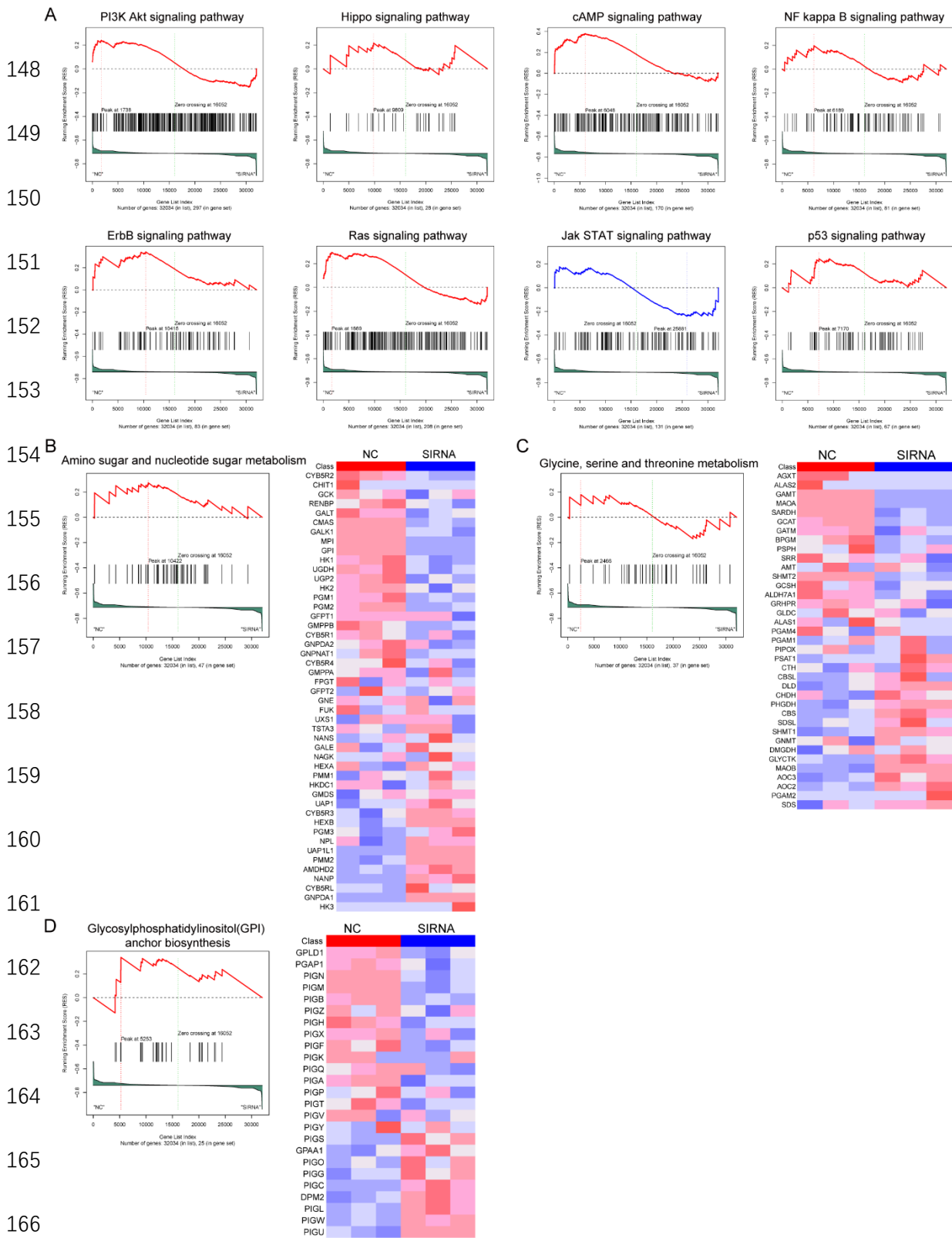
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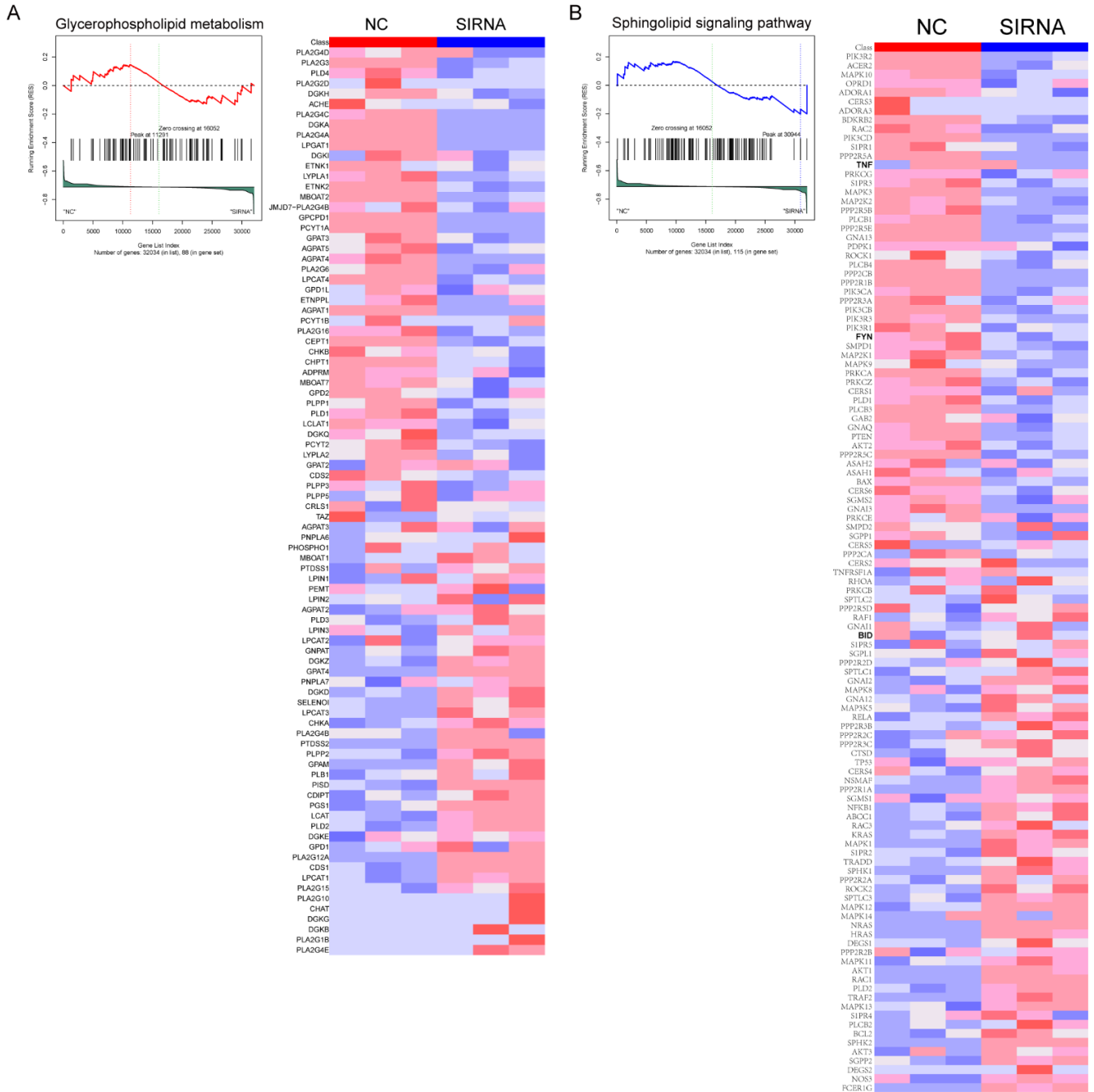
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167 **Supplementary Fig.5** Pathway enrichment analysis of SERINC2-knockdown HeLa cells. (A) GSEA
 168 analysis revealed several critical pathways that enriched in siSERINC2 HeLa cells compared to siNC
 169 HeLa cells. (B-D) GSEA analysis and gene expression profiles in siSERINC2 HeLa cells versus siNC

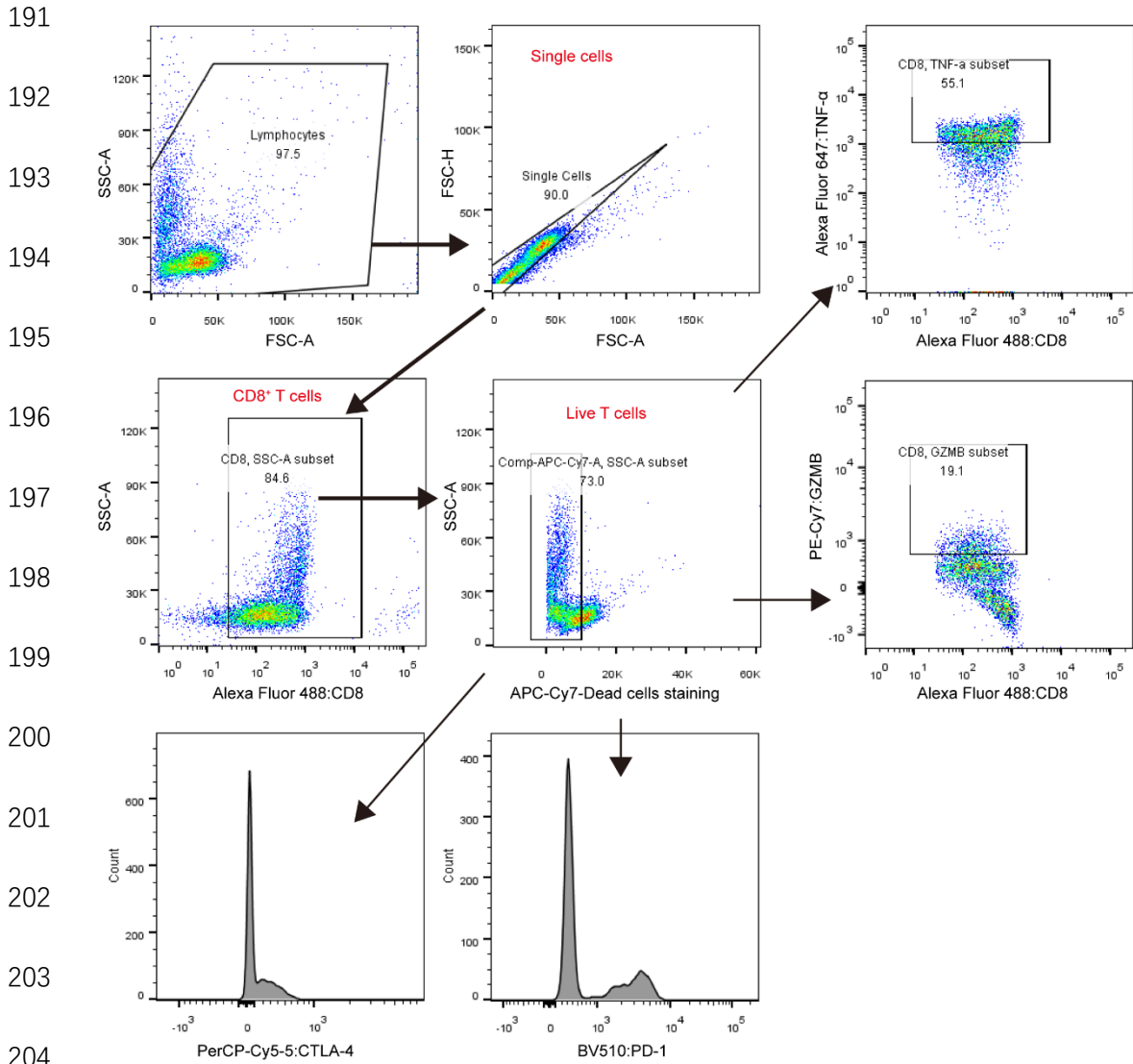


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188 **Supplementary Fig.6** Pathway enrichment analysis of SERINC2-knockdown HeLa cells. **(A, B)** GSEA

189 analysis and gene expression profiles in siSERINC2 HeLa cells versus siNC HeLa cells.

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Supplementary Fig.7 Gating strategy for CD8⁺ T cells functional assay.

210 Supplementary Table1 Primers used in this study.

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	Forward Primer (5'-3')	Reverse Primer (5'-3')
<i>ATF4</i>	TCCAACAACAGCAAGGAGGA	TACCCAACAGGGCATCCAAG
<i>PHGDH</i>	CTGCGGAAAGTGCTCATCAGT	TGGCAGAGCGAACAATAAGGC
<i>PSAT1</i>	TGCCGCACTCAGTGTTGTTAG	GCAATTCCCGCACAAAGATTCT
<i>PSPH</i>	GAGGACGCGGTGTCAGAAAT	GGTTGCTCTGCTATGAGTCTCT
<i>SHMT1</i>	CTGGCACAACCCCTCAAAGA	AGGCAATCAGCTCCAATCCAA
<i>MTHFD1</i>	AGGATGTGGATGGATTGACTAGC	CCCTTAGGCGTACAAGGAATG
<i>MTHFR</i>	CCGCCGTGAACTACTGTGG	AGATGGCCCGTGATCTCCTC
<i>SHMT2</i>	CATCGTCACCACCACTACTCACAAG	CAGGGATGGGAACACGGCAAAG
<i>MTHFD2</i>	GATCCTGGTTGGCGAGAATCC	TCTGGAAGAGGCAACTGAACA
<i>MTHFD1L</i>	GGCTCTGTATAATCGGCTGGTTCC	CTCTTCTGTCAGTGTGCTCGGATC
<i>MTHFD2L</i>	CAGTTACCACTACCAGACCACG	GGCAGGTATGAGAGAATGCTGA
<i>DHFR</i>	GAGAACTCAAGGAACCTCCACAAGG	CAGAACTGCCACCAACTATCCAGAC
<i>TYMS</i>	GGAGTGAAAATCTGGGATGCC	ACTGGAAGCCATAAACTGGGC

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213 Supplementary Table 2 Antibody used in this study.

	Catalogue Number	Company
Anti-mTOR	2972S	Cell Signaling Technology
Anti-p-mTOR(S2481)	2974T	Cell Signaling Technology
Anti-AKT	9272S	Cell Signaling Technology

Anti-p-AKT(S473)	9271T	Cell Signaling Technology
Anti-Puromycin	MABE343	Merck
Anti-ATF4	10835-1-AP	Proteintech
Anti-Na,K-ATPase	GB11400-100	Servicebio

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216 Cell membrane and cytosol protein extraction assay

217 Cell membrane and cytosol protein extraction was performed using Membrane and Cytosol Protein

218 Extraction Kit (Beyotime, China, Cat#P0033) according to manufacturer's guideline. Briefly, a total of

219 1×10^7 cells were harvested and resuspended in reagent A containing phenylmethanesulfonyl fluoride

220 (PMSF, Beyotime, China, Cat#ST506). The samples were subjected to 2 cycles of freeze and thaw by

221 liquid nitrogen, followed by centrifugation at 700 g for 10 mins in 4 °C. The supernatant was then

222 centrifuged at 14000 g for 30 mins and the precipitation was resuspended in reagent B. After ice-bath for

223 40 mins, samples were centrifuged at 14000 g for 5 mins and the supernatant was maintained at -80°C

224 or prepared for immunoblotting sample loading.