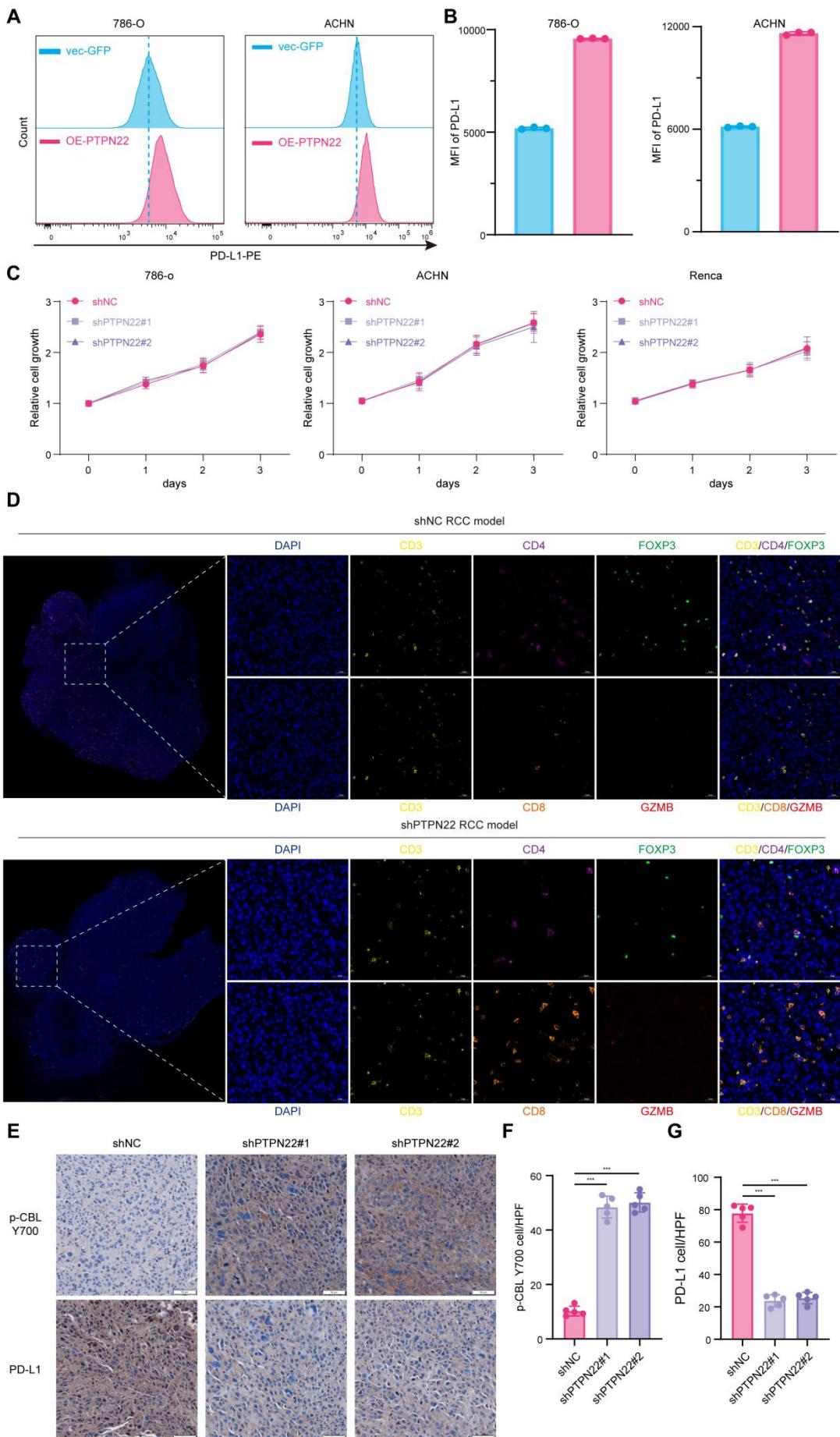
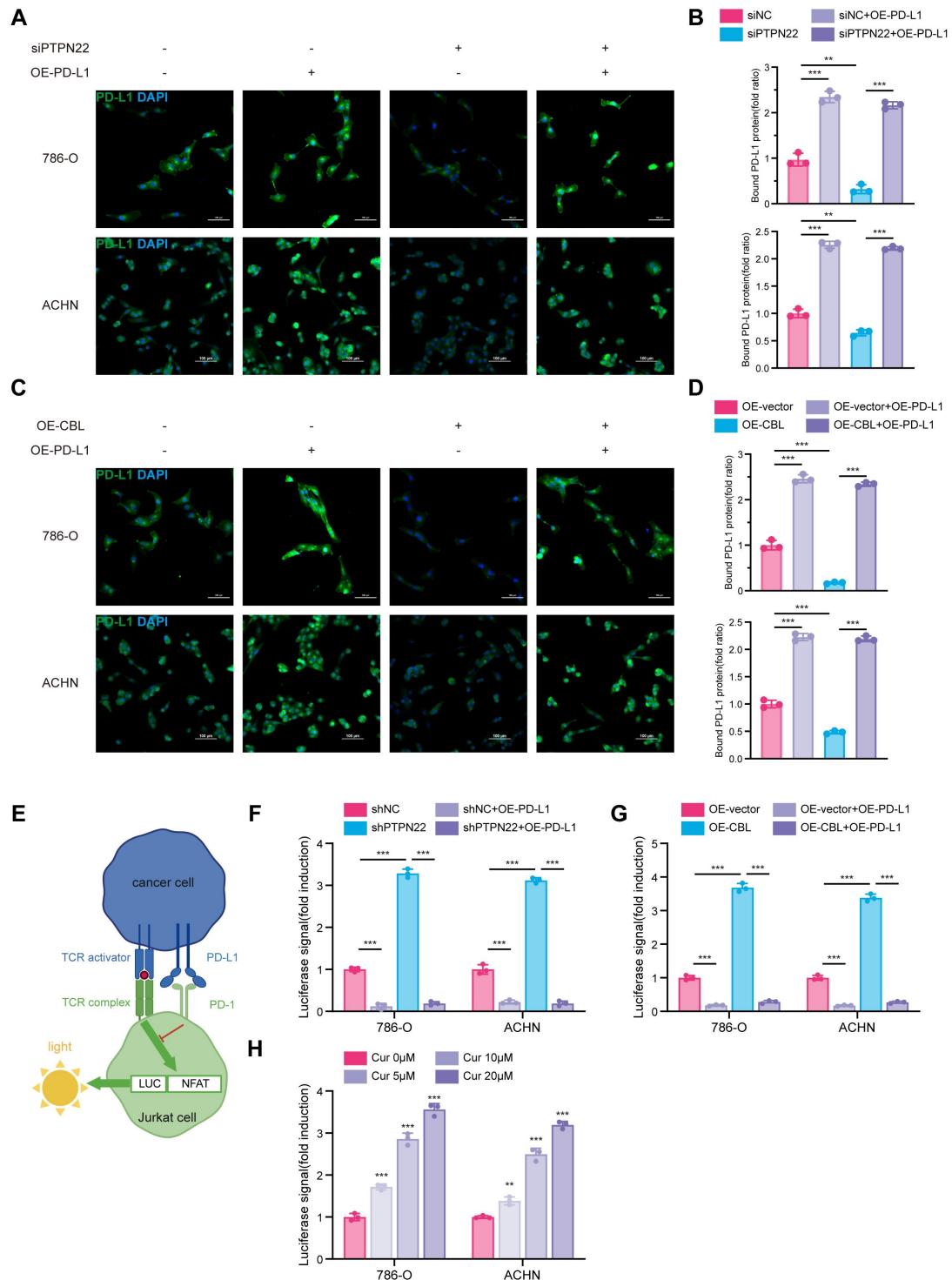


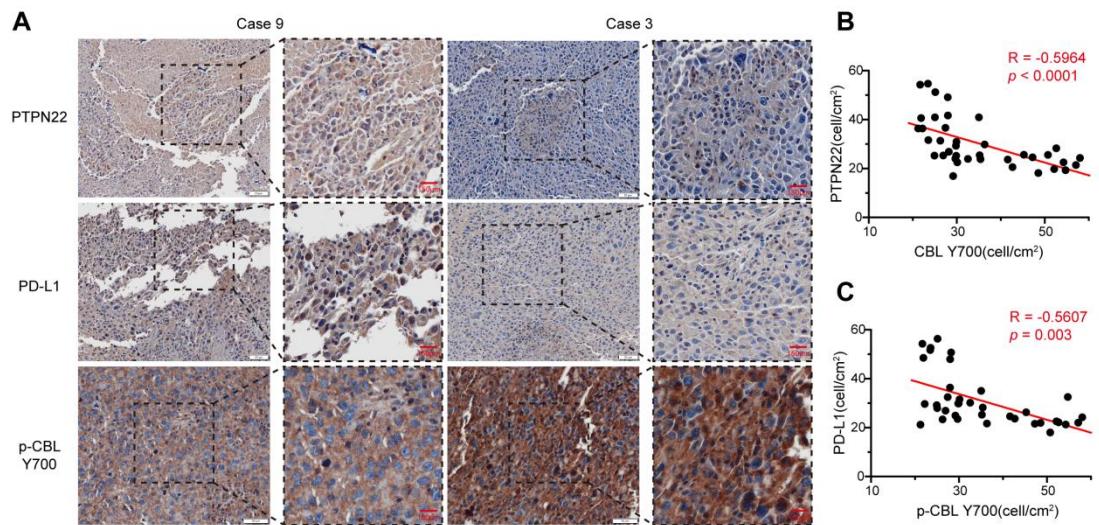
**Figure S1.** PTPN22 is overexpressed and is associated with immunosuppression in RCC. A) Representative IHC images for PTPN22 expression in normal and tumor tissues. Scale bar = 150  $\mu$ m. B) Correlation heatmap of PTPN22 between differential immune fraction. C) The survival of RCC patients stratified by the expression of PTPN22 or PD-L1 was compared by two-sided log-rank analysis. ns, not significant;  $*p < 0.05$ ,  $**p < 0.01$  and  $***p < 0.001$ .



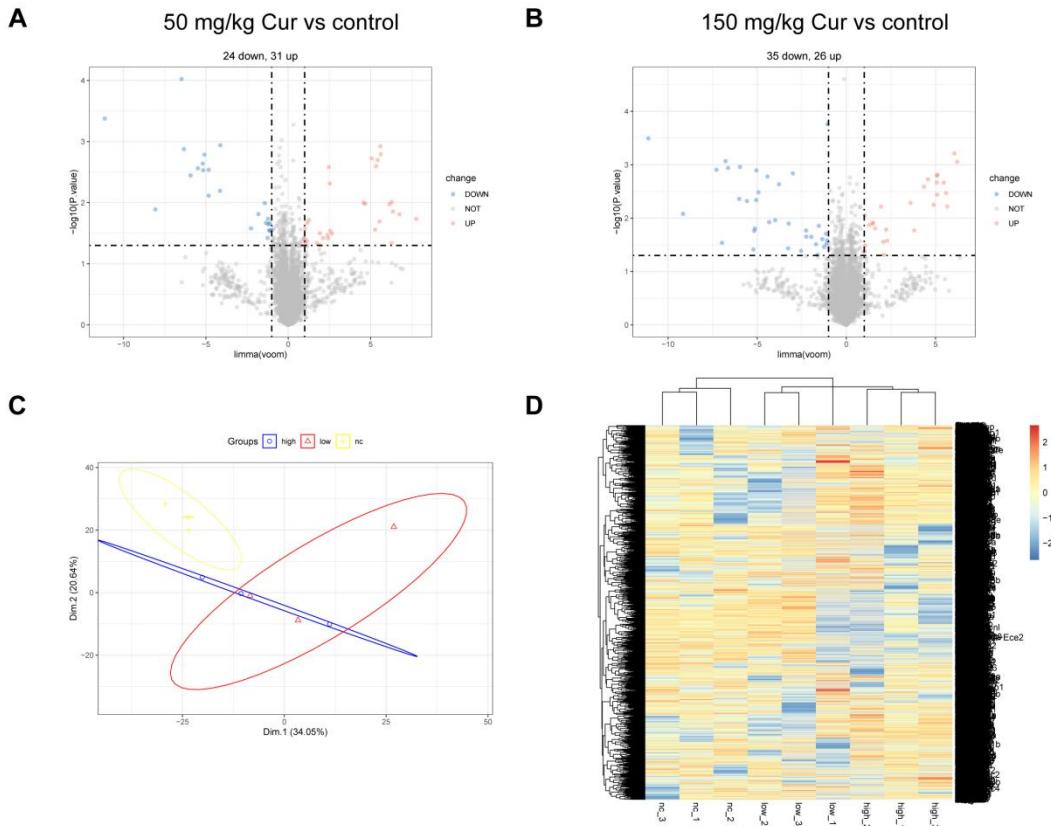
**Figure S2.** PTPN22 influences tumor growth by modulating T cell infiltration in the tumor immune microenvironment. A) Flow cytometry analysis of PD-L1 in 786-O and ACHN cells after PTPN22 overexpression. B) Statistical analysis of PD-L1 MFI. C) CCK8 analysis assays of RCC cells with or without shPTPN22. D) Representative images of mIF for DAPI (blue), CD3 (yellow), CD4 (purple), FOXP3 (green), CD8 (orange), GZMB (red). E-G) Representative IHC staining images (left panel) and quantification (right panel) for PD-L1 and p-CBL Y700 of BALB/c mice with indicated treatment. ns, not significant;  $*p < 0.05$ ,  $**p < 0.01$  and  $***p < 0.001$ .



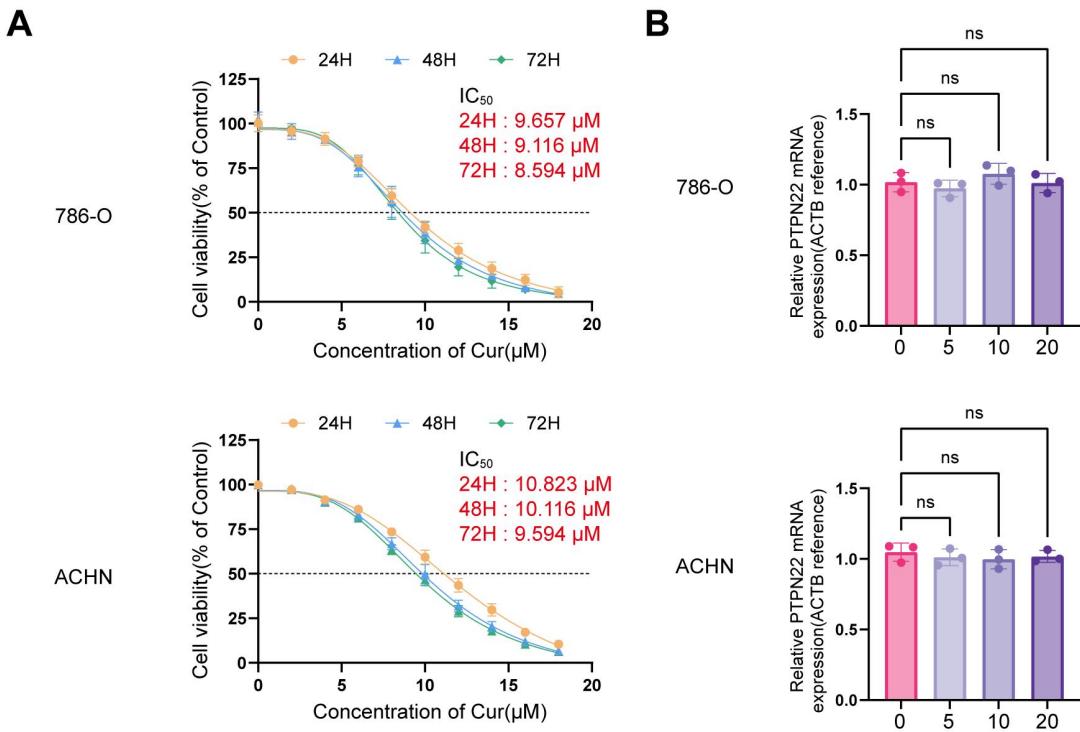
**Figure S3.** Silencing PTPN22 reduces the binding between tumor cells and PD-1, and promotes T cell activation. A, B) PD-L1/PD-1 binding assay (left panel) and quantification (right panel) in RCC cells with indicated treatment. C, D) PD-L1/PD-1 binding assay (left panel) and quantification (right panel) in RCC cells with indicated treatment. E) Schematic diagram of the PD-L1/PD-1 blockade assay. F-H) The statistical analysis of PD-L1/PD-1 blockade assays in different treatment groups. ns, not significant; \* $p < 0.05$ , \*\* $p < 0.01$  and \*\*\* $p < 0.001$ .



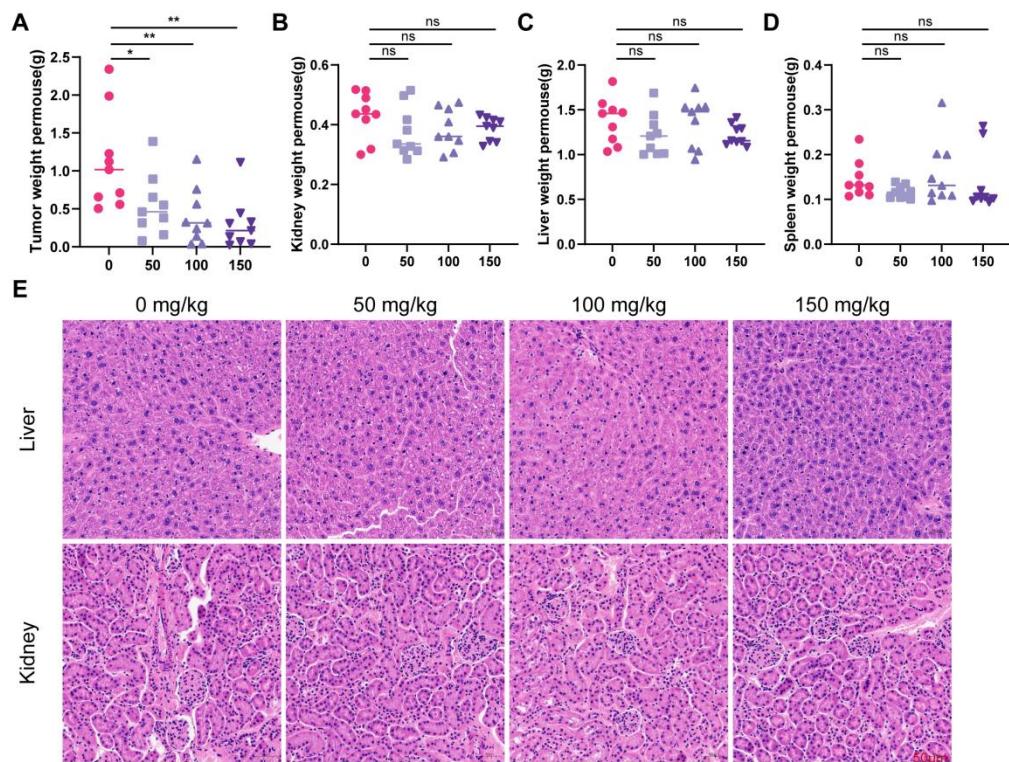
**Figure S4.** Validation of the correlation among PTPN22, p-CBL Y700, and PD-L1 in tumor tissues. A) Representative immunohistochemistry (IHC) images of PTPN22, PD-L1 and p-CBL Y700 expression in patient-derived tissues. B) Correlation analysis of PTPN22 and p-CBL Y700 protein levels in patient-derived tissues.



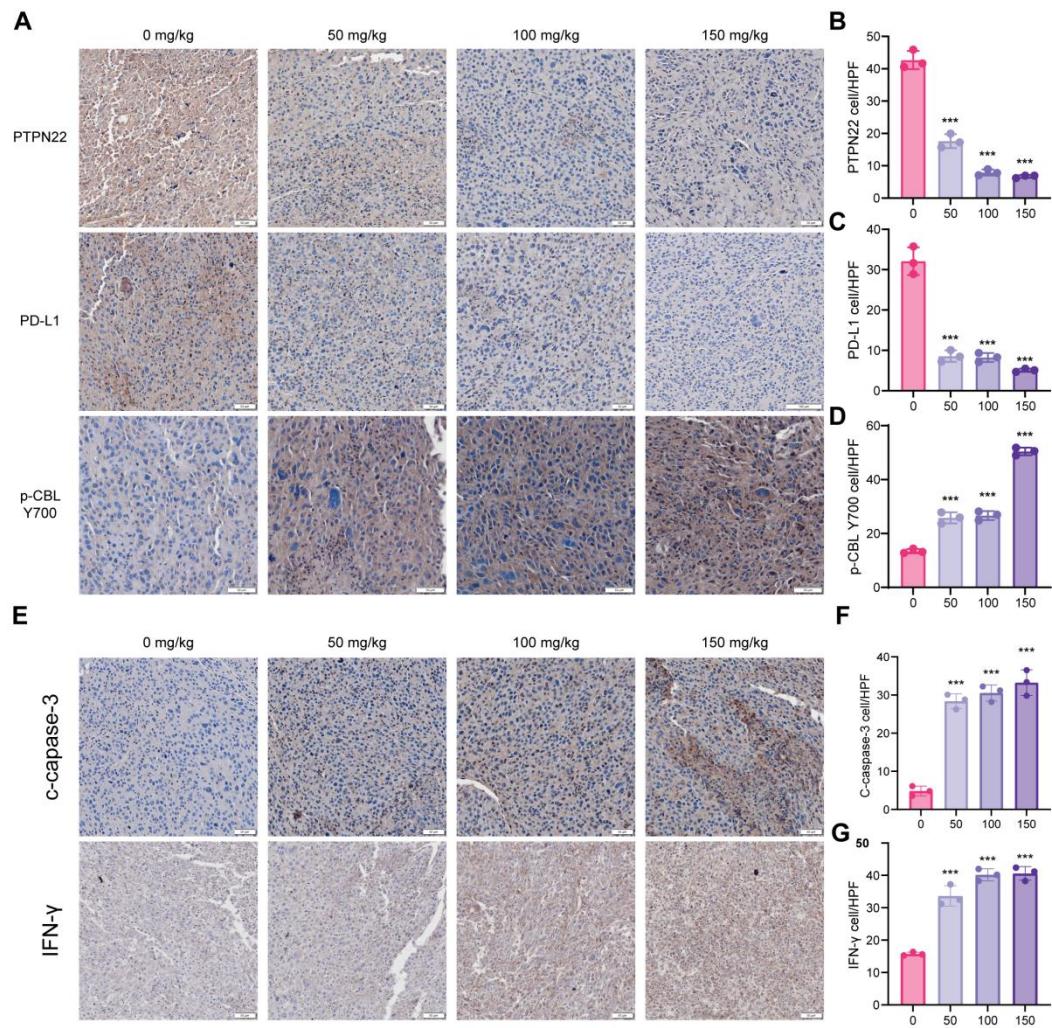
**Figure S5.** Proteomic analysis of Cur effects after treatment of Renca tumor in vivo. A) Volcano plot of DEPs analysis between 50 mg/kg Cur treatment with solvent control. B) Volcano plot of DEPs analysis between 150 mg/kg Cur treatment with solvent control. C) PCA showing the expression of proteins in Cur-treated tumors. D) Heatmap showing the expression of proteins in Cur-treated tumors.



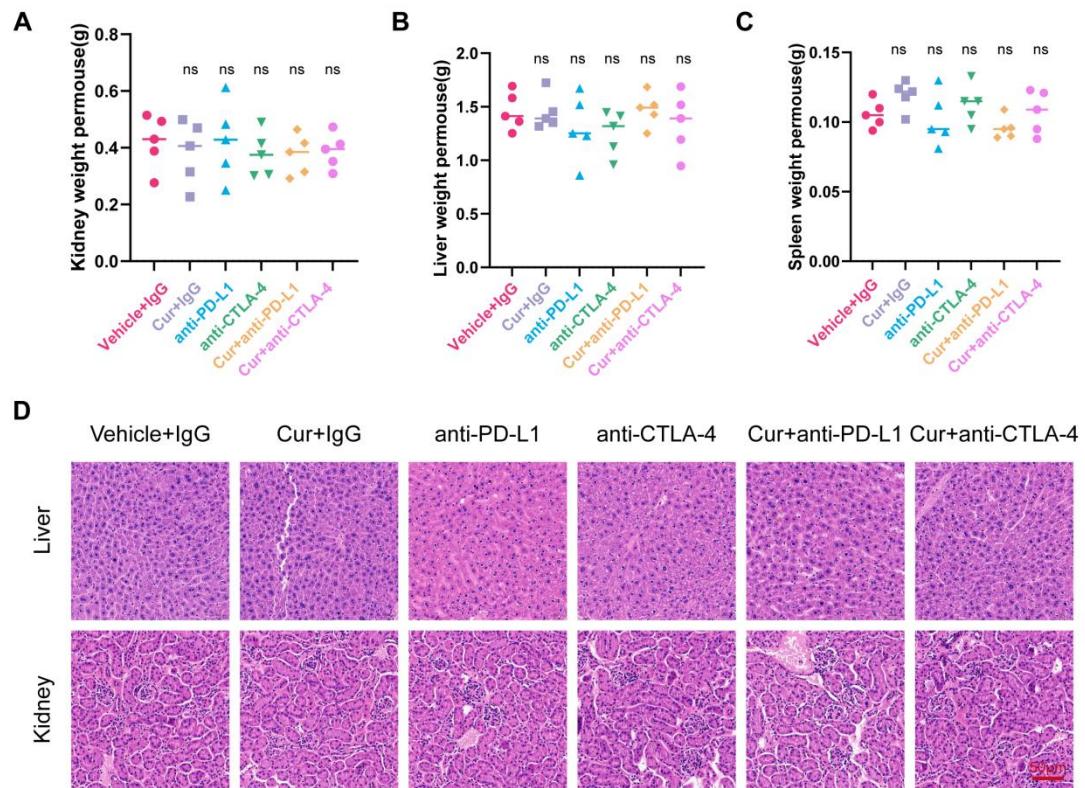
**Figure S6.** Curcumin promotes PTPN22 degradation in RCC cells. A) CCK-8 assay showing drug effects on cell viability. B) qRT-PCR showing relative PTPN22 RNA expression in 786-O and ACHN cells treated with different curcumin concentrations. ns, not significant;  $*p < 0.05$ ,  $**p < 0.01$  and  $***p < 0.001$ .



**Figure S7.** Curcumin's anti-tumor effects and safety. A-D) Tumor weight (A), kidney weight (B), liver weight (C) and spleen weight (D) of BALB/c mice with indicated treatment. (n = 9 per group). E) Representative H&E images of BALB/c mice with indicated treatment. (n = 9 per group). ns, not significant; \*p < 0.05, \*\*p < 0.01 and \*\*\*p < 0.001.



**Figure S8.** Curcumin effectively downregulates PTPN22 and PD-L1 and promotes tumor apoptosis and immunocide. A) Representative IHC staining images (left panel) and quantification (right panel) for PTPN22 and PD-L1 of BALB/c mice with indicated treatment. B) Representative IHC staining images (left panel) and quantification (right panel) for c-caspase-3 and INF-γ of BALB/c mice with indicated treatment. ns, not significant; \* $p < 0.05$ , \*\* $p < 0.01$  and \*\*\* $p < 0.001$ .



**Figure S9.** Curcumin in combination with ICIs does not cause systemic toxicity in mice. A,B,C) kidney weight (A), liver weight (B) and spleen weight (C) of BALB/c mice with indicated treatment. (n = 5 per group). D) Representative H&E images of BALB/c mice with indicated treatment. (n = 5 per group). ns, not significant; \*p < 0.05, \*\*p < 0.01 and \*\*\*p < 0.001.

**Table S1:** Clinical characteristics of renal cell carcinoma patient's cohort and applications

Name	Sex	Location	Diagnosis	Staging	Primary or	Application
					Recurrence	
M051	Male	Left Kidney	ccRCC	T <sub>2</sub> N <sub>0</sub> M <sub>0</sub>	Primary	IHC
M074	Female	Left Kidney	ccRCC	T <sub>1</sub> N <sub>0</sub> M <sub>0</sub>	Primary	IHC
M065	Female	Right Kidney	ccRCC	T <sub>3</sub> N <sub>1</sub> M <sub>0</sub>	Primary	IHC
M103	Female	Left Kidney	ccRCC	T <sub>1</sub> N <sub>1</sub> M <sub>0</sub>	Primary	IHC
M110	Female	Left Kidney	ccRCC	T <sub>1</sub> N <sub>0</sub> M <sub>0</sub>	Primary	IHC
M123	Female	Right Kidney	ccRCC	T <sub>3</sub> N <sub>0</sub> M <sub>0</sub>	Primary	IHC
M137	Male	Left Kidney	ccRCC	T <sub>2</sub> N <sub>1</sub> M <sub>0</sub>	Primary	IHC
M157	Female	Left Kidney	ccRCC	T <sub>3</sub> N <sub>0</sub> M <sub>0</sub>	Primary	IHC
M217	Male	Right Kidney	ccRCC	T <sub>3</sub> N <sub>0</sub> M <sub>0</sub>	Primary	IHC
M216	Male	Right Kidney	ccRCC	T <sub>2</sub> N <sub>1</sub> M <sub>0</sub>	Primary	IHC
M299	Male	Right Kidney	ccRCC	T <sub>1</sub> N <sub>0</sub> M <sub>0</sub>	Primary	IHC
R007	Male	Left Kidney	ccRCC	T <sub>1</sub> N <sub>0</sub> M <sub>0</sub>	Primary	IHC
R055	Female	Right Kidney	ccRCC	T <sub>1</sub> N <sub>0</sub> M <sub>0</sub>	Primary	IHC
R052	Male	Left Kidney	ccRCC	T <sub>3</sub> N <sub>1</sub> M <sub>0</sub>	Primary	IHC

**Table S2:** Sequences of shRNAs in this study and primers for quantitative real-time PCR

Name	Target Sequence	Source
shPTPN22#1	5'-GTACGGACACCTGAATCATT-3'	TranSheepBio, Shanghai, China
shPTPN22#2	5'-GAATTGATACAGCAGAGAGAA-3'	TranSheepBio, Shanghai, China
shCBL#1	5'-GTACGTATGAAGCAATGTATA-3'	TranSheepBio, Shanghai, China
shCBL#2	5'-GATTCTCCGAACGTGTCACGT-3'	TranSheepBio, Shanghai, China

rtPCR primers	Sequence(5' to 3')
PD-L1 Forward	TGGCATTGCTGAACGCATT
PD-L1 Reverse	TGCAGCCAGGTCTAATTGTTTT
PTPN22 Forward	TGCTACCCAGGGTCCTTATC
PTPN22 Reverse	AACTCCATACATGCCATGACAAT
ACTB Forward	CATGTACGTTGCTATCCAGGC
ACTB Reverse	CTCCTTAATGTCACGCACGAT

**Table S3.** Antibodies used in this study

Antibodies	Company	Cat no.
GAPDH	Affinity	AF7021
PTPN22	Abclonal	A1406
PD-L1	Proteintech	28076-1-AP
PD-L1	Huabio	HA721176
PD-L1	Huabio	HA722184
PD-L1-FITC	Abclonal	A22304
PD-L1-mCherry	Abclonal	A24570
UB	Huabio	ET1609-21
CBL	Abclonal	A0732
Pan Phospho-Tyrosine	Abclonal	AP1162
Phospho-CBL (Tyr647)	Affinity	AF3225
Phospho-CBL (Tyr700)	Abclonal	AP0780
Phospho-CBL (Tyr731)	Affinity	AF8006
Phospho-CBL (Tyr774)	Abclonal	AP0794
K48-UB	CST	4289
K63-UB	CST	5621
FLAG	Huabio	HA722780
His	Huabio	HA722798
Myc	Huabio	R1208-1
HA	Huabio	ET1611-49
PSMB5	Proteintech	19178-1-AP
FOXP3	Abclonal	A27744
GZMB	Abclonal	A2557
c-capase-3	Huabio	ET1608-64
IFN-γ	Proteintech	30293-1-AP
L/D,FVS780	BD Pharmingen	565388
CD45,FITC	BD Pharmingen	553079
CD3,PE	BD Pharmingen	553063
CD8,R718	BD Pharmingen	566985
CD4,APC	BD Pharmingen	553051
CD25,PE-CY7	BD Pharmingen	552880
FOXP3,BV421	Thermo	404-5773-82
GZMB,BV785	Biolegend	396438

**Table S4.** 114 traditional Chinese medicine monomers

Betulinic acid	Cepharanthine	18 β-Glycyrrhetic acid	(E)-Ethyl p-methoxycinnamate	β-Elemonic acid
Chelerythrine (chloride)	Ayanin	Fisetin	Columbianadin	Hydroxytyrosol
Aloperine	Osthole	4'-Hydroxychalcone	18α-Glycyrrhetic acid	Fraxetin
Lupenone	Chlorogenic acid	Asiatic acid	Liquiritin	Terpinen-4-ol
Zingerone	Macelignan	1,4-Cineole	Liquiritigenin	Morusin
Sinomenine	Solasodine	Avicularin	Glabridin	Carnosic acid
Sinomenine hydrochloride	Sarsasapogenin	Fraxinellone	Decursinol	Aucubin
Resveratrol	Naringenin	Aloesin	Wogonin	Quercetagetin
Genipin	Isoliquiritigenin	Cordycepin	4(3H)-Quinazolinone	trans-Chalcone
Decursin	Sophocarpine	Ligustrazine	Sophoricoside	Coumarin
Triptonide	Sophocarpine (monohydrate)	Caffeic acid phenethyl ester	Licochalcone D	Carvacrol
Melatonin	Rhein	Colcemid	Acacetin	8-O-Acetylharpagide
Sinapine	Salicin	Esculetin	(-)-Alkannin	Isoorientin
(-)-Epicatechin	Nobiletin	Lycorine	Britannin	Isopimpinellin
Oridonin	Oxymatrine	Lycorine (hydrochloride)	6-Hydroxycoumarin	1-beta-D-Arabinofuranosyluracil
Baohuoside I	Luteolin	Ginsenoside Rk1	Nitidine (chloride)	Isorhamnetin
Bergenin	Magnolol	Ziyuglycoside II	Gallic acid (hydrate)	Camphor
Diphyllin	Beta-Sitosterol (purity>80%)	Farrerol	Allantoin	Abietic acid
Arctigenin	Beta-Sitosterol	Pseudolaric Acid B	Wedelolactone	Catalpol
Pterostilbene	Ailanthonne	Lapachol	Salvigenin	Rhapontigenin
Jaceosidin	Artemisic acid	Isorhapontigenin	Crebanine	4-Hydroxybenzyl alcohol

Curcumin	Withaferin A	Methyl 3,4-dihydroxyben zoate	Curcumenol	Betulonic acid
Benzothiaz ole	Acetylshikoni n	Citric acid	Isoeugenol acetate	

**Table S5.** The prediction performance of 114 traditional Chinese medicine monomers docked with PTPN22 protein

monomers	Mean Vina	monomers	Mean Vina
18- $\beta$ -Glycyrrhetic acid	-8.62	Chlorogenic_acid	-7.23
Solasodine	-8.25	Ailanthonone	-7.23
Curcumin	-8.23	Lupenone	-7.20
Withaferin_A	-8.11	Betulonic_acid	-7.17
Cepharanthine	-7.52	Diphyllin	-7.13
Sarsasapogenin	-7.52	Quercetagetin	-7.10
18- $\alpha$ -Glycyrrhetic acid	-7.46	Luteolin	-7.10
Ziyuglycoside_II	-7.32	Betulinic_acid	-7.07
Liquiritin	-7.30	Glabridin	-7.07
Morusin	-7.27	Sophoricoside	-7.07