

1 **Table S1. The information of clinical samples**

<i>Liver transplant recipient information</i>	
Age (years)	51.4 ± 14,2
Sample source and type	Paraffin embedded block of liver tissue
Gender(M/F)	16/4
Time of biopsy	2023.8.1-2025.8.2
Medical history	All donors were not diagnosed with hepatitis
Donation Status	Donation after brain death

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3 **Table S2. Stard10^{-/-} knockout homozygous mice related sequences and primers**

<i>Names</i>	<i>Species</i>	<i>Sequence (5' to 3')</i>
sgRNA	<i>Mus musculus</i>	GTCAGCGTTGACAGTCAAGC
Identifying Primers	<i>Mus musculus</i>	Sense: GATAGGGAGGGCATAGACACAGTG Antisense: CAGCCCTTACCTCATTGCTCTGAC
Sequencing Primer	<i>Mus musculus</i>	CATGTGGCACACA ACTGCATATGC

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5 **Table S3. Knockdown and overexpression sequences with AAV8**

<i>Names</i>	<i>Species</i>	<i>Sequence (5' to 3')</i>
Mir30-m- <i>Stard10</i>	<i>Mus musculus</i>	AAGGTATATTGCTGTTGACAGTGAGCGGGTG CAACATGCAGACTCATAGTGAAGCCACAGAT GTATGAGTCTGCATGTTGCACCTGCCTACTGC CTCG
OE- <i>Stard10</i>	<i>Mus musculus</i>	Transcript ID: NM_001360460.1

OE-Acs11 *Mus musculus* Transcript ID: XM_017312563.3

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7 **Table S4. Primers for RT-qPCR**

<i>Names</i>	<i>Species</i>	<i>Sequence (5' to 3')</i>
STARD10	<i>Mus musculus</i>	Sense: TGAGACTTTCGACATCGCCC Antisense: CCGAGGTGGGTATTTAGGGTG
IL-6	<i>Mus musculus</i>	Sense: CCAAGAGGTGAGTGCTTCCC Antisense: CTGTTGTTTCAGACTCTCTCCCT
IL-1 β	<i>Mus musculus</i>	Sense: CCGTGGACCTTCCAGGATGA Antisense: GGGAACGTCACACACCAGCA
TNF- α	<i>Mus musculus</i>	Sense: GACGTGGAAGTGGCAGAAGAG Antisense: TTGGTGGTTTGTGAGTGTGAG
SPTLC2	<i>Mus musculus</i>	Sense: CCATGCGTCACTGGTTCTA Antisense: GTCCGAGGCTGACCATAAA
CERS2	<i>Mus musculus</i>	Sense: TCATCCCTTCTCAGTATTGGT Antisense: ATCCTTTCGCTTGACATCAG
β -ACTIN	<i>Mus musculus</i>	Sense: GTGACGTTGACATCCGTAAAGA Antisense: GCCGGACTCATCGTACTCC

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9 **Table S5. Details of the antibodies used**

<i>Antibodies</i>	<i>Supplier</i>	<i>Catalog number</i>	<i>Dilution</i>
			WB 1:1000
STARD10	Thermo Fisher	PA5-106793	IHC 1:100 IF 1:400
			WB 1:1000
YBX1	Proteintech	20339-1-AP	IF: 1:400
			WB 1:1000
β -Tubulin	Proteintech	14555-1-AP	WB 1:1000
			IHC 1:500
CD11b	Servicebio	GB11058-100	IHC 1:500
			WB 1:1000
ACSL1	Proteintech	13989-1-AP	WB 1:1000
			IHC 1:1:1000
MPO	Servicebio	GB12224-100	IHC 1:1:1000

4-HNE	Thermo Fisher	MA5-27570	IHC 1:600
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11 **Table S6. The IP-MS information of YBX1**

Accession	P62960
Gene name	Ybx1
Abundances (Scaled): F2: Sample, S_IP	100
Abundances (Normalized): F2: Sample, S_IP	10840777.88
Abundance: F2: Sample, S_IP	10840777.88
Coverage [%]	43
# Peptides	9
# PSMs	10
# Unique Peptides	9
# AAs	322
MW [kDa]	35.7
calc. pI	9.88
Score Sequest HT: Sequest HT	29.71
# Peptides (by Search Engine): Sequest HT	9
Sum PEP Score	57.161

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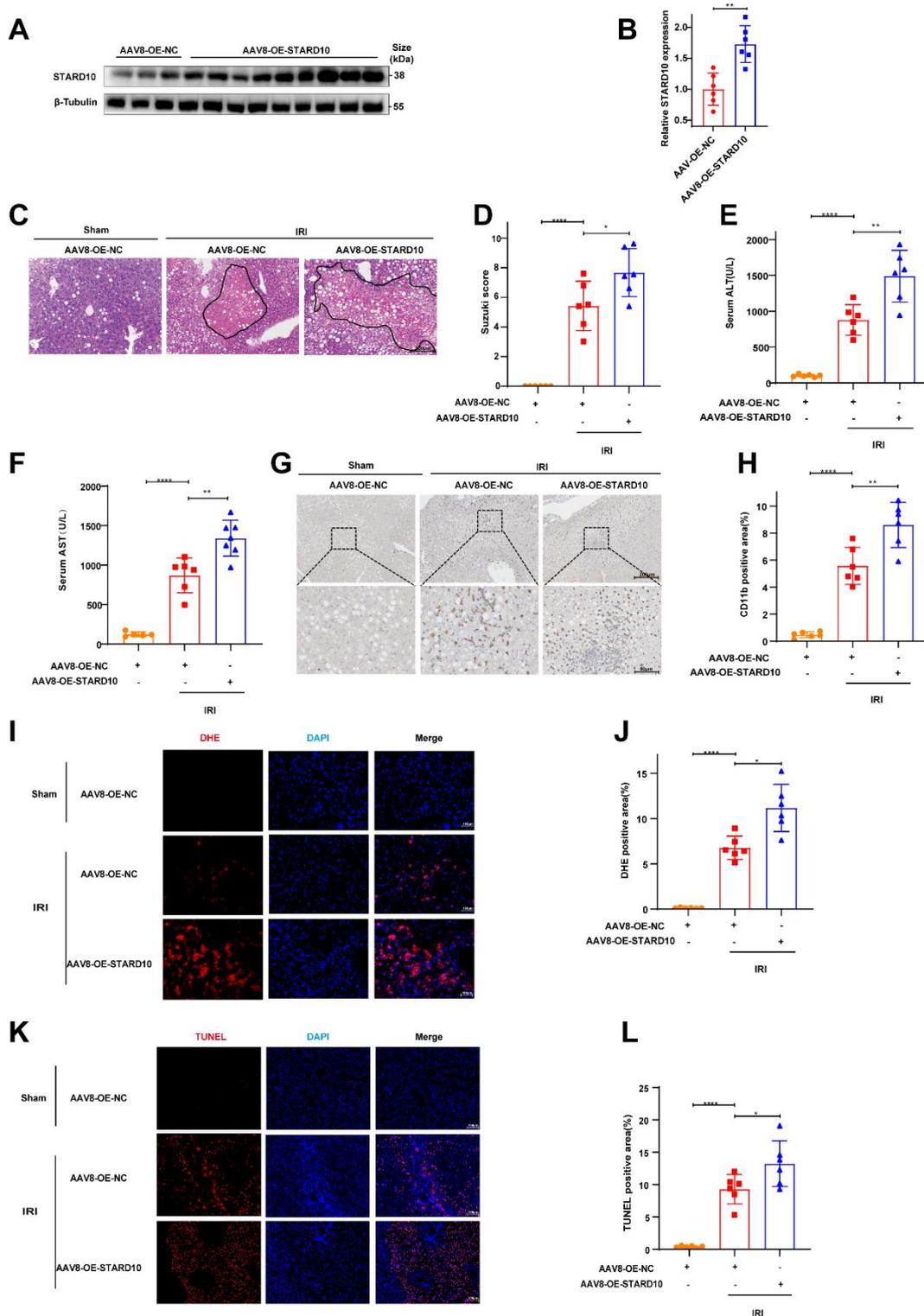
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21 Fig. S1.



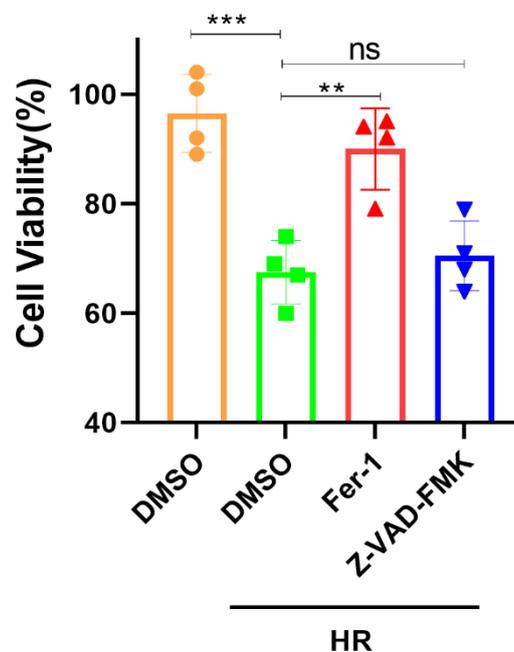
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23 Fig. S1. Overexpression of STARD10 exacerbates hepatic IRI in fatty liver. Mice received tail

24 vein injections of AAV8 vectors for STARD10 overexpression and were assessed after 8 weeks of
 25 intervention. (A-B) Western blot analysis and quantification of hepatic STARD10 levels. (C-D)
 26 Representative H&E staining images and Suzuki injury scores of liver sections. (E-F) Serum ALT
 27 and AST levels. (G-H) Representative IHC staining for CD11b and quantification of CD11b-
 28 positive cells in liver tissues. (I-J) DHE staining and quantification of ROS levels in frozen liver
 29 sections. (K-L) TUNEL staining and quantification of apoptotic cells in frozen liver sections. Data
 30 are presented as mean \pm SEM (n = 6). *P < 0.05, **P < 0.01, ***P < 0.001, ****P < 0.0001. AAV8,
 31 adeno-associated virus serotype 8; IHC, immunohistochemistry; IRI, ischemia-reperfusion injury;
 32 ROS, reactive oxygen species; STARD10, steroidogenic acute regulatory protein-related lipid
 33 transfer domain 10; TUNEL, terminal deoxynucleotidyl transferase dUTP nick end labeling.

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35 **Fig. S2.**



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37 **Fig. S2. Predominant cell death in steatotic AML12 HR injury is ferroptosis, not apoptosis.**

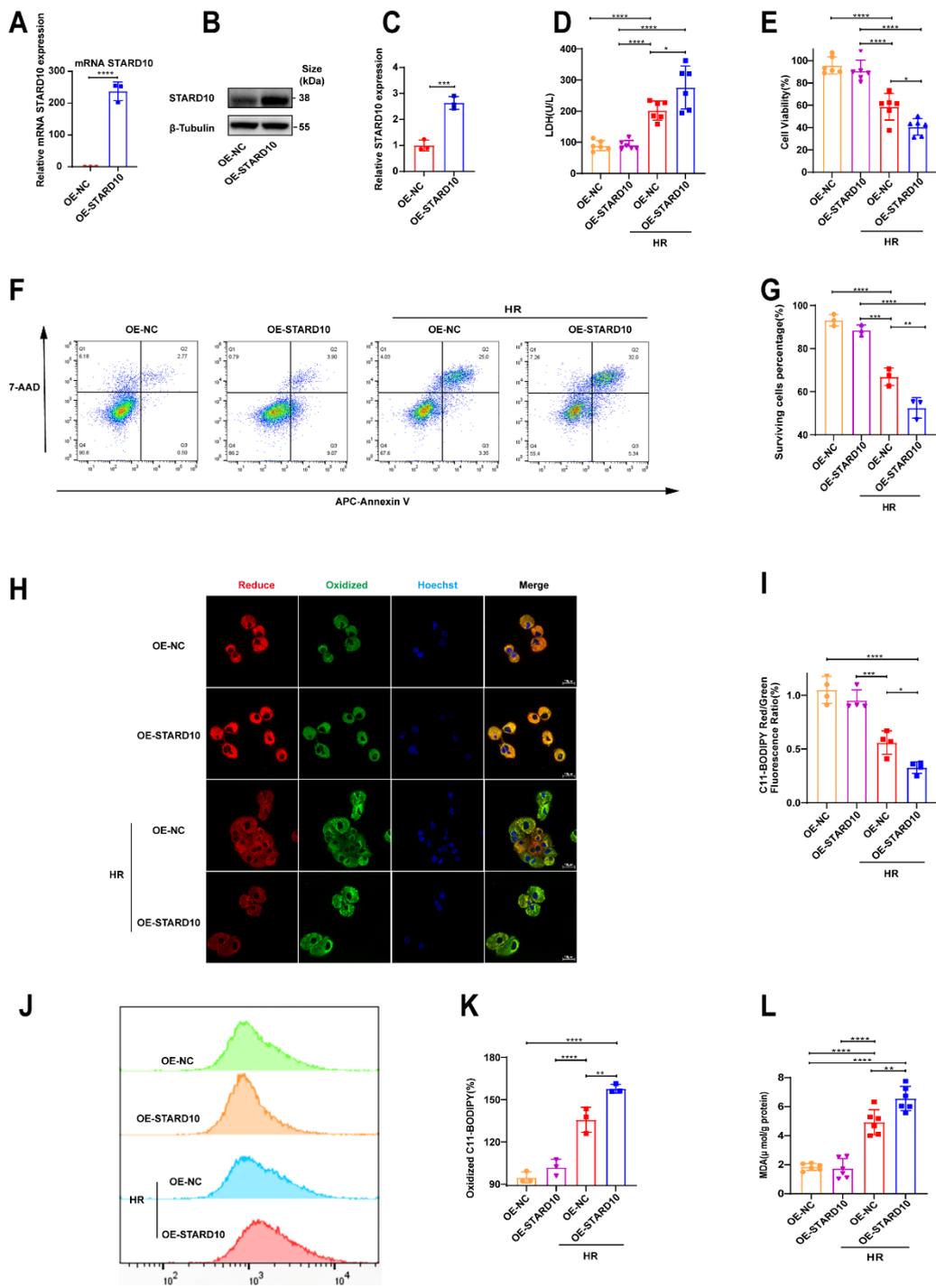
38 Steatotic AML12 cells undergoing HR injury were treated with either the ferroptosis inhibitor
 39 Ferrostatin-1 (Fer-1, 4 μ M) or the pancaspase inhibitor Z-VAD-FMK (50 μ M) during the
 40 reoxygenation phase. Cell viability was subsequently measured. Data are presented as mean \pm SEM
 41 (n = 4). **P < 0.01, ***P < 0.001. Fer-1, ferroptatin-1; Z-VAD-FMK, pancaspase inhibitor.

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45 **Fig. S3.**



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47 **Fig. S3. Overexpression of STARD10 exacerbates hypoxia-reoxygenation injury in steatotic**
 48 **AM12 cells.** After stable transfection of AML12 cells with overexpression lentivirus, (A) STARD10
 49 mRNA expression was analyzed by RT-PCR, and (B-C) STARD10 expression levels were
 50 determined by Western blotting. The following assessments were conducted: (D) LDH release, (E)

51 cell viability, and (F-G) apoptosis levels. Finally, (H-K) lipid peroxidation levels were evaluated by
52 staining cells with C11-BODIPY and analyzing them via confocal microscopy and flow cytometry,
53 as well as by measuring (L) MDA levels. Data are presented as mean \pm SEM with $n \geq 3$. *P < 0.05,
54 **P < 0.01, ***P < 0.001. PA, palmitic acid; HR, hypoxia-reoxygenation; Ctrl, control; MDA,
55 malondialdehyde.

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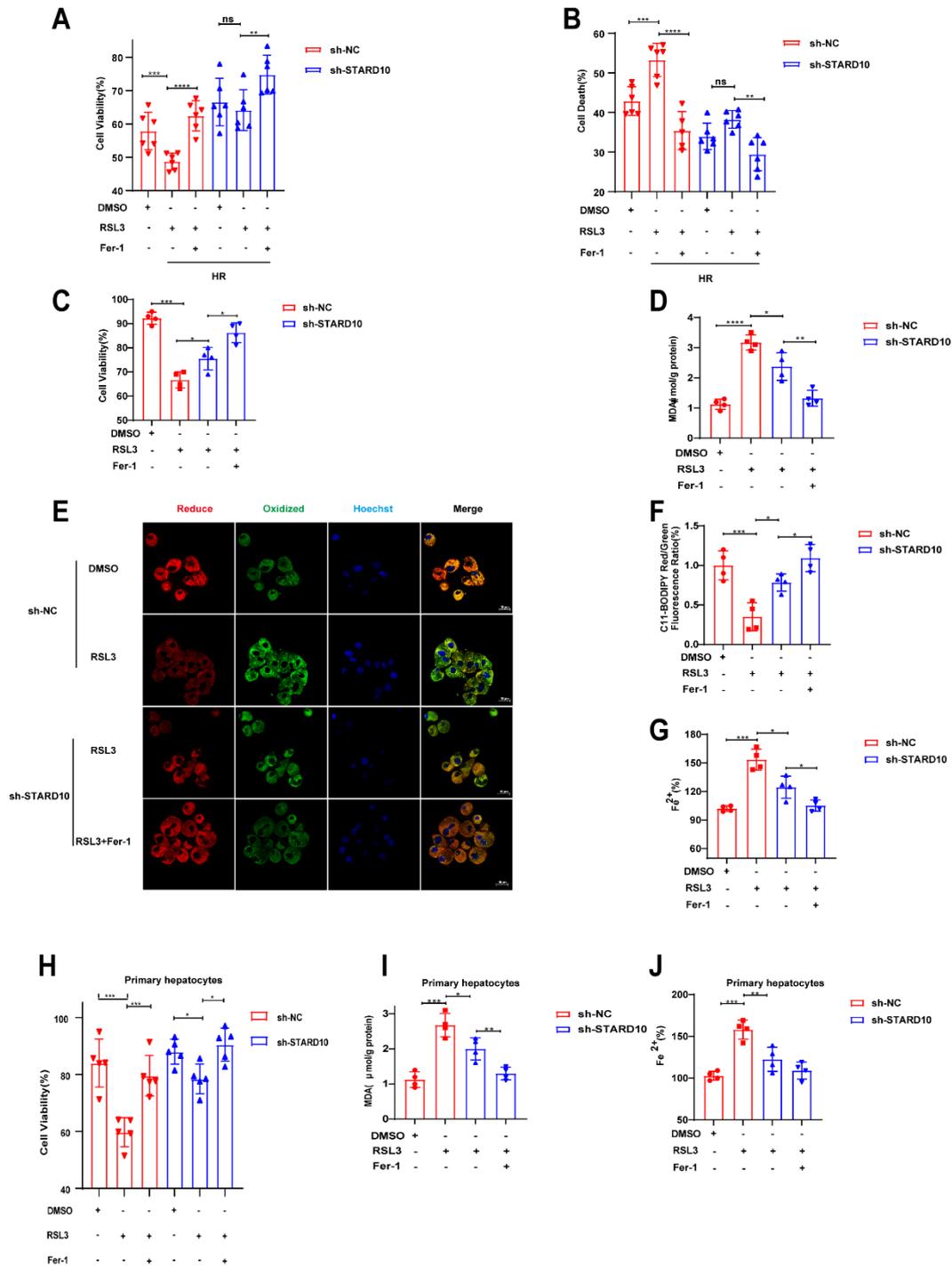
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80 **Fig. S4. Knockdown of STARD10 attenuates ferroptosis in steatotic hepatocytes.** Steatotic
 81 AML12 were subjected to hypoxia followed by treatment with 5 μM RSL3 and 4 μM Fer-1 during
 82 the reoxygenation phase for 6 h. Subsequently, (A) cell viability and (B) cell death were assessed.
 83 Additionally, steatotic AML12 cells were directly treated with the same concentrations of drugs for

84 24 h, after which (C) cell viability and (D) MDA levels were measured. (E-F) C11-BODIPY staining
85 was observed under a fluorescent microscope, and (G) Fe²⁺ accumulation was detected. Primary
86 hepatocytes were isolated from mice subjected to AAV8 intervention. After inducing steatosis with
87 300 μM PA for 24 h, cells were treated with 1 μM RSL3 and 4 μM Fer-1 for 12 h, followed by
88 measurements of cell viability, MDA, and Fe²⁺ levels. Data are expressed as mean ± SEM with
89 n≥3. *P < 0.05, **P < 0.01, ***P < 0.001, ****P < 0.0001. RSL3, Ras-selective lethal small molecule
90 3; Fer-1, Ferrostatin-1; DMSO, dimethyl sulfoxide; MDA, malondialdehyde.

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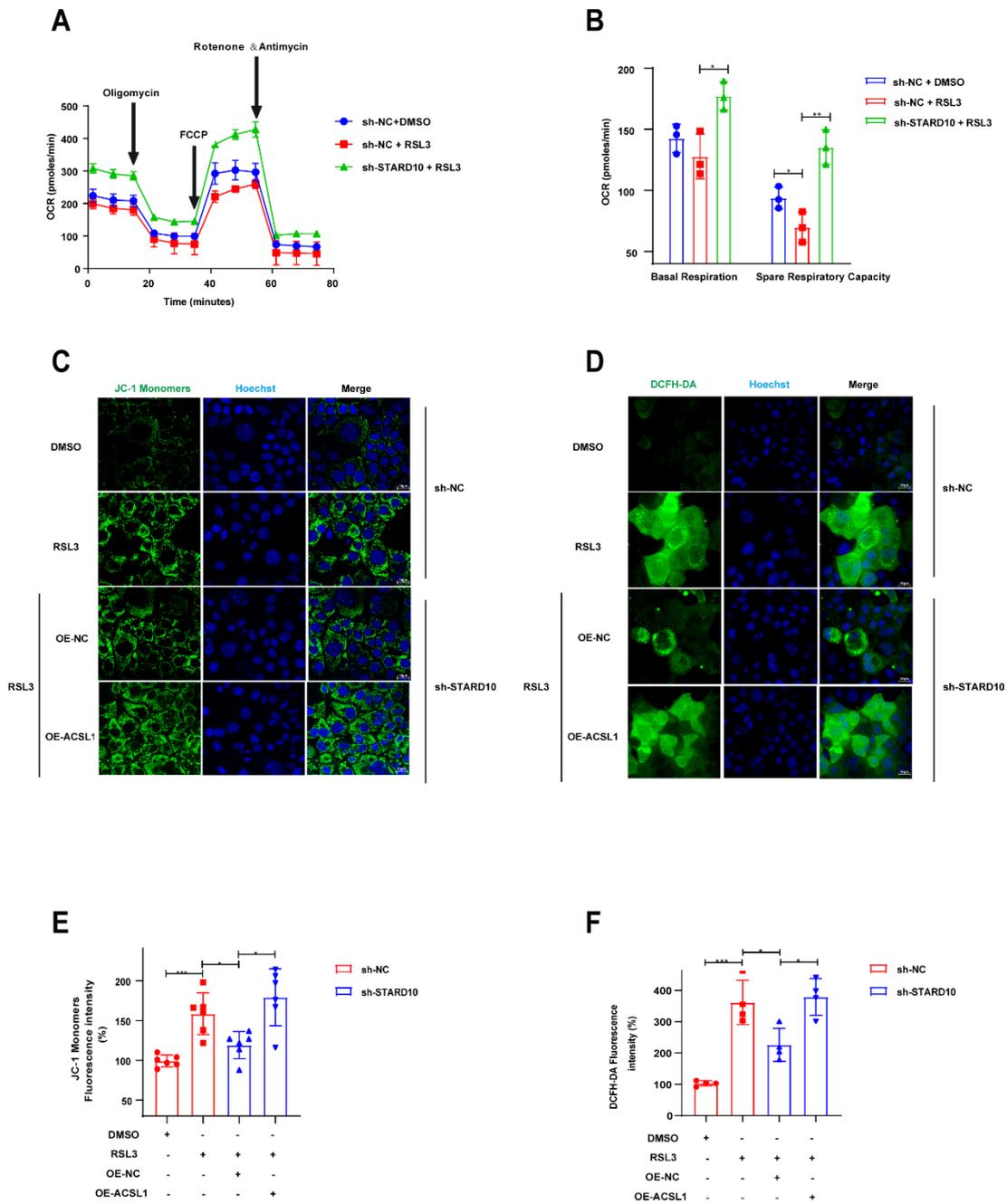
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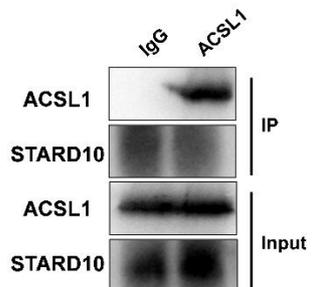
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114 **Fig. S5. STARD10 induces mitochondrial dysfunction via ACSL1.** In primary hepatocytes with
 115 STARD10 knockdown, after treatment with 1µM RSL3 for 12 h, measurements were taken of: (A)
 116 mitochondrial OCR, (B) baseline respiration and spare respiratory capacity. In steatotic AML12
 117 with 5 µM RSL3 and 4 µM Fer-1 treating for 24h were then stained with JC-1 and DCFH-DA to
 118 determine: (C, E) changes in mitochondrial membrane potential, and (D, F) production of ROS. All
 119 values are presented as mean ± SEM with n≥3. *P < 0.05, **P < 0.01, ***P < 0.001. OCR, oxygen

120 consumption rate; ROS, reactive oxygen species.

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122 **Fig. S6.**



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124 **Fig. S6. Co-IP of STARD10 with ACSL1 in primary hepatocytes.**

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