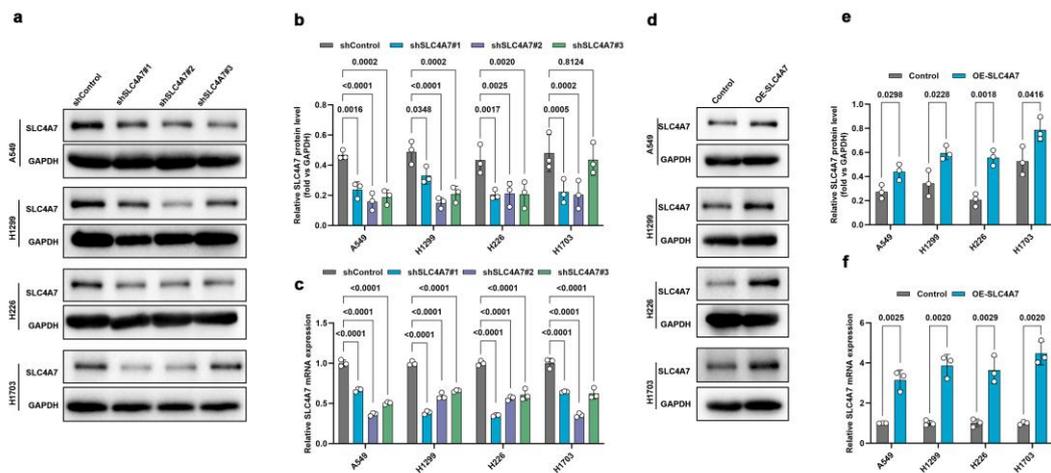


**Figure S1. SLC4A7 expression is elevated in NSCLC**

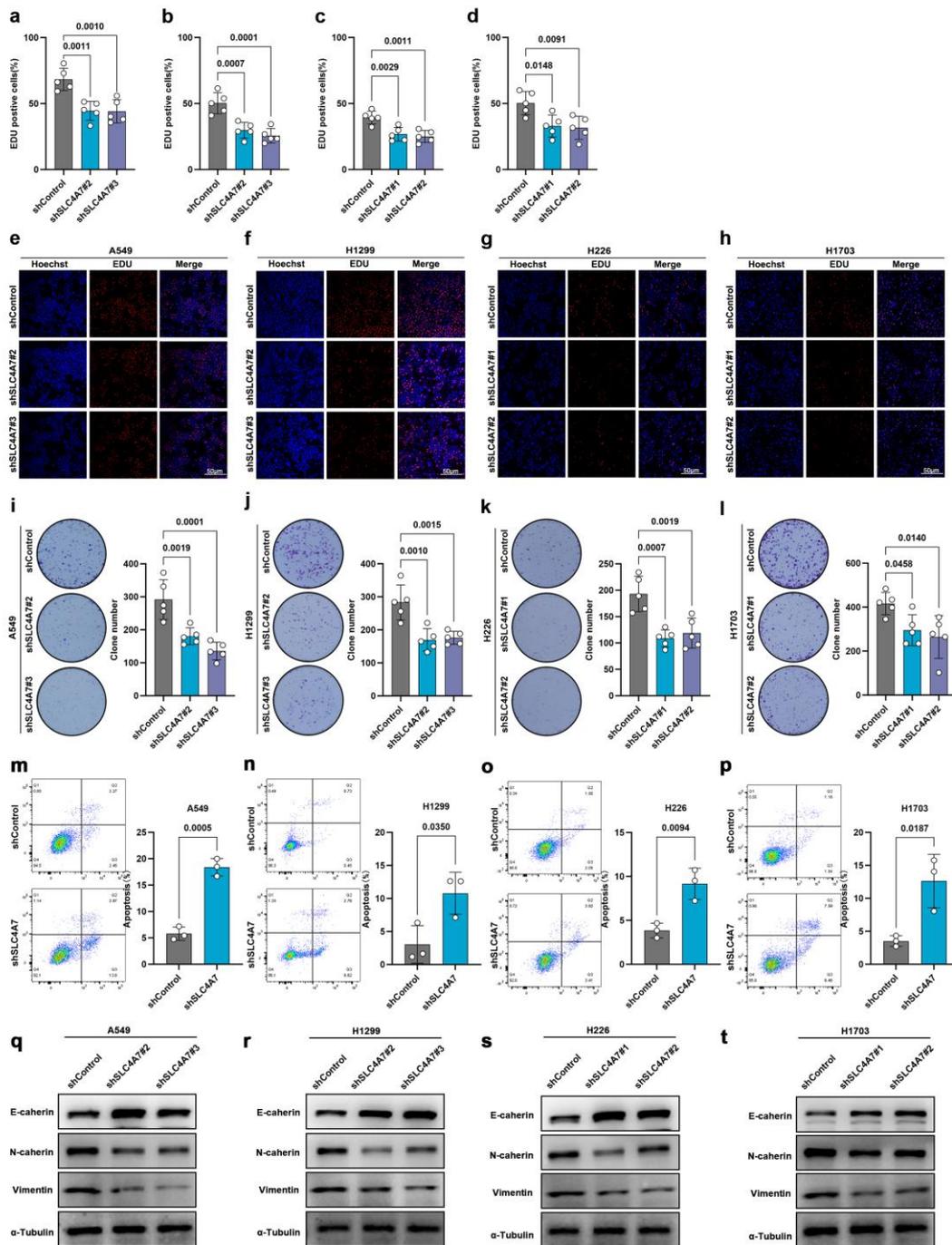
**a-c**, Expression of SLC4A4 (**a**), SLC4A5 (**b**) and SLC4A8 (**c**) in tumor and normal tissues in TCGA and CPTAC databases. **d**, Representative images of western blot analysis of SLC4A7 expression in BEAS-2B, H1975, PC-9, A549, H1299, H226, H1703 and SK-MES-1 cells **e**, Expression of SLC4A7 in various tumors in the TCGA database. **d**, Representative images of immunohistochemical staining of SLC4A7 in NSCLC, bar =

1000 $\mu\text{m}$ . *P* value was assessed two-tailed Student's *t*-test.



**Figure S2. Construction of stable cell lines with overexpression and knockdown of SLC4A7**

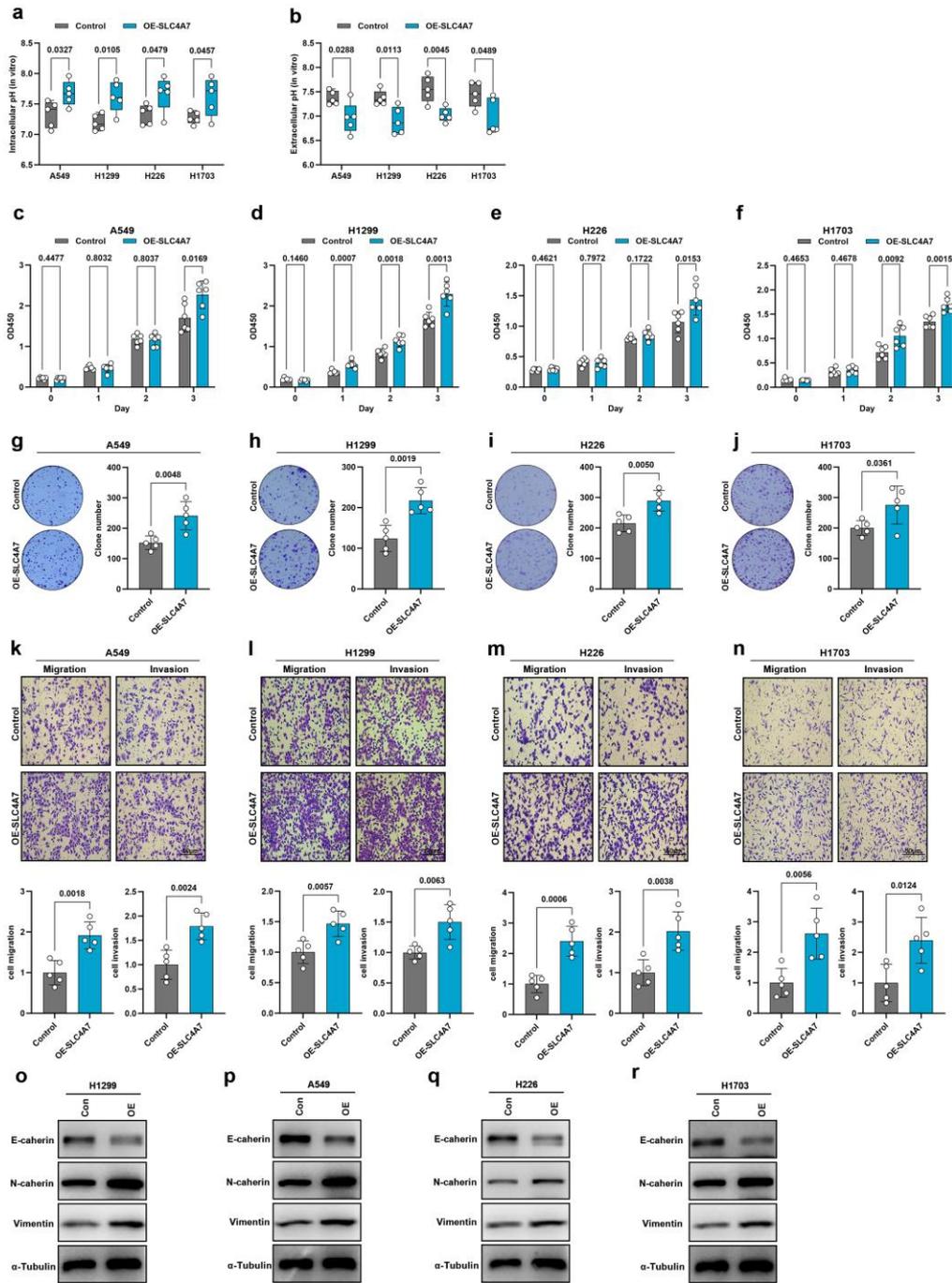
**a-c**, Western blot (**a**, **b**) and qPCR (**c**) analysis of SLC4A7 expression in A549, H1299, H226 and H1703 cells with or without SLC4A7 knockdown ( $n = 3$ ). **d-f**, Western blot (**d**, **e**) and qPCR (**f**) analysis of SLC4A7 expression in A549, H1299, H226 and H1703 cells with or without SLC4A7 overexpression ( $n=3$ ).  $P$  value was assessed by two-way ANOVA with Tukey's multiple comparison test (**a-c**) and two-tailed Student's  $t$ -test (**d-f**).



**Figure S3. SLC4A7 knockdown inhibits proliferation of non-small cell lung cancer cells**

**a-d**, EDU assay for detecting the proliferation of A549, H1299, H226 and H1703 cell with or without SLC4A7 knockdown (n = 5). **e-h**, Representative images of EdU staining of A549, H1299, H226 and H1703 cells after SLC4A7 knockdown (n=5), bar = 50µm. **i-l**, Representative images and statistical charts of clone formation of A549, H1299, H226 and H1703 cells after SLC4A7 knockdown (n=5). **m-p**, Flow cytometry for detecting the

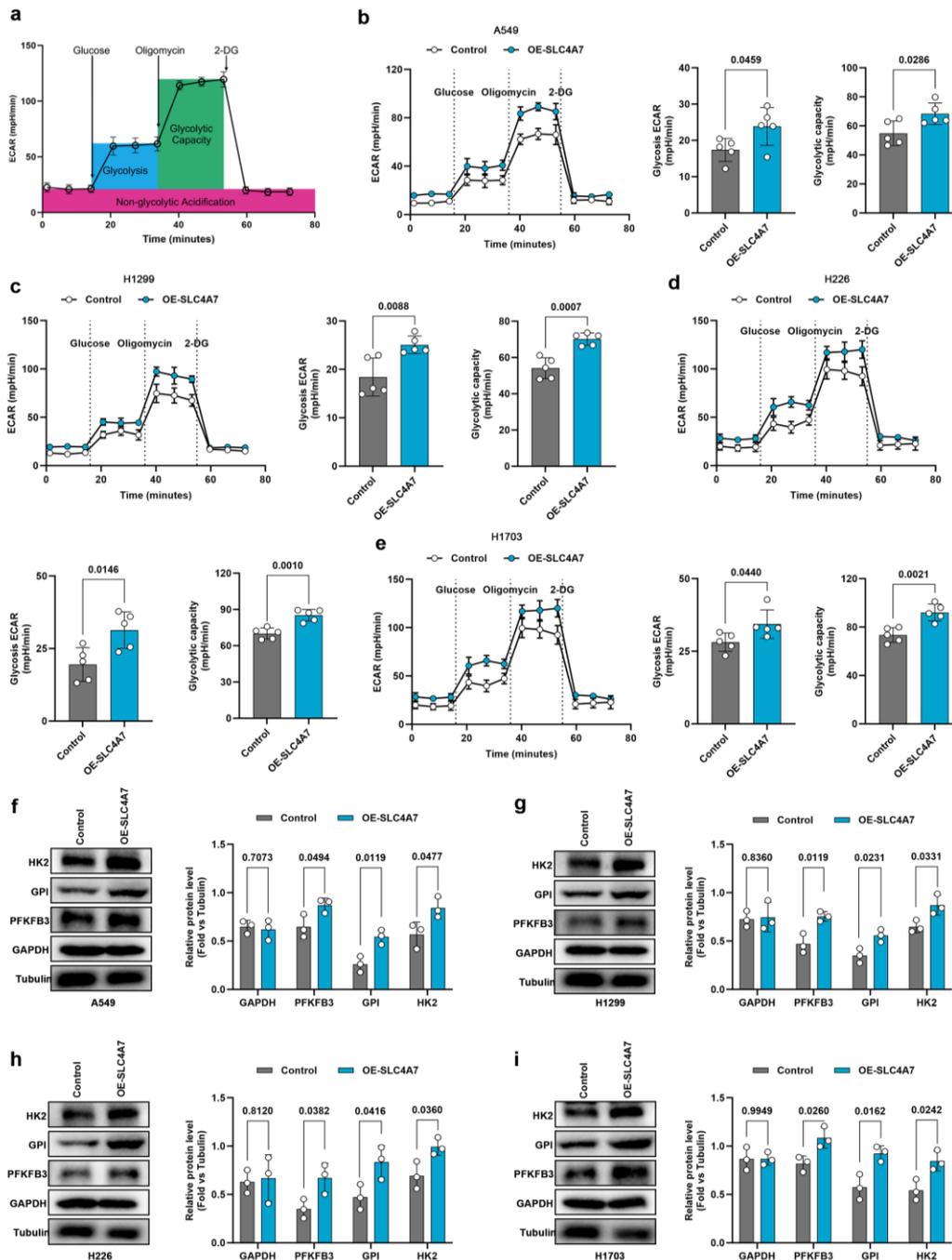
apoptosis rate of A549, H1299, H226 and H1703 cell with or without SLC4A7 knockdown (n = 3). **q-t**, Representative images of Western blot analysis of E-cadherin, N-cadherin, and vimentin expression in A549, H1299, H226, and H1703 cells with or without SLC4A7 knockdown. *P* value was assessed by one-way ANOVA followed by the Tukey's post hoc.



**Figure S4. SLC4A7 overexpression promotes proliferation and metastasis of NSCLC cells**

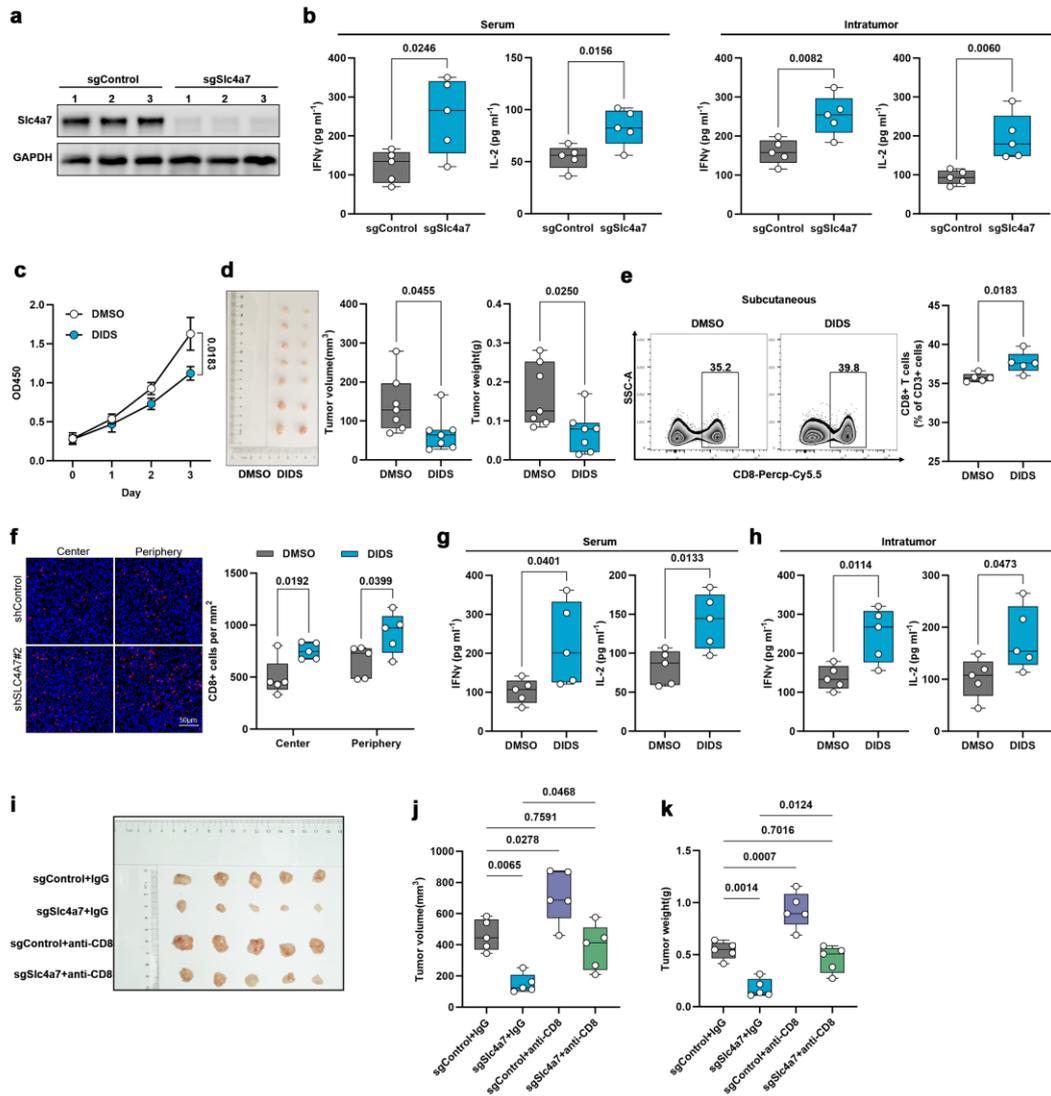
**a**, Quantitative analysis of intracellular pH using pHrodo Red intracellular pH indicator dye in A549, H1299, H226 and H1703 cells with or without SLC4A7 overexpression (n=5). **b**, Quantitative analysis of extracellular pH values using single-barreled H<sup>+</sup>-sensitive microelectrodes in A549, H1299, H226 and H1703 cells with or without

SLC4A7 overexpression (n=5). **c-f**, CCK8 assay was used to detect the proliferation ability of A549, H1299, H226 and H1703 cells after SLC4A7 overexpression (n = 6). **g-j**, Representative images and statistical charts of clone formation of A549, H1299, H226 and H1703 cells after SLC4A7 overexpression (n=5). **k-n**, Representative images and statistical charts of traswell assay of A549, H1299, H226 and H1703 cells after SLC4A7 overexpression (n=5), bar = 50 $\mu$ m. **o-r**, Western blot analysis of E-cadherin, N-cadherin and Vimentin expression levels with or without SLC4A7 overexpression (n = 3). *P* value was assessed by two-tailed Student's t-test.



**Figure S5. SLC4A7 overexpression promotes glycolysis in NSCLC cells**

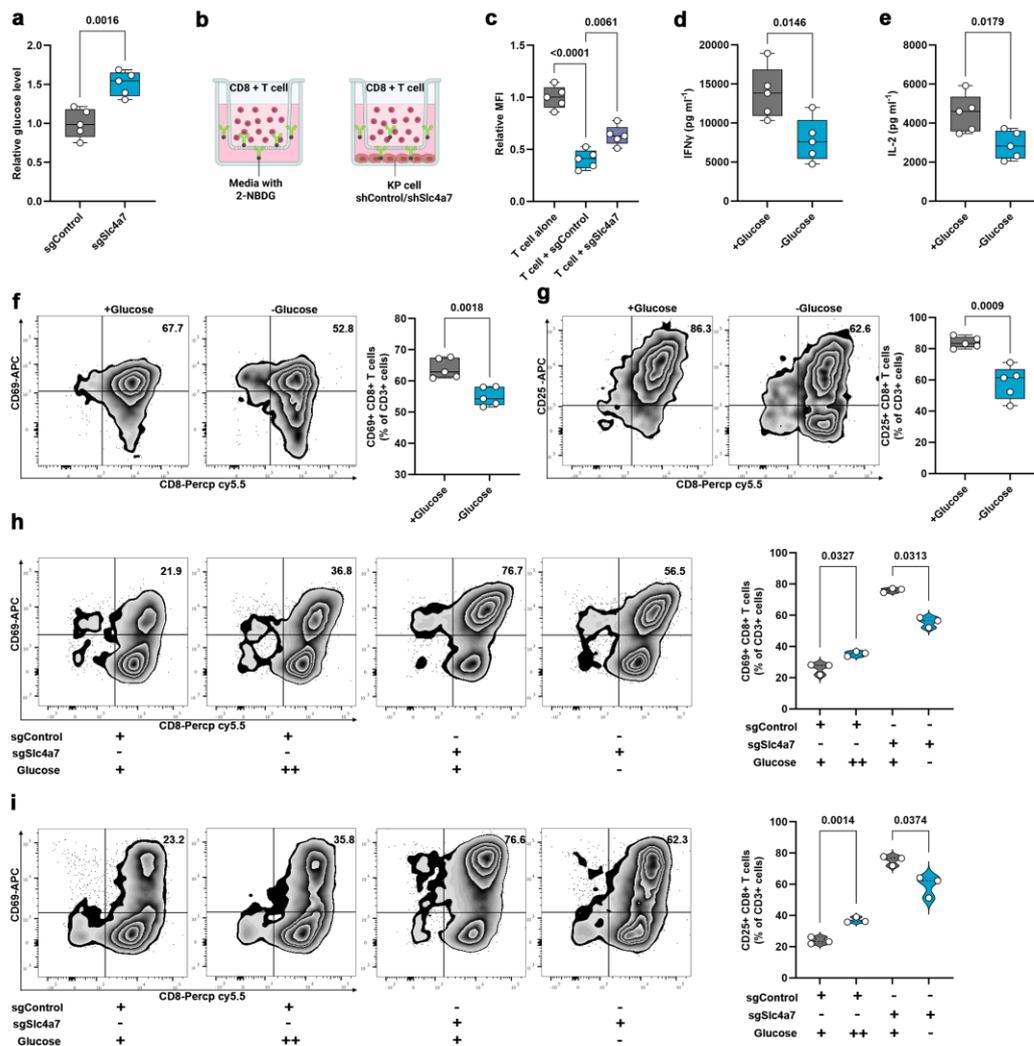
**a**, Schematic diagram of the seahorse experiment. **b-e**, Seahorse experimental results showed that SLC4A7 overexpression increased the level of glycolysis and the maximum glycolytic capacity of A549, H1299, H226 and H1703 cells ( $n = 5$ ). **f-i**, Western blot analysis of the expression levels of key glycolytic proteins HK2, PFKFB3, GPI, and GAPDH in A549, H1299, H226, and H1703 cells with or without SLC4A7 overexpression ( $n = 3$ ).  $P$  value was assessed two-tailed Student's  $t$ -test.



**Figure S6. The anti-tumor effect of SLC4A7 knockdown is related to CD8+ T cells**

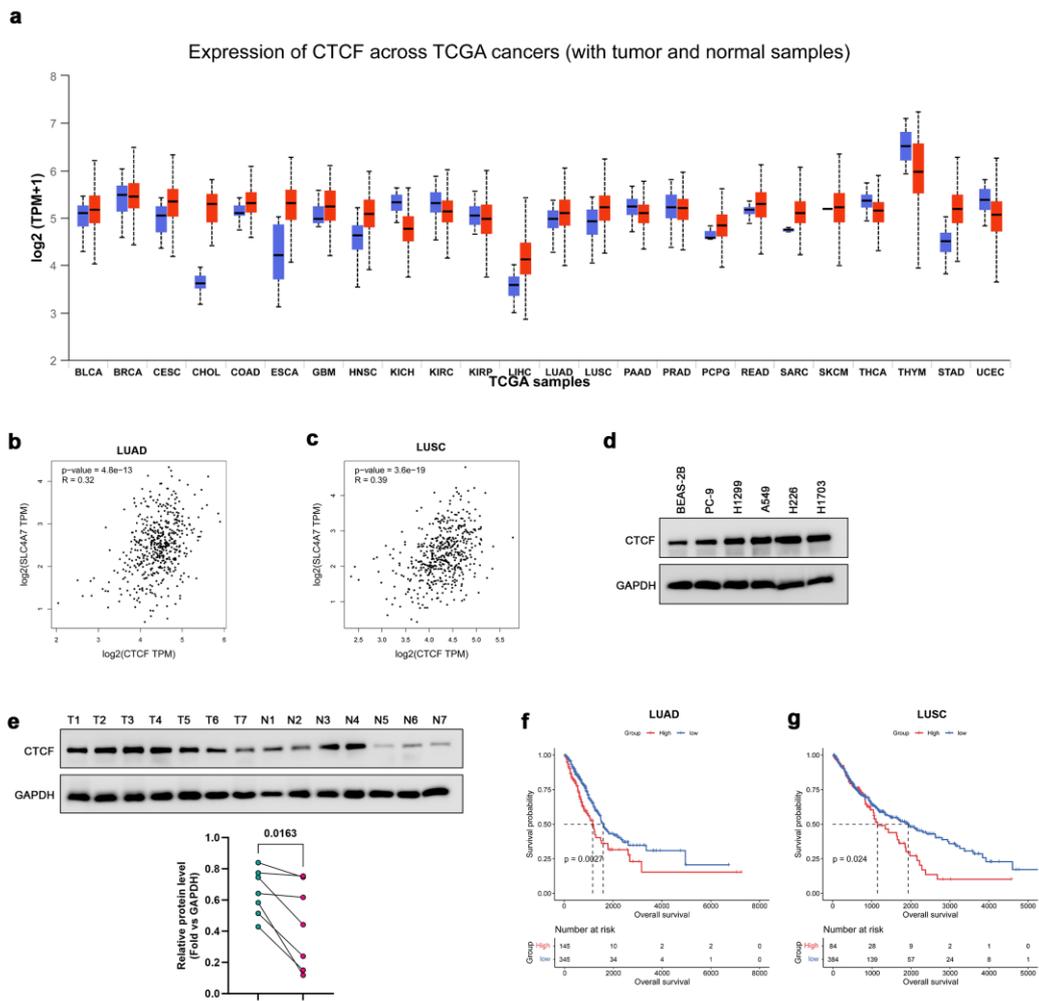
**a**, Representative image of western blot analysis of Slc4a7 knock out in KP cells. **b**, ELISA was used to detect the concentrations of IFN $\gamma$  and IL2 in the serum and tumors of mice with subcutaneous tumors (n = 5). **c**, CCK8 assay was used to detect the proliferation ability of KP cells after DIDS treatment (n = 3). **d**, Volume and weight of subcutaneous tumors in C57 mice after DIDS or DMSO treatment (n = 7). **e**, Flow cytometric analysis of the percentage of CD3+CD8+ T cells in subcutaneous tumors spleen in C57 mice after DIDS or DMSO treatment (n = 5). **f**, Representative images of infiltrating CD8+ T cells in KP subcutaneous tumors after DIDS or DMSO treatment (n = 5), bar = 50 $\mu$ m. **g, h**, ELISA was used to detect the concentrations of IFN $\gamma$  and IL2 in

the serum and tumors of mice with subcutaneous tumors (n = 5). **i-k**, Tumor volume and weight of subcutaneous KP tumors in mice treated with anti-CD8 or IgG (n = 5). *P* value was assessed by two-tailed Student's t-test (**b-h**), one-way ANOVA followed by the Tukey's post hoc (**j, k**).



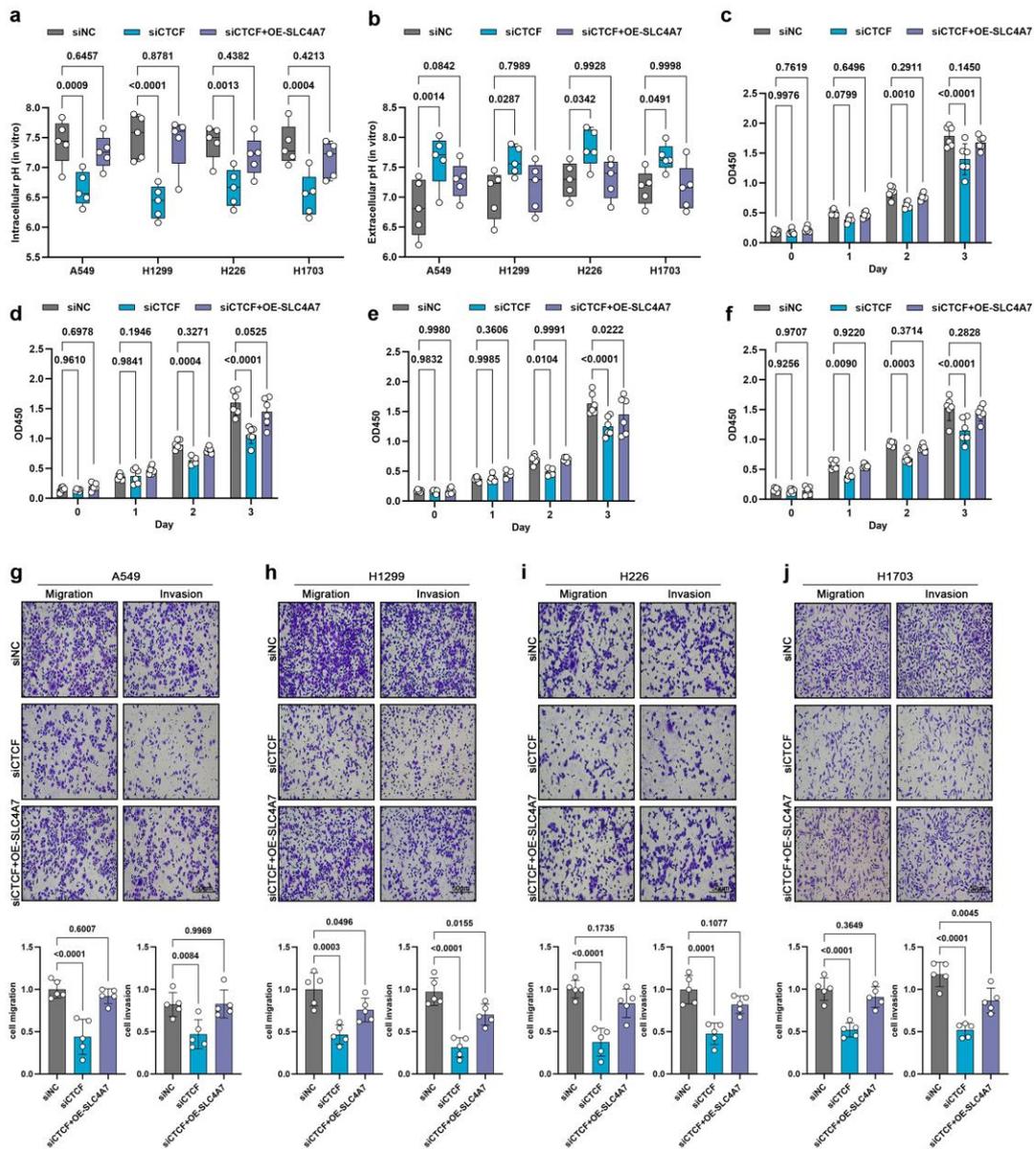
**Figure S7. Glucose is essential for T cell activation**

**a**, Glucose concentration in the supernatant of KP cells with or without Slc4a7 knockdown was analyzed using a glucose detection kit ( $n = 5$ ). **b,c**, Glucose uptake in CD8+ T cells cultured with or without KP tumor cells was measured using fluorescent 2-NBDG using a fluorescence microplate reader ( $n = 5$ ). **d,e**, ELISA analysis of IFN $\gamma$  and IL2 secretion levels after 48 h of culture at medium with or without glucose ( $n = 5$ ). **f,g**, Flow cytometry analysis of the proportion of CD69+ and CD25+ CD8+ T cells after 48 h of culture in the presence or absence of T glucose ( $n = 5$ ). **h,i**, Flow cytometry analysis of the effects of adding or removing glucose in KP-conditioned medium on CD8+ T cell activation after 48 hours of culture ( $n = 3$ ). *P* value was assessed by two-tailed Student's *t*-test (**a, c-g**), one-way ANOVA followed by the Tukey's post hoc (**h, i**).



**Figure S8. CTCF expression is elevated in lung cancer and is associated with clinical prognosis**

**a**, Expression of CTCF in various tumors in the TCGA database. **b,c**, Correlation analysis between CTCF and SLC4A7 in TCGA database. **d**, Western blot analysis of SLC4A7 expression in BEAS-2B, pc-9, h1299, a549, h226, h1703 cells. **e**, Western blot analysis of SLC4A7 expression in tumor and normal tissue (n = 7) **f, g**, TCGA database shows that high expression of CTCF shortens survival time



**Figure S9. CTCF knockdown leads to intracellular acidification and inhibits proliferation and metastasis of NSCLC cells**

**a**, Quantitative analysis of intracellular pH using pHrodo Red intracellular pH indicator dye in A549, H1299, H226 and H1703 cells with or without CTCF knockdown (n=5). **b**, Quantitative analysis of extracellular pH values using single-barreled H<sup>+</sup>-sensitive microelectrodes in A549, H1299, H226 and H1703 cells with or without CTCF knockdown (n=5). **c-f**, CCK8 assay was used to detect the proliferation ability of A549, H1299, H226 and H1703 cells after CTCF knockdown. (n = 6). **g-j**, Representative images and statistical charts of transwell assay of A549, H1299, H226 and H1703 cells

after CTCF knockdown (n=5), bar = 50µm. *P* value was assessed by one-way ANOVA followed by the Tukey's post hoc.

**Table S1. Materials and reagents used in this article**

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Antibodies		
SLC4A7 Polyclonal antibody	Proteintech	Cat# 29442-1-AP; RRID: AB_3086133
SLC4A7 Polyclonal Antibody	Invitrogen	Cat# PA5-57433; RRID: AB_2647530
SLC4A7 Antibody	Affinity	Cat# DF9927; RRID: AB_2843121
CTCF (D31H2) XP Rabbit mAb	Cell Signaling Technology	Cat# 3418; RRID: AB_2086791
Monoclonal Anti-CTCF antibody produced in mouse	Sigma Aldrich	Cat# AMAB90663; AB_2665624
GPI Monoclonal antibody	Proteintech	Cat# 67178-1-Ig; RRID: AB_2882474
Hexokinase 2 Monoclonal antibody	Proteintech	Cat# 66974-1-Ig; RRID: AB_2882294
PFKFB3 Polyclonal antibody	Proteintech	Cat# 13763-1-AP; RRID: AB_2162854
Alpha Tubulin Monoclonal antibody	Proteintech	Cat# 66031-1-Ig; RRID: AB_11042766
GAPDH Monoclonal antibody	Proteintech	Cat# 60004-1-Ig; RRID: AB_2107436
E-cadherin Monoclonal antibody	Proteintech	Cat# 60335-1-Ig; RRID: AB_2881444
N-cadherin Monoclonal antibody	Proteintech	Cat# 66219-1-Ig; RRID: AB_2881610
Vimentin Monoclonal antibody	Proteintech	Cat# 60330-1-Ig; RRID: AB_2881439
Fixable Viability Stain 510	BD Biosciences	Cat# 564406; RRID: AB_2869572

Fixable Viability Stain 450	BD Biosciences	Cat#562247; RRID: AB_2869405
APC-Cy7 Rat Anti-Mouse CD45	BD Biosciences	Cat# 557659; RRID: AB_396774
APC Hamster Anti-Mouse CD3e	BD Biosciences	Cat# 553066; RRID: AB_398529
FITC anti-mouse CD3ε Antibody	Biolegend	Cat# 100306; RRID: AB_312671
R718 Rat Anti-Mouse CD8a	BD Biosciences	Cat# 566985; RRID: AB_2869989
PerCP-Cy5.5 Rat Anti-Mouse CD8a	BD Biosciences	Cat# 551162; RRID: AB_394081
FITC anti-mouse CD4 Antibody	Biolegend	Cat# 100406; RRID: AB_312691
FOXP3 Monoclonal Antibody (FJK-16s), PE	Thermo Fisher Scientific	Cat# 12-5773-82; RRID: AB_465936
F4/80 Monoclonal Antibody (BM8), PE-eFluor™ 610	Thermo Fisher Scientific	Cat# 61-4801-82; RRID:AB_2574612
FITC anti-mouse/human CD11b Antibody	Biolegend	Cat# 101206; RRID: AB_312789
PE/Cyanine7 anti-mouse CD86 Antibody	Biolegend	Cat# 105115; RRID: AB_493601
Alexa Fluor 647 Rat Anti-Mouse CD206	BD Biosciences	Cat# 565250; RRID: AB_2739133
BV421 Hamster Anti-Mouse CD11c	BD Biosciences	Cat# 562782; RRID: AB_2737789
MHC Class II (I-A/I-E) Monoclonal Antibody (M5/114.15.2), PE-Cyanine7	Thermo Fisher Scientific	Cat# 25-5321-82; RRID: AB_10870792
FITC anti-mouse CD19 Antibody	Biolegend	Cat# 115505; RRID: AB_313640
PE/Cyanine7 anti-mouse NK-1.1 Antibody	Biolegend	Cat# 108714; RRID: AB_389364
BV421 Hamster Anti-Mouse CD69	BD Biosciences	Cat# 562920; RRID: AB_2687478
CD69 Monoclonal Antibody (H1.2F3), APC	Thermo Fisher Scientific	Cat# 17-0691-80; RRID:AB_1210795
CD25 Monoclonal Antibody (PC61.5), APC	Thermo Fisher Scientific	Cat# 17-0251-81; RRID: AB_469365

CD25 Monoclonal Antibody (PC61.5), FITC	Thermo Fisher Scientific	Cat# 11-0251-80; RRID:AB_2802176
Anti-pan Cytokeratin antibody	Abcam	Cat# ab7753; RRID: AB_306047
Anti-CD8 alpha antibody	Abcam	Cat# ab237709; RRID: AB_2892677
InVivoMAb anti-mouse CD8 $\alpha$	BioCell	Cat# BE0004-1; RRID: AB_1107671
InVivoMAb anti-mouse PD-1 (CD279)	BioCell	Cat# BE0146; RRID: AB_10949053
InVivoMAb anti-mouse PD-L1 (B7-H1)	BioCell	Cat# BE0101; RRID: AB_10949073
Chemicals, peptides, and recombinant proteins		
Seahorse XF 1.0 M glucose solution	Aligen	Cat#103577-100
Seahorse XF 100 mM pyruvate solution	Aligent	Cat#103578-100
Seahorse XF 200 mM glutamine solution	Aligent	Cat#103579-100
3-aminopropyl phosphonate	Sigma-Aldrich	Cat#268615
HEPES (1M)	Thermo Fisher Scientific	Cat#15630080
MEM NEAA	Thermo Fisher Scientific	Cat#11140050
SODIUM PYRUVATE	Thermo Fisher Scientific	Cat#11360070
GLUTAMAX I, 100X	Thermo Fisher Scientific	Cat#35050061
Sodium bicarbonate ( $^{13}\text{C}$ , 99%)	Cambridge Isotope Laboratories	Cat#87081-58-1
Advanced DMEM/F12	Thermo Fisher Scientific	Cat# 12634010
Glutamax	Gibco	Cat# 35050

N2 SUPPLEMENT	Thermo Fisher Scientific	Cat#A1370701
B-27 SUPPLEMENT	Thermo Fisher Scientific	Cat#12587010
EGF	PeproTech	Cat#AF-100-15-500UG
Noggin	MedChemExpre ss	Cat#HY-P700143AF
R-spondin-1	Thermo Fisher Scientific	Cat#120-38-100UG
FGF10	MedChemExpre ss	Cat#HY-P7342AF
N-Acetyl-L-cysteine	MedChemExpre ss	Cat#HY-B0215
Y-27632	MedChemExpre ss	Cat#HY-10071
A83-01	MedChemExpre ss	Cat#HY-10432
Matrigel	Corning	Cat#354234
TrypLE Express	Thermo Fisher Scientific	Cat#12604021
Critical commercial assays		
Cell-Light EdU Apollo567 In Vitro Kit	Ribobio	Cat#C10310-1
Seahorse XF Cell Mito Stress Test Kit	Aligent	Cat#103015-100
Seahorse XF Glycolysis Stress Test Kit	Aligent	Cat#103020-100
Dynabeads Mouse T-Activator CD3/CD28 for activation of mouse T cells	Thermo Fisher Scientific	Cat#11456D
Mouse CD8+ T-Cell Isolation Kit	Selleck	Cat#B90011

Magnetic CHIP Kit	Sigma-Aldrich	Cat#17-10086
Intracellular pH Calibration Buffer Kit	Invitrogen	Cat#P35379
Deposited data		
Single cell transcriptomic	CZ CELLxGENE	<a href="https://cellxgene.cziscience.com/">https://cellxgene.cziscience.com/</a>
RNA sequencing	This paper	
ATAC sequencing	This paper	
TCGA data RNA-seq	GDC	<a href="https://portal.gdc.cancer.gov/">https://portal.gdc.cancer.gov/</a>
Experimental models: Cell lines		
NCI-H1299	Procell	Cat#CL-0165
A549	Procell	Cat#CL-0016
NCI-H226	Procell	Cat#CL-0396
NCI-H1703	Procell	Cat#CL-0390
293T	Procell	Cat#CL-0005
KP cells	This paper	NA
Experimental models: Organisms/strains		
BALB/c-Nude	GemPharmatec h	Strain NO. D000521
C57BL/6J	Charles River	NA
NOD/ShiLtJGpt- Prkdc <sup>em26Cd52</sup>   2rg <sup>em26Cd22</sup> /Gpt	GemPharmatec h	Strain NO. T001475
Oligonucleotides		
shSLC4A7#1 target sequence GCAATGAAACTCTAGCACAAT	This paper	NA
shSLC4A7#2 target sequence CCAGTATTTGACCGTATAAA	This paper	NA

shSLC4A7#3 target sequence CCATGAAATTGGACGATCAAT	This paper	NA
sgSlc4a7#1 target sequence GTCGGCGTCACAAGCATCGA	This paper	NA
sgSlc4a7#2 target sequence GAGGGCACAACATCACCAC	This paper	NA
sgSlc4a7#3 target sequence GCTAAGACACGCCATCGCAG	This paper	NA
siCTCF#1 target sequence (5'-3') GUGGAGGAGUCCGAAACUUTT AAGUUCCGACUCCUCCACTT	This paper	NA
siCTCF#2 target sequence (5' -3' ) GGAAAGUGAACCCAUGAUATT UAUCAUGGGUUCACUUUCCTT	This paper	NA
siCTCF#3 target sequence (5' -3' ) GGGACACAUACAAGCUGAATT UUCAGCUUGUAUGUGUCCCTT	This paper	NA
Software and algorithms		
FlowJo	FlowJo LLC	<a href="https://flowjovx.software.informer.com/">https://flowjovx.software.informer.com/</a>
GraphPad Prism 10.0	GraphPad Prism	<a href="https://graphpad-prism.software.informer.com/">https://graphpad-prism.software.informer.com/</a>



**Table S2. Tissue microarray clinical information of all sample**

Variable	Overall, N = 172 <sup>1</sup>	High, N = 24 <sup>1</sup>	Low, N = 56 <sup>1</sup>	Middle, N = 92 <sup>1</sup>	p- value <sup>2</sup>
<b>Pathological type</b>					0.084
LUAD	82 (48%)	12 (50%)	33 (59%)	37 (40%)	
LUSC	90 (52%)	12 (50%)	23 (41%)	55 (60%)	
<b>Gender</b>					0.036
female	35 (20%)	3 (12%)	18 (32%)	14 (15%)	
male	137 (80%)	21 (88%)	38 (68%)	78 (85%)	
<b>Age</b>					0.8
<=65	114 (66%)	15 (62%)	36 (64%)	63 (68%)	
>65	58 (34%)	9 (38%)	20 (36%)	29 (32%)	
<b>Tumor stage</b>					<0.001
I	54 (31%)	7 (29%)	30 (54%)	17 (18%)	
II	56 (33%)	9 (38%)	17 (30%)	30 (33%)	
III	60 (35%)	8 (33%)	9 (16%)	43 (47%)	
IV	2 (1.2%)	0 (0%)	0 (0%)	2 (2.2%)	
<b>Distant metastasis</b>					>0.9
negative	168 (98%)	24 (100%)	55 (98%)	89 (97%)	
positive	4 (2.3%)	0 (0%)	1 (1.8%)	3 (3.3%)	
<b>Lymph node metastasis</b>					0.008
negative	92 (53%)	13 (54%)	39 (70%)	40 (43%)	
positive	80 (47%)	11 (46%)	17 (30%)	52 (57%)	
<b>EGFR</b>					0.3

**Table S2. Tissue microarray clinical information of all sample**

Variable	Overall, N = 172 <sup>1</sup>	High, N = 24 <sup>1</sup>	Low, N = 56 <sup>1</sup>	Middle, N = 92 <sup>1</sup>	p- value <sup>2</sup>
negative	52 (30%)	9 (38%)	21 (38%)	22 (24%)	
positive	10 (5.8%)	2 (8.3%)	2 (3.6%)	6 (6.5%)	
unknown	110 (64%)	13 (54%)	33 (59%)	64 (70%)	
<b>ALK</b>					0.2
negative	51 (30%)	8 (33%)	17 (30%)	26 (28%)	
positive	14 (8.1%)	4 (17%)	6 (11%)	4 (4.3%)	
unknown	107 (62%)	12 (50%)	33 (59%)	62 (67%)	
<b>Status</b>					<0.001
alive	76 (44%)	6 (25%)	37 (66%)	33 (36%)	
dead	96 (56%)	18 (75%)	19 (34%)	59 (64%)	

<sup>1</sup>n (%)<sup>2</sup>Pearson's Chi-squared test; Fisher's exact test

**Table S3. Tissue microarray clinical information of LUAD**

Variable	Overall, N = 82 <sup>1</sup>	High, N = 12 <sup>1</sup>	Low, N = 33 <sup>1</sup>	Middle, N = 37 <sup>1</sup>	p- value <sup>2</sup>
<b>Gender</b>					0.071
female	32 (39%)	3 (25%)	18 (55%)	11 (30%)	
male	50 (61%)	9 (75%)	15 (45%)	26 (70%)	
<b>Age</b>					0.8
<=65	57 (70%)	9 (75%)	24 (73%)	24 (65%)	
>65	25 (30%)	3 (25%)	9 (27%)	13 (35%)	
<b>Tumor stage</b>					0.084
I	32 (39%)	4 (33%)	18 (55%)	10 (27%)	
II	23 (28%)	5 (42%)	9 (27%)	9 (24%)	
III	26 (32%)	3 (25%)	6 (18%)	17 (46%)	
IV	1 (1.2%)	0 (0%)	0 (0%)	1 (2.7%)	
<b>Distant metastasis</b>					>0.9
negative	80 (98%)	12 (100%)	32 (97%)	36 (97%)	
positive	2 (2.4%)	0 (0%)	1 (3.0%)	1 (2.7%)	
<b>Lymph node metastasis</b>					0.041
negative	46 (56%)	6 (50%)	24 (73%)	16 (43%)	
positive	36 (44%)	6 (50%)	9 (27%)	21 (57%)	
<b>EGFR</b>					0.4
negative	52 (63%)	9 (75%)	21 (64%)	22 (59%)	
positive	10 (12%)	2 (17%)	2 (6.1%)	6 (16%)	
unknown	20 (24%)	1 (8.3%)	10 (30%)	9 (24%)	
<b>ALK</b>					0.086
negative	51 (62%)	8 (67%)	17 (52%)	26 (70%)	
positive	14 (17%)	4 (33%)	6 (18%)	4 (11%)	

**Table S3. Tissue microarray clinical information of LUAD**

Variable	Overall, N = 82 <sup>1</sup>	High, N = 12 <sup>1</sup>	Low, N = 33 <sup>1</sup>	Middle, N = 37 <sup>1</sup>	p- value <sup>2</sup>
unknown	17 (21%)	0 (0%)	10 (30%)	7 (19%)	
<b>Status</b>					0.036
alive	33 (40%)	3 (25%)	19 (58%)	11 (30%)	
dead	49 (60%)	9 (75%)	14 (42%)	26 (70%)	

<sup>1</sup>n (%)

<sup>2</sup>Fisher's exact test; Pearson's Chi-squared test

**Table S4. Tissue microarray clinical information of LUAD**

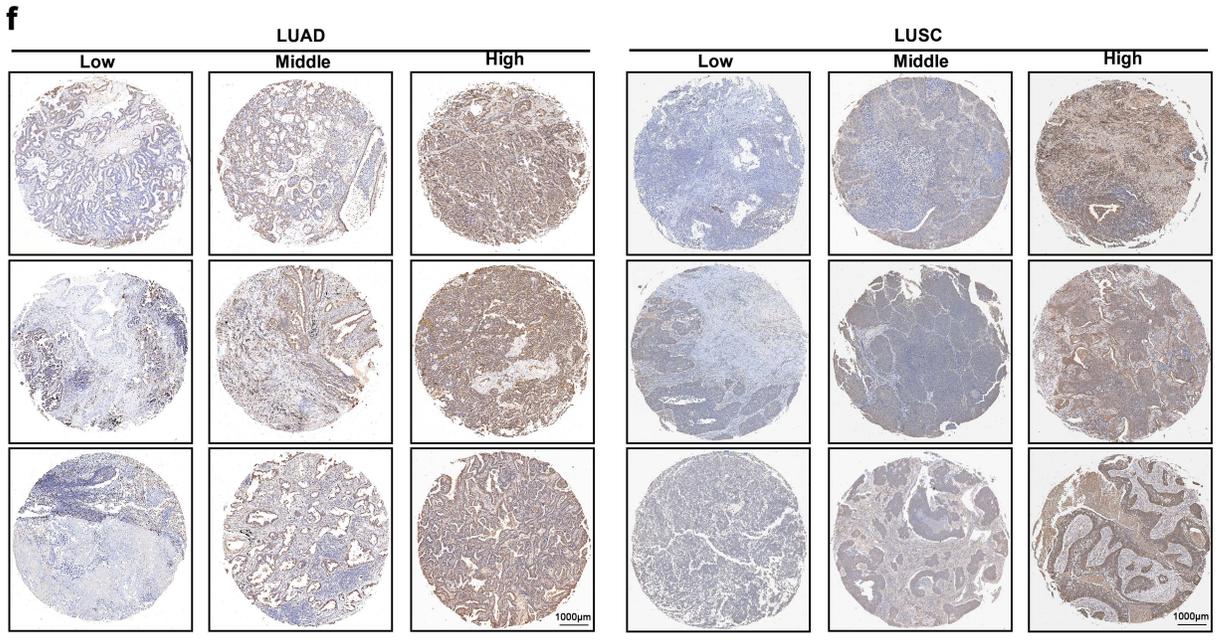
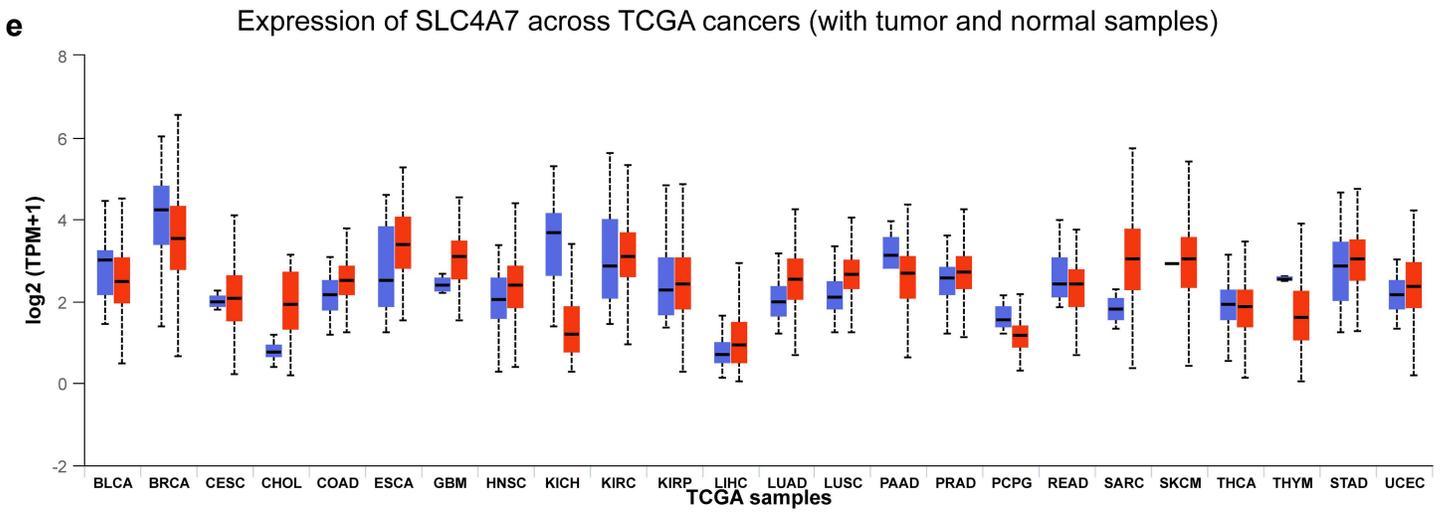
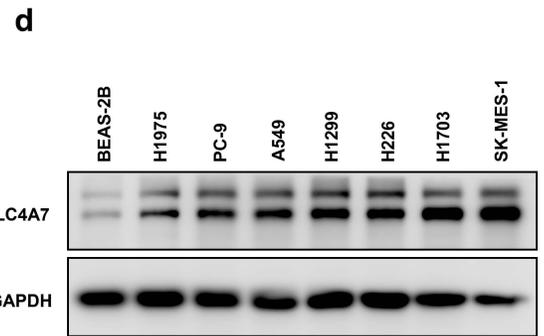
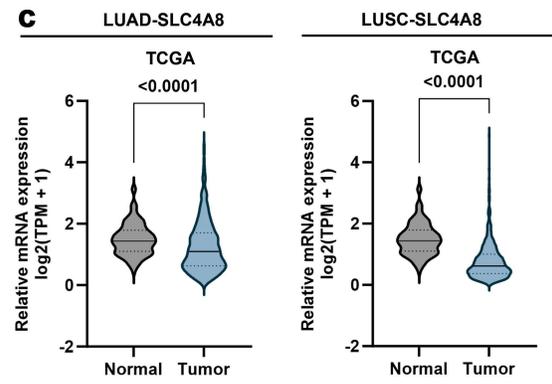
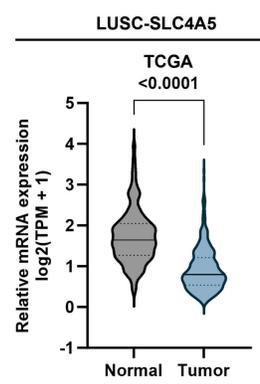
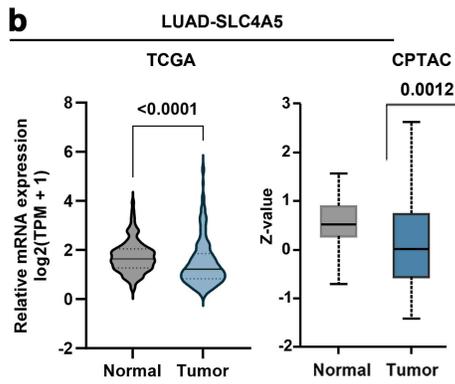
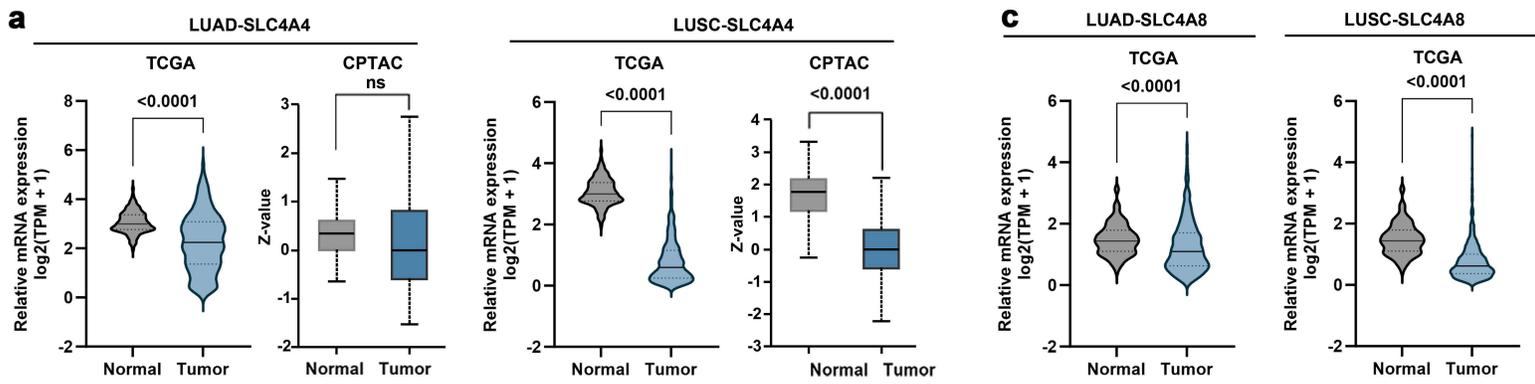
Variable	Overall, N = 90 <sup>1</sup>	High, N = 12 <sup>1</sup>	Low, N = 23 <sup>1</sup>	Middle, N = 55 <sup>1</sup>	p- value <sup>2</sup>
<b>Gender</b>					0.7
female	3 (3.3%)	0 (0%)	0 (0%)	3 (5.5%)	
male	87 (97%)	12 (100%)	23 (100%)	52 (95%)	
<b>Age</b>					0.14
<=65	57 (63%)	6 (50%)	12 (52%)	39 (71%)	
>65	33 (37%)	6 (50%)	11 (48%)	16 (29%)	
<b>Tumor stage</b>					0.006
I	22 (24%)	3 (25%)	12 (52%)	7 (13%)	
II	33 (37%)	4 (33%)	8 (35%)	21 (38%)	
III	34 (38%)	5 (42%)	3 (13%)	26 (47%)	
IV	1 (1.1%)	0 (0%)	0 (0%)	1 (1.8%)	
<b>Distant metastasis</b>					>0.9
negative	88 (98%)	12 (100%)	23 (100%)	53 (96%)	
positive	2 (2.2%)	0 (0%)	0 (0%)	2 (3.6%)	
<b>Lymph node metastasis</b>					0.2
negative	46 (51%)	7 (58%)	15 (65%)	24 (44%)	
positive	44 (49%)	5 (42%)	8 (35%)	31 (56%)	
<b>EGFR</b>					
unknown	90 (100%)	12 (100%)	23 (100%)	55 (100%)	
<b>ALK</b>					
unknown	90 (100%)	12 (100%)	23 (100%)	55 (100%)	
<b>Status</b>					0.002
alive	43 (48%)	3 (25%)	18 (78%)	22 (40%)	
dead	47 (52%)	9 (75%)	5 (22%)	33 (60%)	

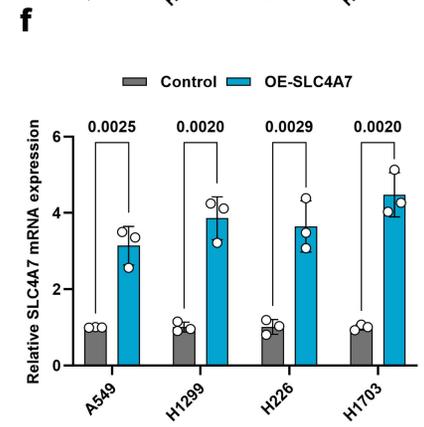
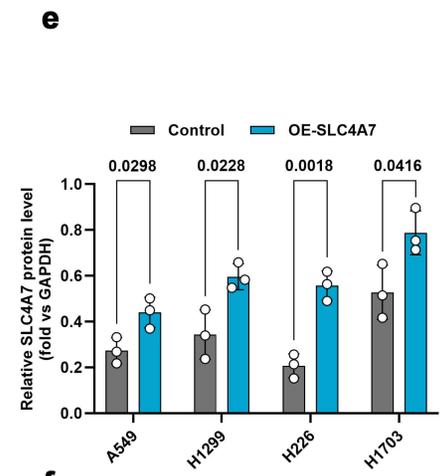
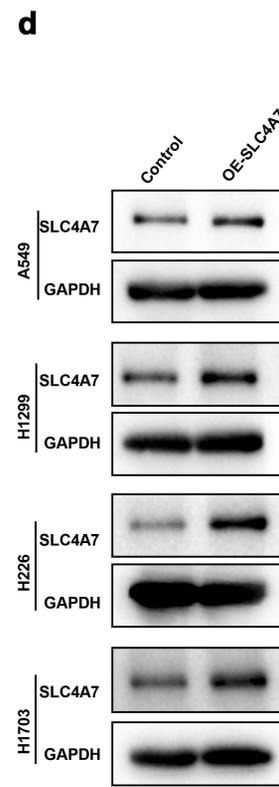
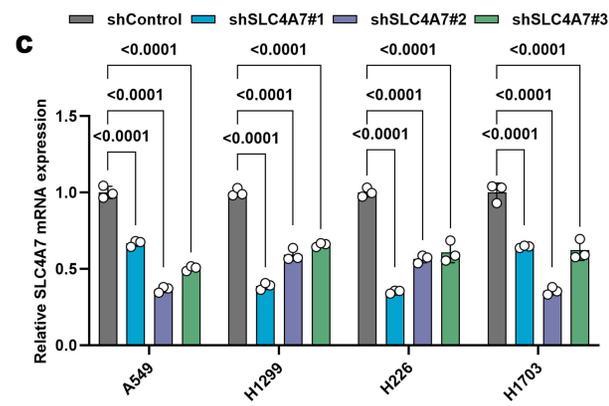
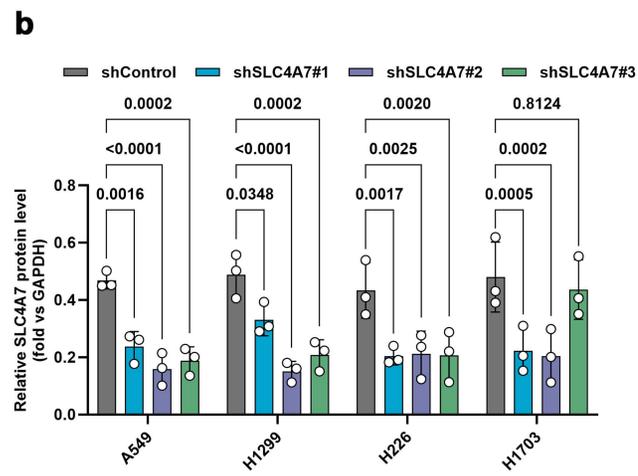
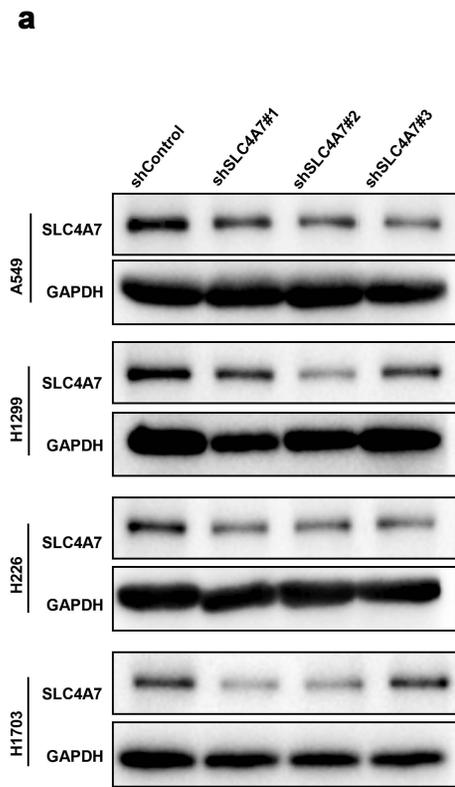
**Table S4. Tissue microarray clinical information of LUAD**

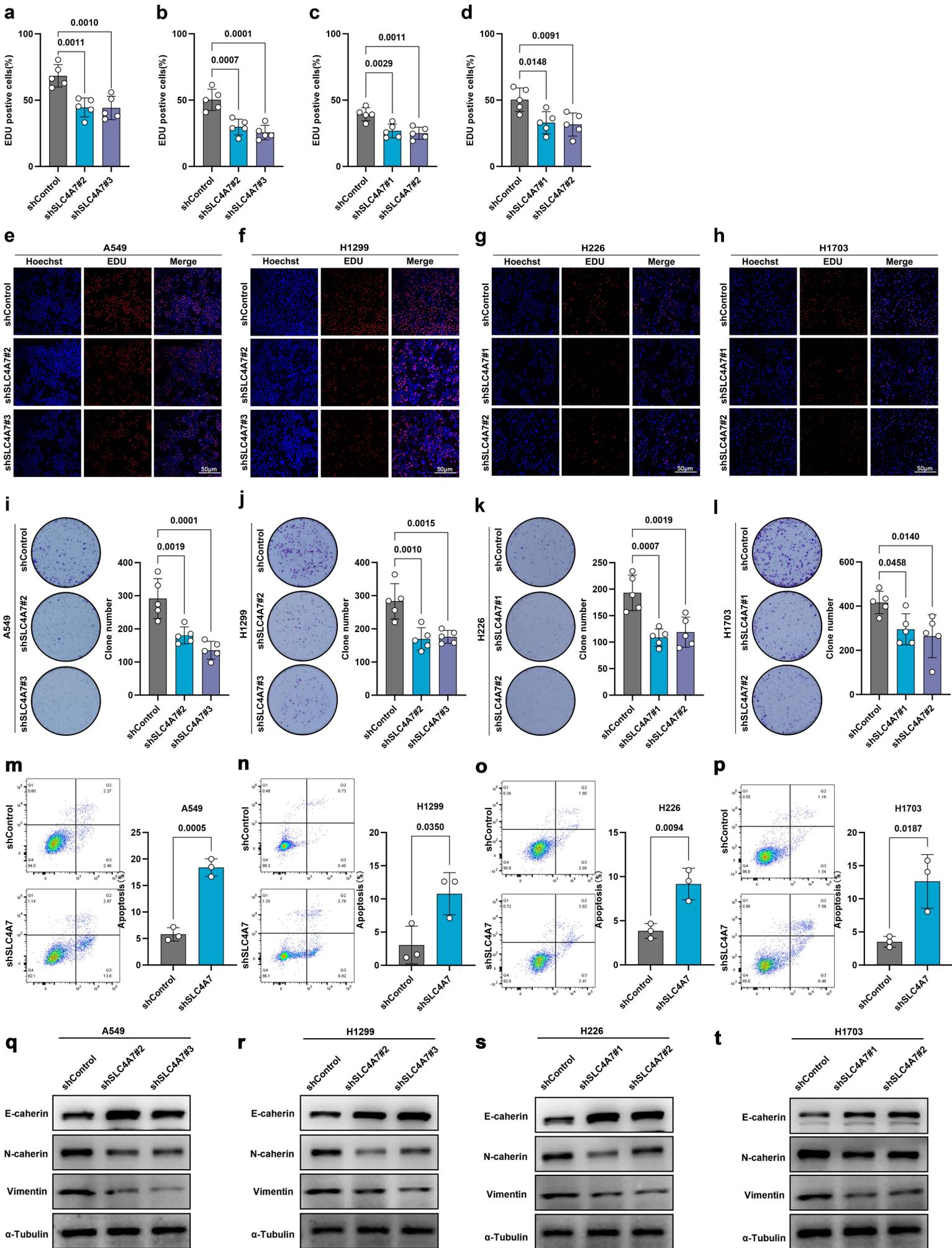
Variable	Overall, N = 90 <sup>1</sup>	High, N = 12 <sup>1</sup>	Low, N = 23 <sup>1</sup>	Middle, N = 55 <sup>1</sup>	p- value <sup>2</sup>
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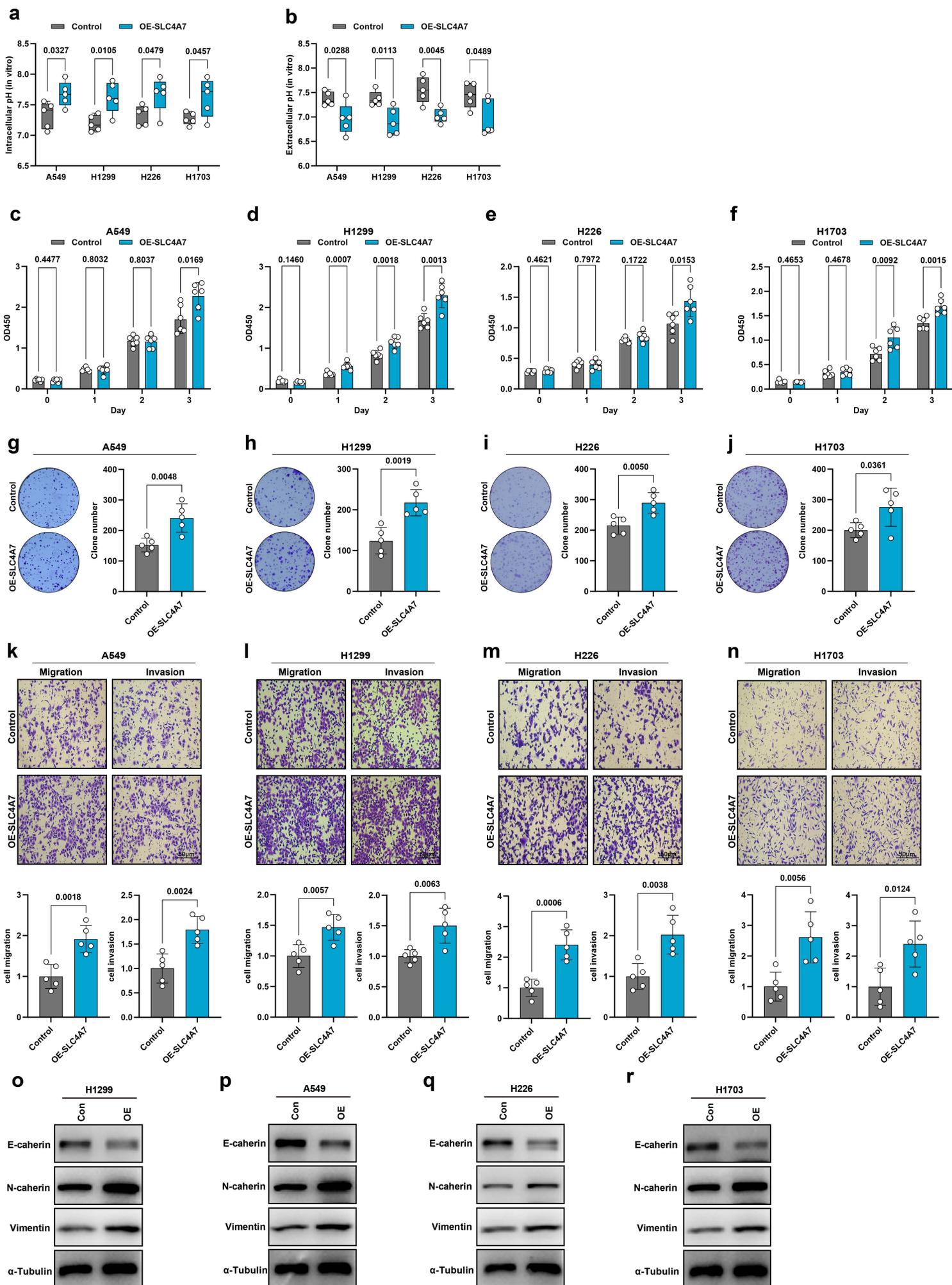
<sup>1</sup>n (%)

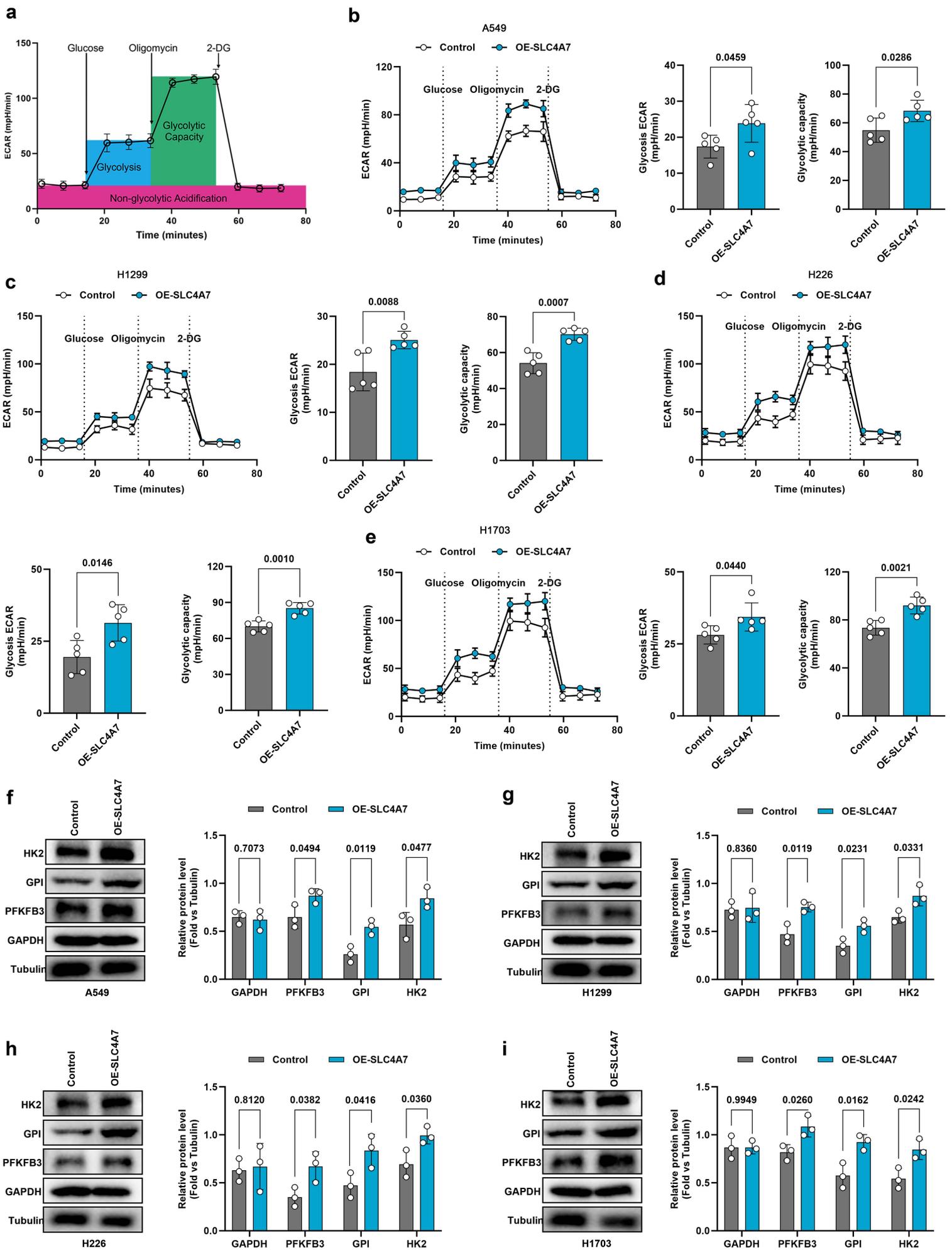
<sup>2</sup>Fisher's exact test; Pearson's Chi-squared test

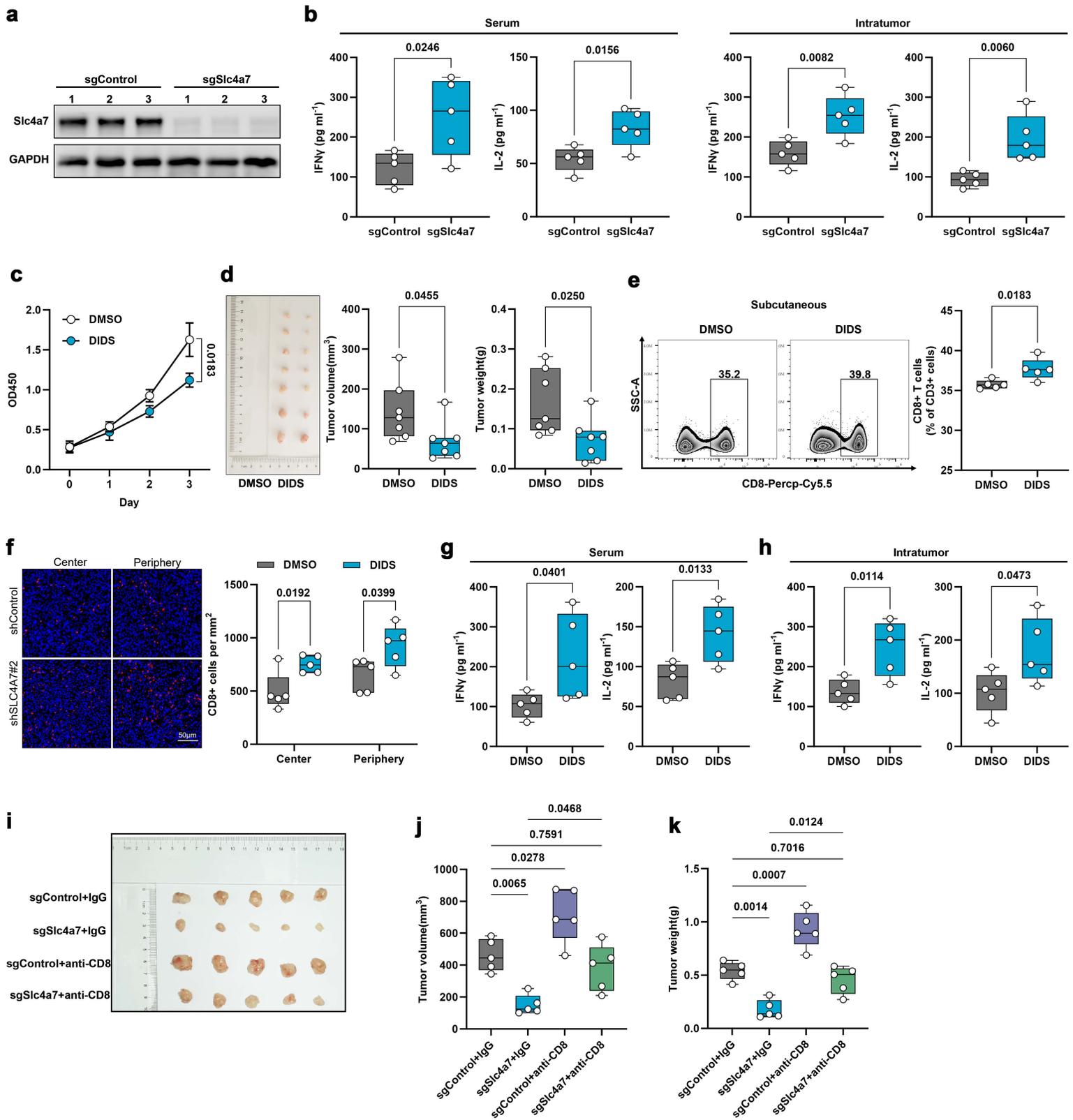


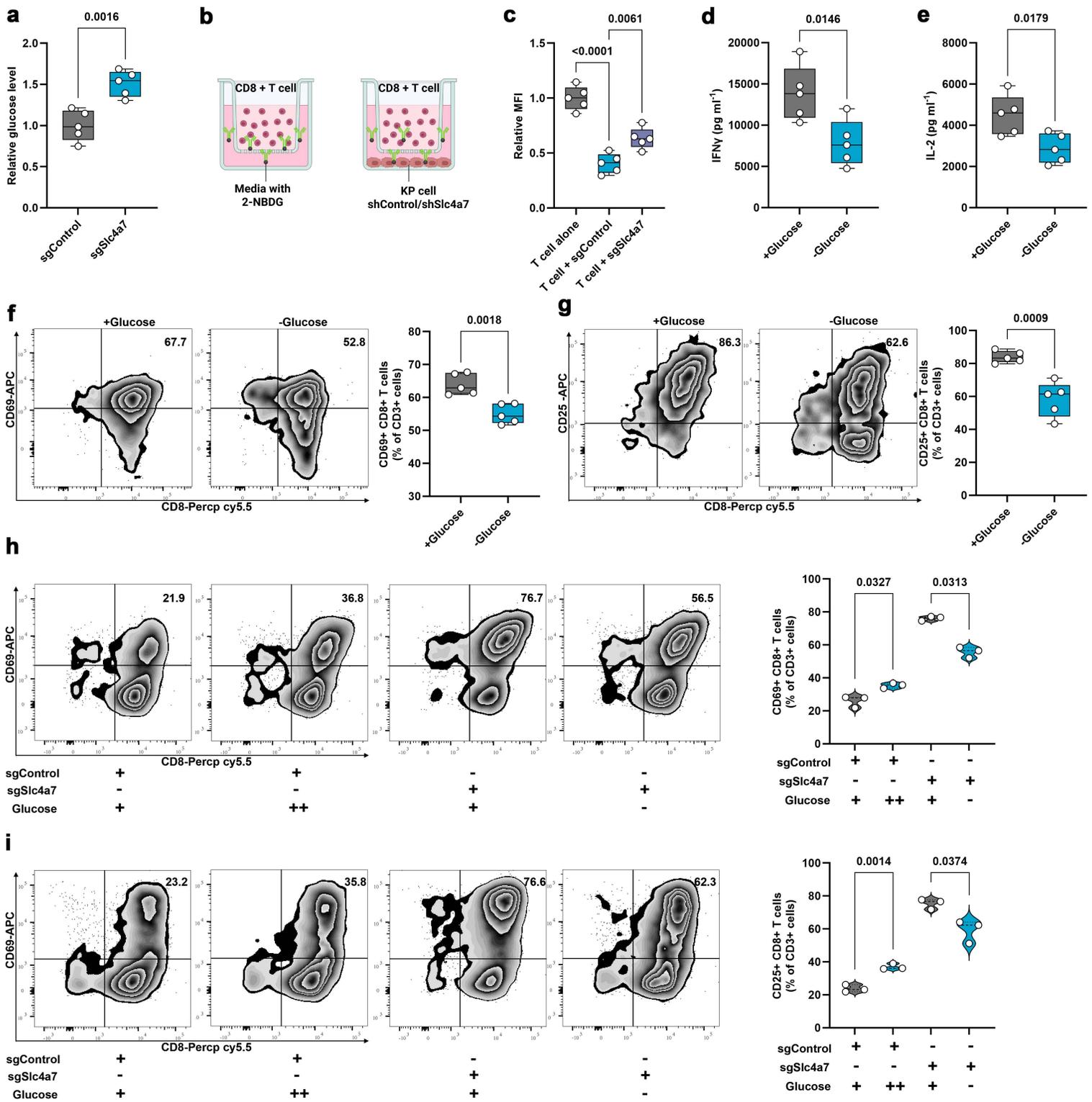






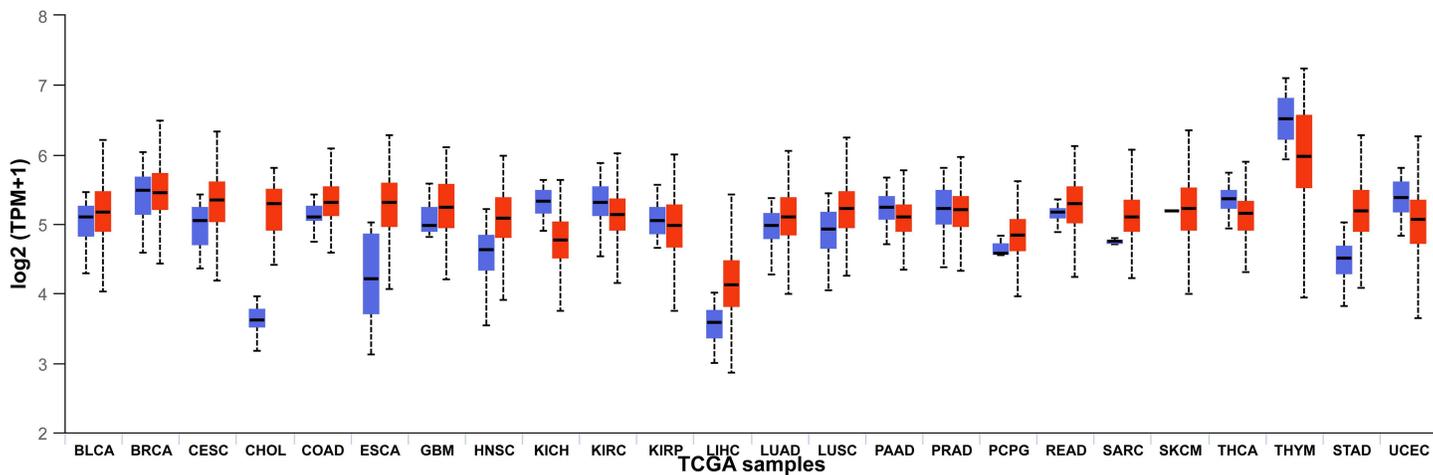
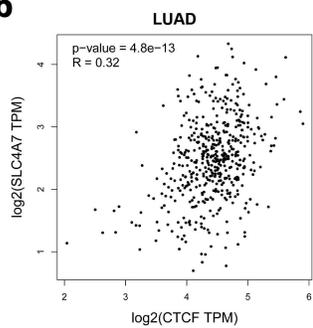
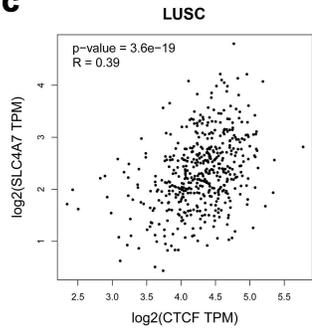
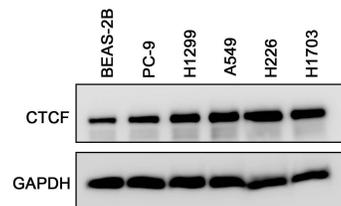
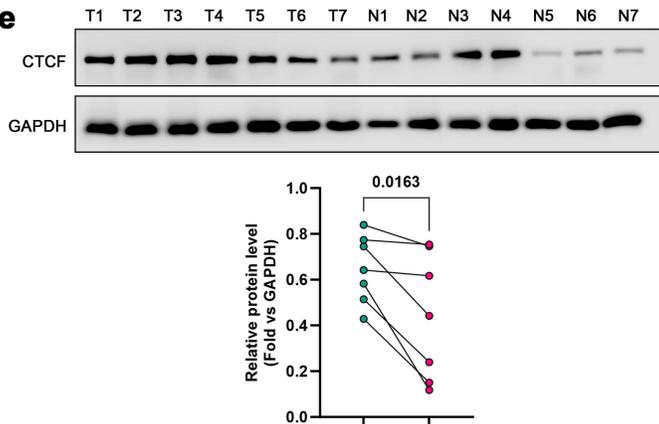
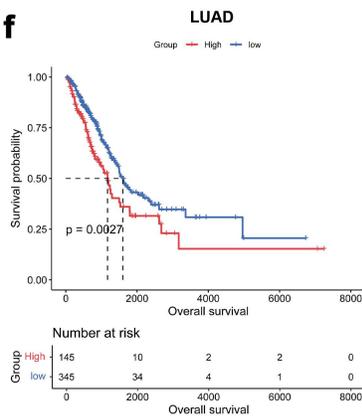






**a**

### Expression of CTCF across TCGA cancers (with tumor and normal samples)

**b****c****d****e****f****g**