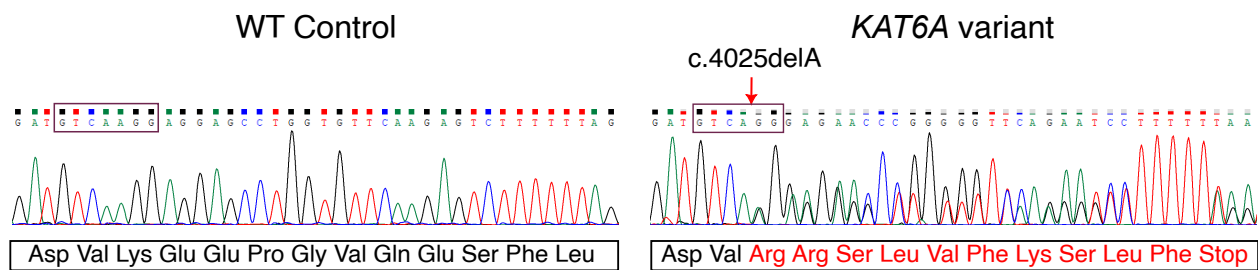


Supplementary Material

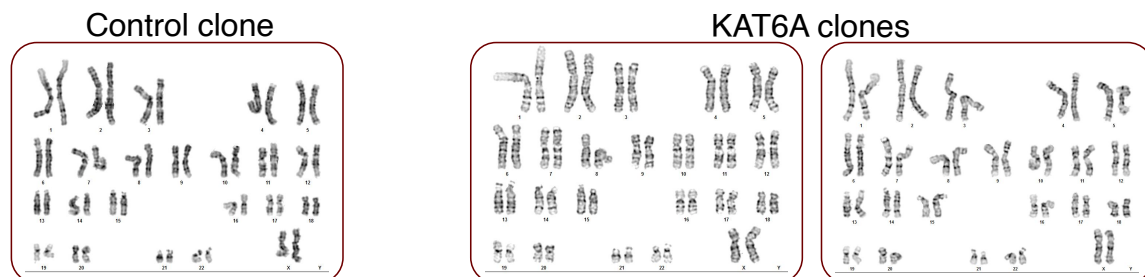
***KAT6A*-Mutant iPSC-Derived Cortical Neurons Display Precocious Maturation, Dysregulated Electrophysiology, and Altered Morphology**

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Wote Amelo Rike², Aviram Shemen², Ritu Nayak², Souvik Chakraborty², Omveer Sharma²,
Tatiana Rabinski^{3,4,5}, Michal Stark¹, Karin Weiss^{6,7}, Gad D. Vatine^{3,4,5}, Shani Stern^{2*},
and Yehuda G. Assaraf^{1*}

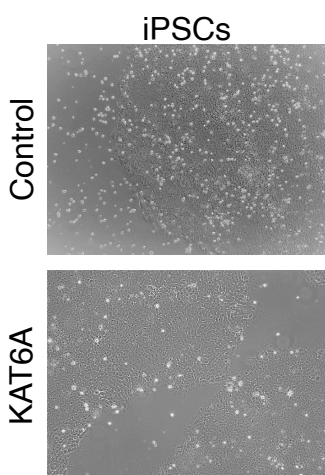
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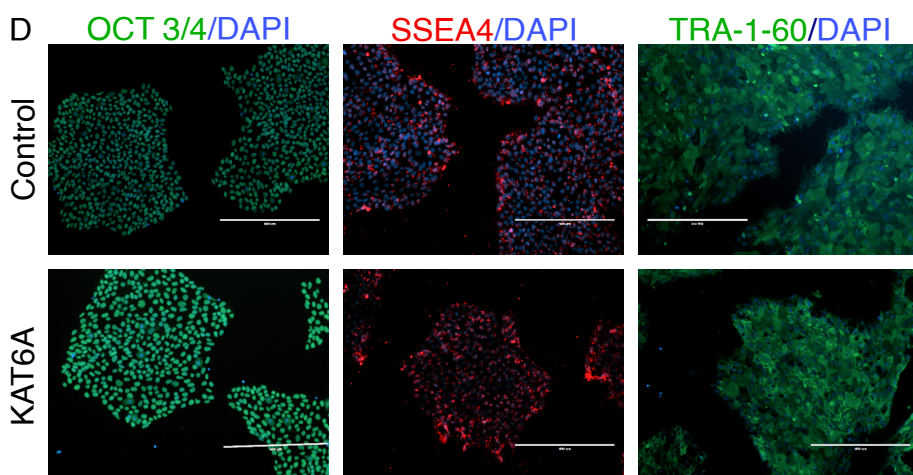
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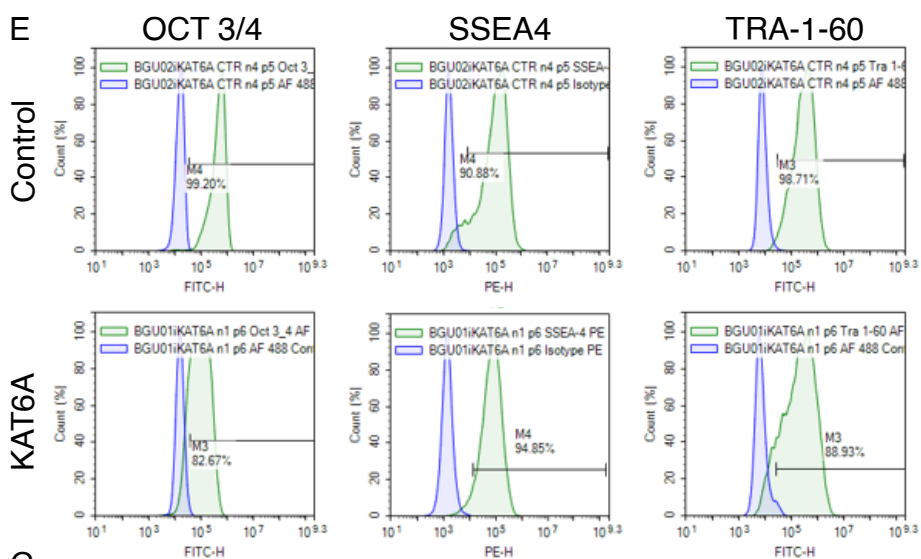
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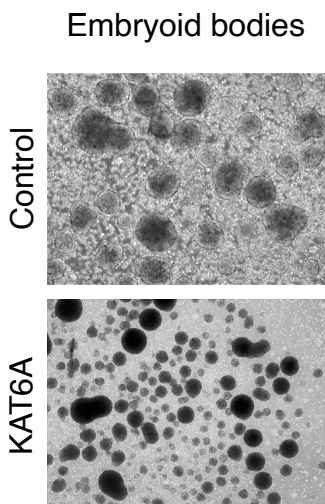
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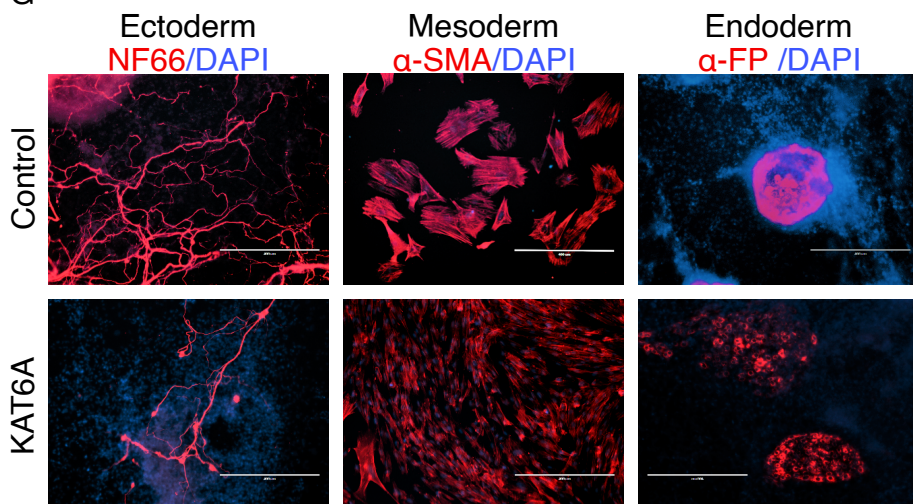


Figure S1. Characterization of iPSC clones derived from a KAT6A syndrome patient and a related control. A) Sanger sequencing of a DNA fragment from exon 17 validated the retention of a heterozygous *KAT6A* c.4025delA pathogenic variant in the patient's iPSC cell line, compared to the sequence of the related unaffected control line. The location of the mutation and the altered amino acid sequence of ten missense mutations followed by a stop codon are indicated (framed by rectangles). B) iPSC clones of the related control and KAT6A syndrome patient maintained a normal 46,XX karyotype and showed no abnormalities. C) Images of iPSC colonies that were generated from dermal fibroblasts derived from the patient and the related unaffected control. (D, E) Pluripotency of the iPSCs was validated by immunofluorescence microscopy analysis (D) (Scale bar= 400 μ m) and flow cytometry analysis (E) of the pluripotency markers OCT3/4, SSEA-4 and TRA-1-60. (F, G) iPSC clones successfully formed EBs (F) and spontaneously differentiated into the three germ layers (G). G) Expression of ectoderm (α -Internexin neurofilament (NF66)), mesoderm (α -SMA) and endoderm (α -FP)) was confirmed by immunofluorescence microscopy analysis.

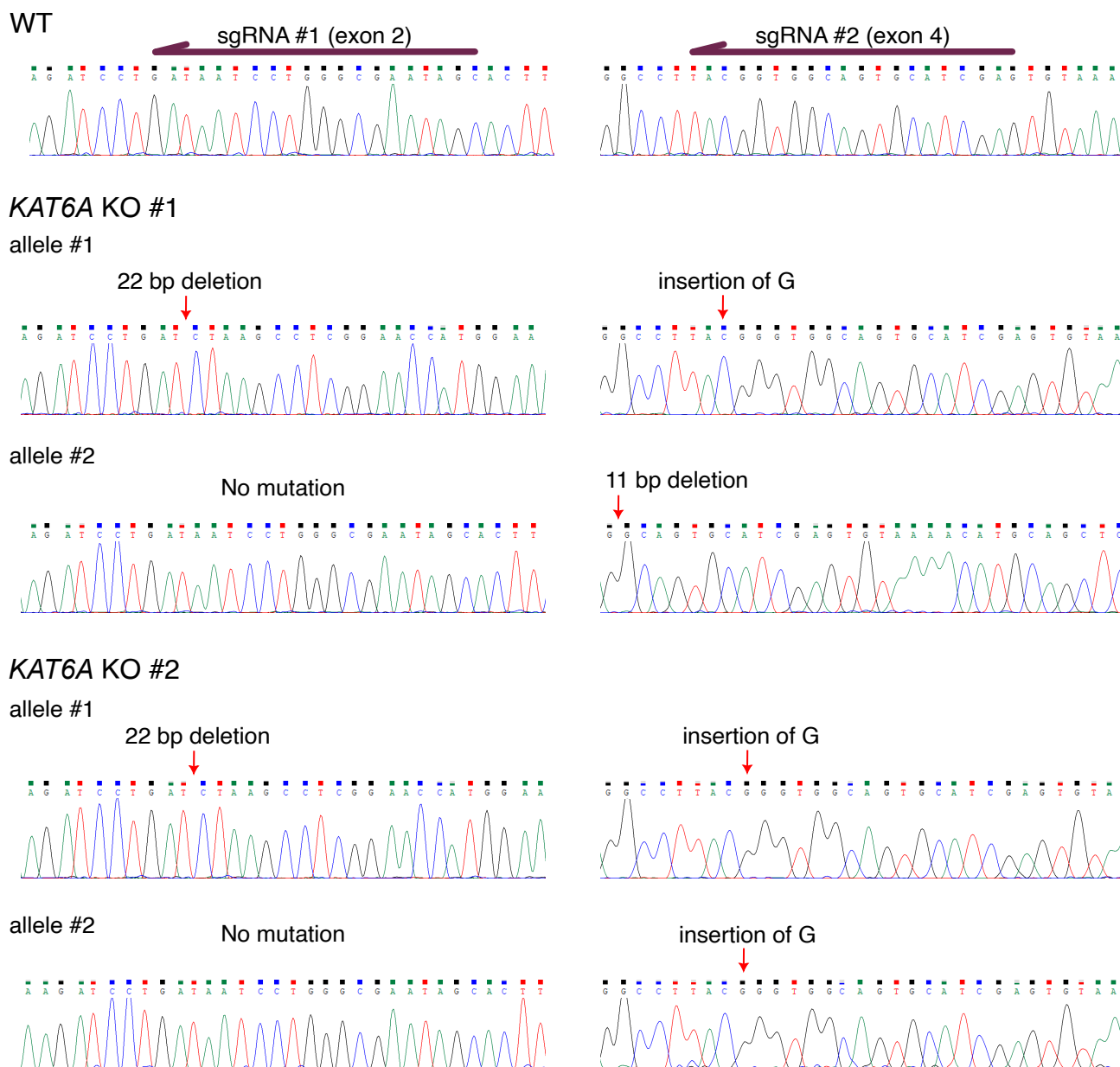


Figure S2. Validation of *KAT6A* KO in SH-SY5Y cells via Sanger DNA sequencing.

Sanger sequencing results of WT control and edited cell lines (*KAT6A* KO clone #1/clone #2). Thirty-one bases of the chromatogram are shown. The sgRNA sequences targeting the *KAT6A* gene are indicated on the WT sequence.

Supplementary Tables S1-3. RNA sequencing results of dysregulated genes and pathways in NPCs (Table S1), cortical neurons at the 1st tp of differentiation (Table S2), and cortical neurons at the 2nd tp of differentiation (Table S3).

Supplementary Table S4. Protein-coding genes that are dysregulated in *KAT6A*-mutant NPCs and cortical neurons, which are also associated with ASD according to the SFARI database.

Supplementary Table S6. A detailed description of all the quantified morphometric parameters .

Supplementary Table S5. List of primers for PCR, sequencing, and RT-PCR.

Primers for <i>KAT6A</i> sequencing	
<i>KAT6A</i> fragment amplification for iPSCs seq:	
<i>KAT6A</i> F	TTGCGAAAACGTGATGTGAAG
<i>KAT6A</i> R	GGGAGTCTTCTTCGTGGTTCGT
<i>KAT6A</i> seq (iPSCs)	CAGAGGAGGAGCAGCAGGAA
<i>KAT6A</i> fragment amplification for SH-SY5Y clones seq:	
<i>KAT6A</i> F	TGGTAAAACTCGCAAACCCG
<i>KAT6A</i> R	GATCTCGACAGGAGCTGCATG
<i>KAT6A</i> seq (SH-SY5Y clones)	TGGTAAAACTCGCAAACCCG
Primers for RT-PCR	
<i>KAT6A</i> F	GCTGAACCAATCCCCATCTGT
<i>KAT6A</i> R	TCTCGACAGGAGCTGCATGTT
<i>KAT6B</i> F	GGATTTGGACGGTTTCTCATTG
<i>KAT6B</i> R	GCCTTGATGCTGATGTGCCT
<i>NTRK2</i> F	TTGAAGCTTATGTGGGACTGAGA
<i>NTRK2</i> R	TCAGACAAGTCAAGGTGACGGA
<i>PSD95</i> F	AAACCAGAAGAGTACAGCCGATT0
<i>PSD95</i> R	GCCTGCCACCACTCCTCAT
<i>SYF</i> F	GAAGGTGCTGCAATGGGTCTT
<i>SYR</i> R	GGTGCAGCCTGAAGGGGTA
<i>ENO2</i> F	GGAGAACAGTGAAGCCTTGGA
<i>ENO2</i> R	CAATCATCCTGGTCAAATGGGT
<i>RSRC1</i> F	TGTTTCAGCAGACATTCAGATCA
<i>RSRC1</i> R	CAGTACTGGGTGGATCAGCA
<i>CRABP2</i> F	CCCTGTAAGAGCCTGGTGAAAT
<i>CRABP2</i> R	GCCGTCATGGTCAGGATCAGT
<i>SHOX2</i> F	CCCGAGTGCAGGTTTGGTT
<i>SHOX2</i> R	AAATGGCATCCTTAAAGCACCTA
<i>EMC7</i> F	CAGACTGCCCTATCCTCTCCA
<i>EMC7</i> R	TCAGGCAACTCATGGTTGGA
<i>GAPDH</i> F	ATGGGGAAGGTGAAGGTCG
<i>GAPDH</i> R	GGGGTCATTGATGGCAACAATA
<i>RPLP0</i> F	GCACCATTGAAATCCTGAGTGA
<i>RPLP0</i> R	ACATTGCGGACACCCTCCA